

Graph Theoretical Analysis, Insilico Modeling and Formulation of Pyrimidine Nanoparticles as p38 α MAP Kinases inhibitors: A Quantitative Proteomics Approach

Authors

Panneerselvam Theivendren¹, Selvaraj Kunjiappan², Saravanan Govindraj³, Jaikanth Chandrasekarn⁴, Parasuraman Pavada⁵, Ganesan Rajalekshmi Saraswathy⁶, Indhumathy Murugan⁷

Affiliations

- 1 Department of Pharmaceutical Chemistry, Karavali College of Pharmacy, Vamanjoor, Mangalore, Karnataka, India
- 2 Sir C.V. Raman Krishnan International Research Centre, Kalasalingam University, Krishnankoil, Tamilnadu, India
- 3 Department of Pharmaceutical Chemistry, MNR College of Pharmacy, Fasalwadi, Sangareddy, Telangana, India
- 4 Department of Pharmacology, Karavali College of Pharmacy, Vamanjoor, Mangalore, Karnataka, India
- 5 Department of Pharmaceutical Chemistry, Faculty of Pharmacy, M S Ramaiah University of Applied Sciences, M S R Nagar, Bengaluru, Karnataka, India
- 6 Pharmacological Modelling and Simulation Centre, Faculty of Pharmacy, M S Ramaiah University of Applied Sciences, M S R Nagar, Bengaluru, Karnataka, India
- 7 Department of Biotechnology, P.S.R Engineering College, Sevalpatti, Sivakasi, Tamilnadu, India

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Correspondence

Dr. T. Panneerselvam

Professor, Department of Pharmaceutical Chemistry

Karavali College of Pharmacy


Vamanjoor, Mangalore- 575028

Karnataka

India

Tel.: +91/759/8224 818

tpsphc@gmail.com

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ABSTRACT

In this study, the optimized 4-(4-hydroxybenzyl)-2-amino-6-hydroxypyrimidine-5-carboxamide derivative was formulated as nanoparticles to evaluate for their anticancer activity. The response surface methodology (RSM) was performed with utilization of Box-Behnken statistical design (BBD) to optimize the experimental conditions for identification of significant synthetic methodology. To explore the stability of the derivative was done by density functional theory (DFT). Graph theoretical analysis was introduced to identify the drug target p38 α MAP Kinases and then insilico modeling was performed to provide straightforward information for further structural optimization. The experimental results under optimal experimental conditions obtained 74.55–76 % yield of 4-(4-hydroxybenzyl)-2-amino-6-hydroxypyrimidine-5-carboxamide, 127 $^{\circ}$ C melting point and Rf value 0.59 were well matched with the predicted results and this was gaining 95 % of confidence level and suitability of RSM. The spectral data were reliable with the assigned structures of synthetic yields. The formulated nanoparticles were exhibited a good anticancer activity against used cancer cell line MCF7. Amusingly the observed docking scores and in-vitro anticancer activity was proving the compound significance and potential as a potent p38 α inhibitor. Further, we have elucidated the mechanism of action at its functional level using label-free quantitative proteomics. Interestingly the observed results were indicating that the derived proteomics data involving in the alteration process in cancer-related regulatory pathways.

Introduction

Despite the fact that, the choice of easily reached and pharmacophoric heterocycles is fairly limited, the development of novel, fast, and healthy routes toward paying attention of synthesis of heterocycles are great significance. In this connection the literature review reveals that, the pyrimidines and fused pyrimidines are important classes of heterocyclic compounds. They are widely used as key input to the structure construction for pharmaceutical agents and it's known to exhibit pharmacological activities like analgesic, anti-inflammatory [1, 2], anticancer [3–6], antifolates [7], antihistaminic [8], antibacterial [9], antiviral [10], hypoglycaemic [11], anticonvulsive [12], PDE4 inhibitor [13]. In addition the heterocyclic pyrimidine ring system is an extensively spread in various structural scaffolds that are present in a number of pharmaceuticals as well natural products. The unique structural collection and the highly marked pharmacological activities are displayed by this class of pyrimidine compounds have made them eye-catching synthetic targets. Led by above facts on pyrimidine chemistry, the compound 4-(4-hydroxy benzyl)-2-amino-6-hydroxypyrimidine-5-carboxamide was chosen to optimization to get significant synthetic route. The combinatorial chemistry is regularly applied into find new biologically active scaffold. In this point of view, multicomponent reactions (MCRs) are very attractive and commanding tool in the current drug discovery course that to lead findings and lead optimization [14–17]. The present study was aiming that to developing and validating 4-synthesis of (4-hydroxy benzyl)-2-amino-6-hydroxypyrimidine-5-carboxamide using response surface methodology because no work has been done so far in this particular area. The RSM is a statistical tools [18, 19] used to analyze quantitative data through simultaneously solve multivariate equations [20]. In this connection to perform this, there are five different parameters such as 4-hydroxy benzaldehyde (X_1), ethyl cyanoacetate (X_2), guanidine hydrochloride (X_3), temperature (X_4) and rotation per minute (X_5) were chosen for investigation. The study of optimization with different independent and dependent levels was employed to look at the optimum conditions with high opinion to synthetic yields of (4-hydroxy benzyl)-2-amino-6-hydroxypyrimidine-5-carboxamide. The synthesized pyrimidine derivative was formulated as nanoparticles to enhance its therapeutic action. Mitogen Activated Protein Kinases (MAP Kinases) are very important signalling path and it involves in cell proliferation and cell death and typically it was linked with cancer cells particularly breast cancer [21–23]. The introduced graph theoretical analysis was producing information that the importance MAP Kinases and its interaction that to improve insilico modeling for identification of target especially in breast cancer agents. The optimization of pyrimidine derivatives and their structures has available in commercial market, like Uramustine, Tegafur, Floxuridine, Fluorouracil, Cytarabine, Methotrexate etc., Herein, we report the stability of designed structure 4-(4-hydroxybenzyl)-2-amino-6-hydroxypyrimidine-5-carboxamide by using DFT and its insilico screening was performed based on the results obtained from graph theoretical analysis, the identified active sites of available crystal structure of MAP Kinases (2FST) was used for docking studies. In addition to increasing our awareness of the significance of active molecule identification, the introduced graph theoretical analysis, DFT label free quantitative proteomics and approach is attractive and it is meaningful to formulate pyrimidine nanoparticles and screen them for their anticancer activity.

Materials and Methods

Materials

The graph theoretical analysis was performed by KEGG database and Cytoscape software 3.3.0. The docking study was performed by Sybyl-x version 2.0, Tripos International, St. Louis, MO, USA, 2012.” The density functional theory was performed by Becke's 3-parameter exchange functional combined with Lee-Yang-Parr correlation potential (B3LYP). The molecular docking was performed with Surflex-Dock program that is interfaced with Sybyl-X 2.0. Label free quantitative proteomics was performed as described in our early publications. Briefly, agilent 1260 infinity HPLC-Chip/MS system was used. The chemicals used for synthesis were purchased as analytical grade procured from Biotium, Inc, Aldrich Chemical Co. and Merck Chemical Co. The melting points were determined in open capillary tubes and are uncorrected. IR spectra were recorded with KBr pellets (FT-IR spectrometer MB 104 ABB Limited, Kalasalingam University, Krishnankoil, Tamilnadu). ^1H NMR spectra (Bruker 300 NMR spectrometer, Punjab University, Chandigarh) were recorded with TMS as internal reference. Mass spectral data were recorded with a quadrupole mass spectrometer (Shimadzu GC MS QP 5000, Punjab University, Chandigarh), and microanalyses were performed using a vario EL V300 elemental analyzer (Analytensysteme GmbH, Kalasalingam University, India). The purity of the compounds was checked by TLC on pre-coated SiO_2 gel (HF_{254} , 200 mesh) aluminum plates (E. Merck). Evidence of structure was achieved by IR, ^1H -NMR, mass spectral data, and elemental analyses.

Methods

Graph theoretical analysis

The graph theoretical analysis was performed by using KEGG database and Cytoscape software [24]. In this study, the preparation and processing of KEGG pathway data (hsa 04010) used with different centrality measures to identify key central node of network. The overview of MAP Kinases signalling pathway from KEGG database is shown in ► **Fig. 1**. Network components were visualized and analyzed using Cytoscape 3.3.0. Nodes are genes, compounds and edges are interaction between the nodes. The network contains 129 nodes with 177 edges.

Insilico modeling

Molecular docking was used to elucidate the binding mode of the compounds to provide straight forward information for further structural optimization. The crystal structure of mitogen activated protein kinase p38alpha (D176A + F327L) activating mutant (PDB ID: 2FST) was extracted from the Brookhaven Protein Database (PDB <http://www.rcsb.org/pdb>). The proteins were prepared for docking by adding polar hydrogen atom with Gasteiger-Huckel charges and water molecules were removed. The 3D sketch up of the structure was implemented by SYBYL program (Tripos Inc., St. Louis, USA) and its energy-minimized conformation was obtained with the help of the Tripos force field using Gasteiger-Huckel charges and molecular docking was performed with Surflex-Dock [25] program that is interfaced with Sybyl-X 2.0 and other miscellaneous parameters were assigned with the default values given by the software.

DFT

The synthesized compound was studied for their stability by DFT with standard drug Uramustine. The DFT was performed through B3LYP [26] and basis set of 6-311++G(d,p). The above parameters have been used in the purpose of optimization and stability studies of compound and Uramustine. The optimized structures of selected compound and Uramustine were shown in ►Fig. 2. In this experiment to understand the stability, highest occupied molecular orbital (HOMO) and lowest unoccupied molecular orbital (LUMO) gap, DFT reactivity parameter have been calculated and further chemical hardness also been calculated using the following formula:

$$\eta = \frac{(I - A)}{2}$$

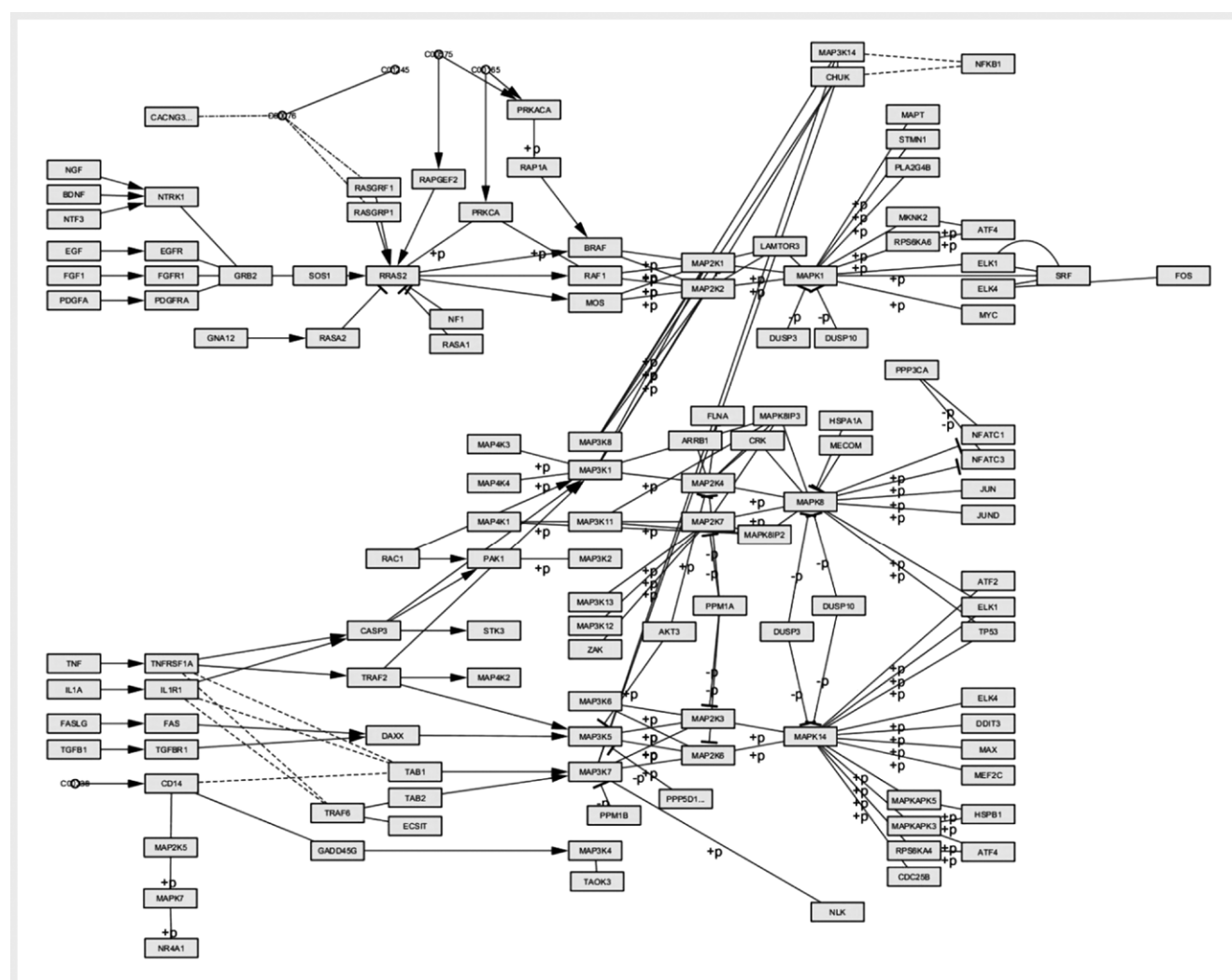
Where I = ionization potential, and A = electron affinity of a molecular system. Based on Koopman's theorem [27] HOMO was related to $I = -E_{\text{HOMO}}$ and LUMO was related to $A = -E_{\text{LUMO}}$. All the calculations part was performed by using NWChem 6.0 program [28].

Optimization

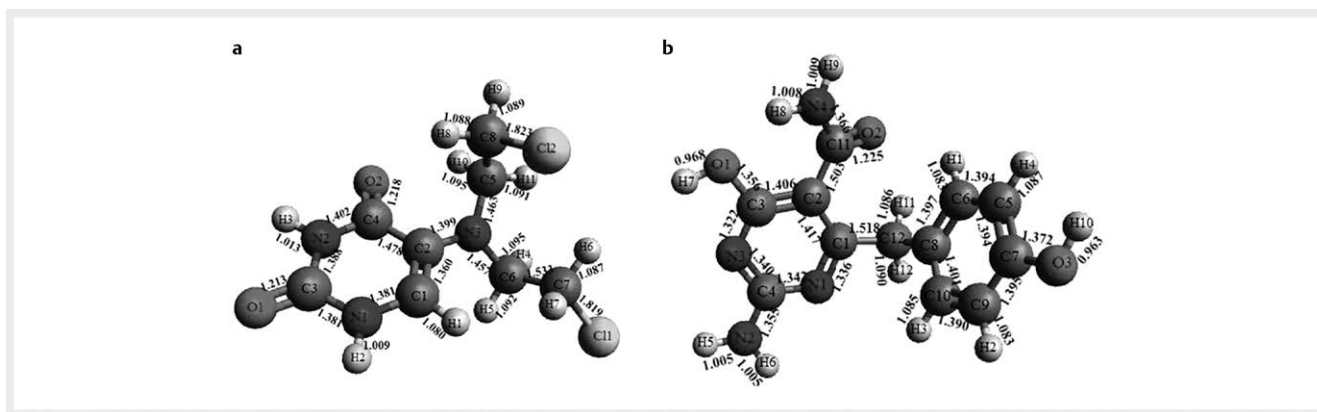
The most effective key parameters on 4-hydroxy benzaldehyde ($X_1 = 0.1-1$ M), ethyl cyanoacetate ($X_2 = 0.1-1$ M), guanidine hydrochloride ($X_3 = 0.1-1$ M) including temperature and rotation per minute were chosen for identification of significant synthetic methodology of 4-(4-hydroxybenzyl)-2-amino-6-hydroxypyrimidine-5-carboxamide. In order to optimize the values of these parameters and reach the best response, BBSD was applied. Three levels three factors BBSD was applied to examine the optimum combination of effective synthetic methodology. The coded and real levels of the independent variables in the Box-Behnken statistical design matrix were listed in ►Table 1. A total of 46 investigational runs with eight factorial points, six axial points and six replicates of the central point's were carried out according to the BBD experimental design and low, middle, and high levels of the coded values were designated for the variables as 1, 0, and -1, respectively.

Synthetic Procedure

The synthetic strategy to prepare the target compounds is depicted in earlier reported ►Fig. 3 [29]. The equimolar mixture of 4-hy-



► Fig. 1 The signalling pathway of MAP Kinases



► **Fig. 2** The optimized DFT structures of compound and Uramustine

► **Table 1** Experimental parameters and range of coded and actual parameters of Box-Behnken statistical design (BBS).D).

| Independent variables | Symbols (x_j) | Factor levels | | | |
|-------------------------|-------------------|---------------|------------|--------|-------------|
| | | Units | Low Actual | Medium | High Actual |
| 4-hydroxy benzaldehyde | (X_1) | M | 0.01 | 0.055 | 0.1 |
| ethyl cyanoacetate | (X_2) | M | 0.01 | 0.055 | 0.1 |
| guanidine hydrochloride | (X_3) | M | 0.01 | 0.055 | 0.1 |
| Temperature | (X_4) | C | 30 | 40 | 50 |
| RPM | (X_5) | RPM | 100 | 200 | 300 |

droxy benzaldehyde, (0.01–1.0 mol), ethyl cyanoacetate (0.01–1.0 mol) and NaOH (0.4 g in 5 ml water) in 25 ml ethanol was stirred mechanically for at least 10 min, then guanidine hydrochloride (0.01–1.0 mol) was added to the above reaction mixture and reaction mixture was stirred with 100–300 rpm using temperature 30–50°C till completion of reaction. Followed by the above reaction mixture was mixed with 1 mmol of TFA in presence of H_2SO_4 in continuous stirring for 4 h at room temperature. After this it was poured into ice cooled water to get the desired product and to end with recrystallization using ethanol to get pure product 4-(4-hydroxy benzyl)-2-amino-6-hydroxypyrimidine-5-carboxamide.

Formulation of nanoparticles

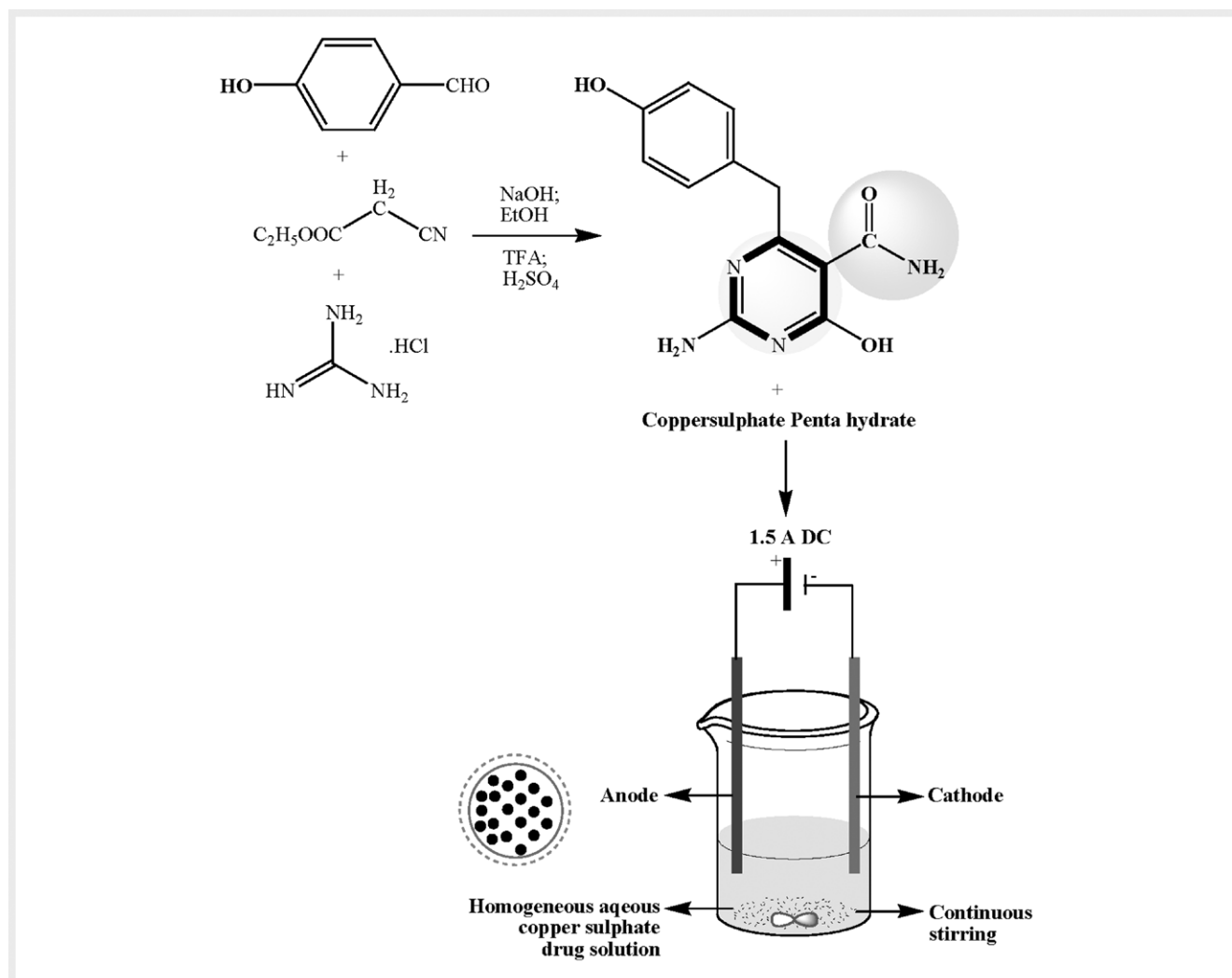
The formulation of metal-drug nanoparticles was produced by electrolytic cathode reduction of copper and the methodology was followed by addition of 0.5 % w/v of 4-(4-hydroxy benzyl)-2-amino-6-hydroxypyrimidine-5-carboxamide and 5 % w/v of copper sulphate penta hydrate into a 500 mL beaker with this 100 mL water was added to make a homogenous aqueous copper sulphate-drug solution. The fresh anode and cathode electrodes of copper plates were placed in this solution and 1.5 A DC a constant current was passed into this solution. At the end of the electrolyzing process a layer of copper-drug nanoparticles deposition was observed on the cathode surface. The copper-drug nanoparticles was obtained through removal of observed layer and it was washed with solution of 100 mL water and 60 mL acetone to remove residual of starting materials. The formulated nanoparticles was characterized by SEM analysis and X-ray diffraction to verify the formation of copper-drug nanoparticles.

Anticancer activity

The potential cell proliferation activity of optimally prepared 4-(4-hydroxybenzyl)-2-amino-6-hydroxypyrimidine-5-carboxamide was examined by 3-(4, 5-dimethylthiazol-2-yl)-2, 5-diphenyltetrazolium bromide (MTT) assay. Briefly, Breast cancer cell lines (MCF7 cell) were seeded in a 96-multiwell plate (1×10^5 cells/well) for 24 h before treatment with 4-(4-hydroxybenzyl)-2-amino-6-hydroxypyrimidine-5-carboxamide, allowing for cell add-on to the plate wall. Different concentrations of 4-(4-hydroxybenzyl)-2-amino-6-hydroxypyrimidine-5-carboxamide compound (5–25 $\mu\text{g}/\text{mL}$) were added to cell monolayer and the respective amount of DMSO in PBS buffer was used as control. The medium was removed after 24 h of treatment and cells were washed with phosphate-buffered saline (PBS, 0.01M, pH 7.4). This was followed by addition of 200 μL (5 mg/mL) of MTT prepared in serum free medium solution to each well and incubated for 4h at 37 °C in a 5% CO_2 . Thus, the obtained cells were fixed, washed and stained with MTT stain. Acetic acid was used to remove excess stain while Tris EDTA buffer was used to remove any attached cells. The color intensity was measured in a microplate reader at 564 nm. The absorption ratio of treated cells to absorption of non treated cells expressed in percentage resulted in percentage of death cells. Thus, every treatment condition was repeated in triplicate and IC_{50} of 4-(4-hydroxybenzyl)-2-amino-6-hydroxypyrimidine-5-carboxamide compound obtained were used for treatment for all further studies.

HPLC-Chip-MS

The charged peptides from HPLC-Chip system were infused into mass-spectrometer for detection. The following HPLC-Chip-MS conditions were used for acquiring the MS and MS/MS spectrum of the peptides. Chip ID: G4240-62030 Chip Name: High Performance Chip,



► **Fig. 3** Synthetic & Formulation protocols

360 nanoliter enrichment column, 150 mm X 75 μ m separation, column Solvent A: 0.1% Formic Acid, Solvent B: 90% ACN/10% (0.1% Formic Acid), Flow Rate: 0.3 μ l/min Run Time: 120 minutes, Sample Volume: 5 μ l, MS Scan Range: 275 to 1700 m/z, MS Scan Rate: 8 spectra/sec, MS/MS Scan Rate: 3 spectra/sec Ion Polarity: Positive Ions Fragmentor Voltage: 170 V, Skimmer Voltage: 65 V, Octopole RF Voltage: 750 V Gas, Temperature: 250 $^{\circ}$ C & Drying Gas: 5 L/min.

Bio-informatics analysis

Protein identification was performed with the following criteria: (a) Trypsin digested peptides with 4 missed cleavages allowed, (b) peptide tolerance < 50 ppm, (c) > 2 unique peptides, (d) FDR < 5%. Fasta files for human proteins were downloaded from the uniprot database (Jaikanth et al., 2017). For the analysis, proteins identified in at least 2 out of 3 replicates in each group were considered. Sum of unique peptide intensity was used for semi-quantitative analysis. Ratio was calculated for proteins identified in both the treated and control groups. Ratio of > 1.5 was considered as “up-regulated” and ratio < 0.5 was considered as down-regulated. The signaling pathway, Gene ontology and interaction network were analyzed by using open source of STRING database.

Results

Graph theoretical analysis

The network was intended to recognize key node of centrality measures like Degree, Stress, Betweenness, Radiality, Closeness, Eccentricity and Eigenvector and it was achieved by using Cytoscape software. The values maximum, minimum and mean were calculated for each and every protein and mean value is in use as a threshold value. In all the parameters a node of exceeds threshold values are measured as key node in network and the results were represented in ► **Table 2**.

Insilico Modeling

The compound and standard Uramustine were docked into the active site as shown in ► **Fig. 4**. The predicted binding energies of the compounds are listed in ► **Table 3**. The figure represents the interaction of Uramustine with the active site of the enzyme and observed results were showed one binding interaction at active site of the enzyme PDB ID: 2FST that is hydrogen atom of 2 $^{\circ}$ amine with oxygen of hydroxyl group of amino acid residue GLU71 (N-H -----

► **Table 2** Signalling pathway of p38 α MAP Kinases and values of Centrality analysis.

| | Degree | Betweenness | Closeness | Eccentricity | EigenVector | Radiality | Stress |
|---|--------|-------------|------------|--------------|-------------|-----------|---------|
| Maximum value | 15 | 7868.777333 | 0.2788671 | 14 | 0.424226773 | 10.96875 | 145938 |
| Mean value | 2.7 | 593.5 | 0.1801 | 10.53 | 0.0551 | 8.86 | 12789.7 |
| Minimum Value | 1 | 0 | 0.11337467 | 7 | 6.38E-05 | 5.953125 | 0 |
| p38α MAP Kinases | 15 | 2860.227536 | 0.19423369 | 11 | 0.234960097 | 9.5078125 | 111398 |



► **Fig. 4** The molecular interactions of compound with p38 α (2FST)

► **Table 3** Surflex Docking score (kcal/mol) of the molecules.

| Compounds | C Score ^a | Crash Score ^b | Polar Score ^c | D Score ^d | PMF Score ^e | G Score ^f | Chem Score ^g |
|----------------------|----------------------|--------------------------|--------------------------|----------------------|------------------------|----------------------|-------------------------|
| Uramustine | 3.38 | -0.97 | 1.28 | -81.418 | -41.487 | 25.558 | -12.468 |
| Synthesized Compound | 3.35 | -2.30 | 2.44 | -109.750 | -48.087 | -206.056 | -21.777 |

^a**C Score** (Consensus Score) integrates a number of popular scoring functions for ranking the affinity of ligands bound to the active site of a receptor and reports the output of total score; ^b**Crash-score** revealing the inappropriate penetration into the binding site. Crash scores close to 0 are favorable. Negative numbers indicate penetration; ^c**Polar-score** indicating the contribution of the polar interactions to the total score. The polar score may be useful for excluding docking results that make no hydrogen bonds; ^d**D-score** for charge and van der Waals interactions between the protein and the ligand; ^e**PMF-score** indicating the Helmholtz free energies of interactions for protein-ligand atom pairs (Potential of Mean Force, PMF); ^f**G-score** showing hydrogen bonding, complex (ligand-protein), and internal (ligand-ligand) energies; ^g**Chem-score** points for H-bonding, lipophilic contact, and rotational entropy, along with an intercept term.

► **Table 4** The HOMO-LUMO gap and Chemical hardness values corresponds to the optimized structures of Uramustine and Synthesized drug compound at B3LYP/6-311++G** level of theory (in eV).

| Compound | HOMO-LUMO gap | Chemical hardness |
|------------------|---------------|-------------------|
| Uramustine | 4.29 | 2.15 |
| Synthesized drug | 4.73 | 2.37 |

OH-GLU71, 2.03 Å). As depicted in figure the synthesized compound makes seven hydrogen bonding interactions at active site of the enzyme (PDB ID 2FST). Among them two interactions are came from the amino group, i. e., hydrogen atom interacts with oxygen of GLU71 and ASP168 (N-----H- GLU71 and H-ASP168, 2.73 Å and 1.80 Å), the CONH₂ makes three hydrogen bonding interactions with carbonyl group of LEU104, and ALA51 (H-----O=C-LEU104, O-LEU104 and O-ALA51, 2.25 Å, 2.38 Å and 1.88 Å). The hydroxyl group present on the phenyl ring makes two interactions with HIS107 and MET109 (O-H-----O=C-HIS107, 1.96 Å; H-O-----H-MET109, 2.07 Å). MAP kinase is one of the major protein kinases, catalyzing phosphorylation of a number of substrates and thereby transducing signals from the cell membrane to the interior of the cell.

DFT

The HOMO-LUMO gap values of Uramustine showed 4.29 eV and interestingly synthesized compound was showed 4.73 eV. The ob-

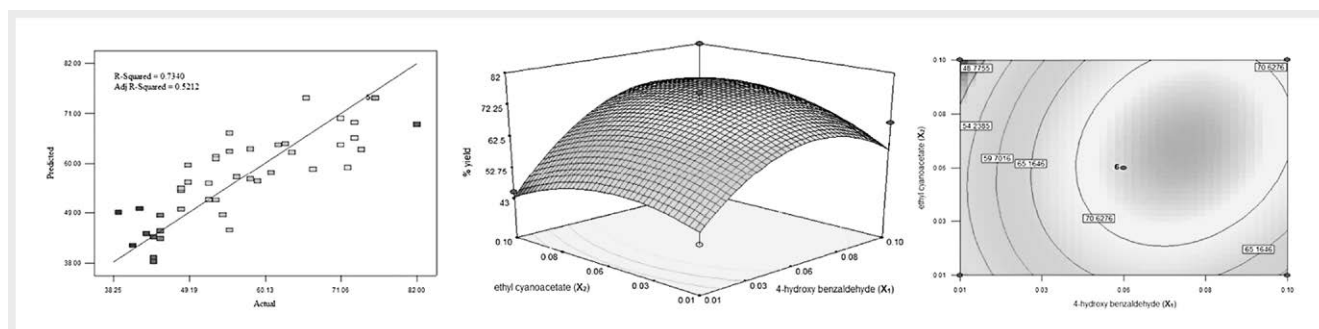
served gap values were indicates that the synthesized compound was more stable than the Uramustine. The bond length values and chemical hardness are listed in ► **Table 4**. In addition the calculated chemical hardness value of synthesized compound was 2.37 eV and it was higher than the Uramustine chemical hardness value 2.15 eV. Based on the maximum hardness principle [30] the consequence of higher chemical hardness value is a clear indication of maximum stability of the structure.

Optimization

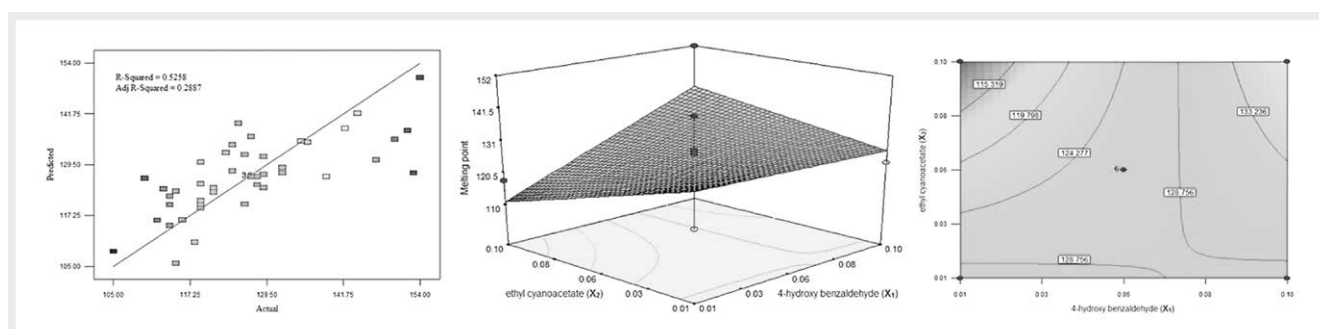
Experimental design, 46 sets of experiments has been conducted by using BBSD and the experiments were performed within the parameters and their range, predicted results experimental results were presented in ► **Fig. 5-7**.

Chemistry

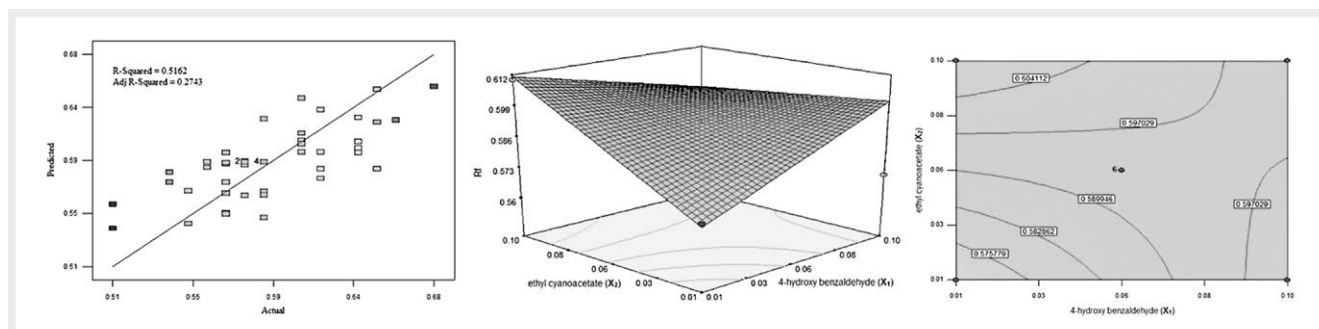
IR: 3368 (OH), 3312 (NH), 3044 (Ar-CH), 2922 (CH₂-CH), 1728 (C=O), 1643 (C=N), 1631 (C=C); ¹H NMR (300 MHz, DMSO-d₆, δ ppm): 9.52 (s, 1H, OH), 8.69 (s, 4H, NH₂), 6.72-7.23 (m, 4H, Ar-H), 3.60 (s, 2H, CH₂); **MS (EI) m/z** 260 [M⁺]; **Elemental analysis:** calculated for C₁₂H₁₂N₄O₃: C, 55.38; H, 4.65; N, 21.53 found: C, 55.36; H, 4.66; N, 21.54.



► Fig. 5 RSM's R2 value and significant effect of percentage yield



► Fig. 6 RSM's R2 value and significant effect of Melting point



► Fig. 7 RSM's R2 value and significant effect of Rf value

Discussion

Graph theoretical analysis

The obtained outcome result from graph theoretical analysis was evidently highlights that the p38 α has crossed all parameters of threshold value. Therefore, p38 α is recognized as a significant protein on the whole network and it will be an ideal target to prevent the development of breast cancer cells.

Insilico Modeling

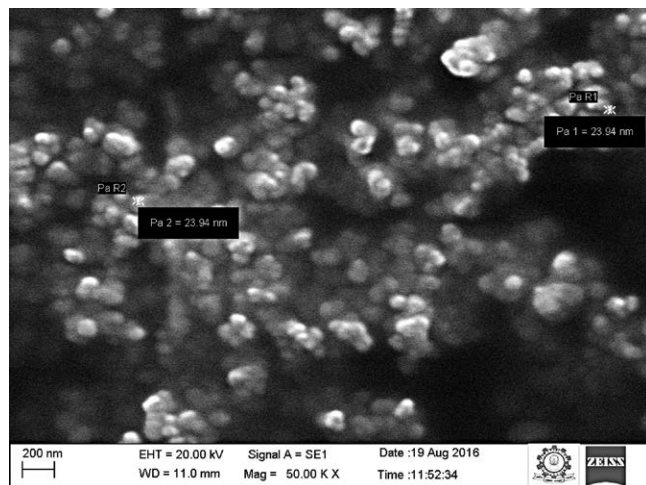
The observed in silico results were showing that the compound interacts with Glu71 which is a conserved residue and this residue lies within the α C helix of ERK1/2 which is mainly it is required for salt-bridging with β 3-lysine and the salt-bridging is essential for kinase to be active [31]. Since the designed compound interacts strongly with

Glu71 it would prevent salt-bridging and rendering the kinase inactive. Further, within the hydrophobic pocket III, Glu71 is capable of making a hydrogen bond with the donor residue.

The beginning activation segment of the DFG [expansion] motif spans is between 167 - 169 amino acids [31], within this region lies Asp 168 and it is interacts with the compound, therefore the compound is likely to affect activation of enzyme. In the enzyme hinge region ranges are from 106 - 109, interestingly the compound is capable of interacting with His107 lying within the hinge region. This hinge region connects N- and C- lobes and interacts with ATP through hydrogen bond [31]. Therefore, the compound can able to effectively inhibit ERK2 since it can interact with 3 different amino acids positioned at 3 different crucial sites necessary for the activity of enzyme. In addition it has already been reported that small molecules inhibitors of protein kinases compete with hinge region to compete with ATP molecules [32].

DFT

The connection of DFT observed results were indicates that synthesized compound of 4-(4-hydroxybenzyl)-2-amino-6-hydroxypyrimidine-5-carboxamide is having more stable than the standard



► **Fig. 8** The drug-copper nanocomposites configuration

Uramustine, hence the synthesized compound gained attraction for their further studies.

Optimization

The observed results were indicates that among the selected independent variables, 4-hydroxy benzaldehyde (X_1) and ethyl cyanoacetate (X_2) has significant effects on maximum yield of 4-(4-hydroxybenzyl)-2-amino-6-hydroxypyrimidine-5-carboxamide (74.55-76%), melting point 127 °C and R_f value of 0.59.

► **Table 5** The Percentage of viability of cell death.

| S. No | Concentration of Nanoparticles ($\mu\text{g/mL}$) | % of cell death |
|-------|---|-----------------|
| 1 | 5 | 21.42 |
| 2 | 10 | 29.36 |
| 3 | 15 | 34.51 |
| 4 | 20 | 46.43 |
| 5 | 25 | 54.31 |
| 6 | 30 | 61.5 |
| 7 | 50 | 72.48 |

► **Table 6** Annotation of up-regulated proteins.

| Uniprot ID | Protein Sequence Coverage (%) | Protein names | Gene names | Number of Unique Peptides | Ratio/Fold change a* |
|------------|-------------------------------|---|----------------------------------|---------------------------|----------------------|
| P06733 | 5.8 | Alpha-enolase | ENO1 ENO1L1 MBPB1 MPB1 | 12 | 1.5 |
| P22314 | 12.9 | Ubiquitin-like modifier-activating enzyme 1 | UBA1 A1S9T UBE1 | 6 | 1.6 |
| P11142 | 15.1 | Heat shock cognate 71 kDa protein | HSPA8 HSC70 HSP73 HSPA10 | 11 | 1.7 |
| V9HWG3 | 52.1 | Epididymis secretory protein Li 45 | HEL-S-45 | 9 | 1.7 |
| P42338 | 13 | Phosphatidylinositol 4,5-bisphosphate 3-kinase catalytic subunit beta isoform | PIK3CB PIK3C1 | 5 | 1.8 |
| P10809 | 23 | 60 kDa heat shock protein, mitochondrial | HSPD1 HSP60 | 4 | 1.8 |
| P07437 | 10.3 | Tubulin beta chain (Tubulin beta-5 chain) | TUBB TUBB5 OK/SW-cl.56 | 12 | 1.9 |
| P08238 | 28.3 | Heat shock protein HSP 90-beta | HSP90AB1 HSP90B HSPC2 HSPCB | 12 | 1.9 |
| P08670 | 14.7 | Vimentin | VIM | 7 | 1.9 |
| P21333 | 23.7 | Filamin-A (FLN-A) | FLNA FLN FLN1 | 16 | 2.1 |
| P35579 | 13.1 | Myosin-9 | MYH9 | 8 | 2.1 |
| P08727 | 7.6 | Keratin, type I cytoskeletal 19 | KRT19 | 5 | 2.3 |
| P04406 | 7 | Glyceraldehyde-3-phosphate dehydrogenase | GAPDH GAPD CDABP0047 OK/SW-cl.12 | 8 | 2.3 |
| P68363 | 17.4 | Tubulin alpha-1B chain | TUBA1B | 6 | 2.7 |
| P60709 | 16.1 | Actin, cytoplasmic 1 (Beta-actin) | ACTB | 4 | 2.7 |
| Q8IWP6 | 31.9 | Tubulin beta chain | | 7 | 3.1 |
| O43707 | 23 | Alpha-actinin-4 | ACTN4 | 6 | 3.1 |
| P07355 | 13.7 | Annexin A2 | ANXA2 ANX2 ANX2L4 CAL1H LPC2D | 6 | 3.1 |
| P62736 | 9.8 | Actin, aortic smooth muscle (Alpha-actin-2) | ACTA2 ACTSA ACTVS GIG46 | 6 | 3.1 |
| P13647 | 37.6 | Keratin, type II cytoskeletal 5 | KRT5 | 6 | 4.2 |
| P05787 | 16.1 | Keratin, type II cytoskeletal 8 | KRT8 CYK8 | 5 | 5.6 |

► **Table 7** Annotation of down-regulated proteins.

| Uniprot ID | Protein Sequence Coverage (%) | Protein names | Gene names | Number of Unique Peptides | Ratio/Fold change a * |
|------------|-------------------------------|---|----------------------------------|---------------------------|-----------------------|
| P13639 | 15.9 | Elongation factor 2 | EEF2 EF2 | 4 | 0.1 |
| P12814 | 24.3 | Alpha-actinin-1 | ACTN1 | 5 | 0.2 |
| P14618 | 18.7 | Pyruvate kinase PKM | PKM OIP3 PK2 PK3 PKM2 | 21 | 0.3 |
| P05783 | 12.9 | Keratin, type I cytoskeletal 18 | KRT18 CYK18 PIG46 | 5 | 0.3 |
| P07900 | 36.1 | Heat shock protein HSP 90-alpha | HSP90AA1 HSP90A HSPC1 HSPCA | 4 | 0.4 |
| P02545 | 8.1 | Prelamin-A/C | LMNA LMN1 | 4 | 0.4 |
| P60174 | 45.2 | Triosephosphate isomerase (TIM) | TPI1 TPI | 12 | 0.4 |
| P11021 | 22.7 | 78 kDa glucose-regulated protein | HSPA5 GRP78 | 4 | 0.4 |
| P35222 | 18.3 | Catenin beta-1 | CTNNB1 CTNNB OK/SW-cl.35 PRO2286 | 4 | 0.5 |
| Q16665 | 9.6 | Hypoxia-inducible factor 1-alpha | HIF1A BHLHE78 MOP1 PASD8 | 4 | 0.5 |
| P42224 | 33.1 | Signal transducer and activator of transcription 1-alpha/beta | STAT1 | 4 | 0.5 |
| F5H5D3 | 14.9 | Tubulin alpha chain | TUBA1C | 6 | 0.5 |
| A0A024R321 | 26.8 | Filamin B, beta | FLNB hCG_27732 | 4 | 0.5 |

► **Table 8** KEGG Pathway enrichment analysis.

| Pathway ID | Pathway description | Observed gene count | False discovery rate | Matching proteins in your network (labels) |
|------------|--------------------------------------|---------------------|----------------------|---|
| 4510 | Focal adhesion | 6 | 3.76E-05 | ACTB,ACTN1,ACTN4,CTNNB1,FLNA,PIK3CB |
| 4670 | Leukocyte transendothelial migration | 5 | 3.76E-05 | ACTB,ACTN1,ACTN4,CTNNB1,PIK3CB |
| 4530 | Tight junction | 5 | 5.13E-05 | ACTB,ACTN1,ACTN4,CTNNB1,MYH9 |
| 4520 | Adherens junction | 4 | 0.000116 | ACTB,ACTN1,ACTN4,CTNNB1 |
| 5200 | Pathways in cancer | 6 | 0.000193 | CTNNB1,HIF1A,HSP90AA1,HSP90AB1,PIK3CB,STAT1 |
| 5215 | Prostate cancer | 4 | 0.000201 | CTNNB1,HSP90AA1,HSP90AB1,PIK3CB |
| 4915 | Estrogen signaling pathway | 4 | 0.000261 | HSP90AA1,HSP90AB1,HSPA8,PIK3CB |
| 4810 | Regulation of actin cytoskeleton | 5 | 0.000287 | ACTB,ACTN1,ACTN4,MYH9,PIK3CB |
| 5205 | Proteoglycans in cancer | 5 | 0.000332 | ACTB,CTNNB1,FLNA,HIF1A,PIK3CB |

Chemistry

The IR, ¹H-NMR, mass spectroscopy and elemental analyses for the new compound are in accordance with the assigned structures. The IR spectrum of compound showed bands of hydroxy group at 3368 cm⁻¹ and amine group stretching bands appears at 3312 cm⁻¹. The appearance of a strong intensity band in the IR spectra of compound in the range of 1728 cm⁻¹ attributable to C=O and it provides a strong evidence for the condensation and also confirms the formation of the title compound. The proton magnetic resonance spectra of pyrimidine derivative have been recorded in CDCl₃. In this signals appear at 9.52 (s, 1H, OH), 8.69 (s, 4H, NH₂), 6.72-7.23 (m, 4H, Ar-H), 3.60 (s, 2H, CH₂) ppm respectively. The position and presence of proton signals in the ¹H-NMR spectra of final compounds and mass spectra were confirms further the title moiety. All these observed facts clearly envisages that the 4-(4-hydroxy benzyl)-2-amino-6-hydroxypyrimidine-5-carboxamide formation as indicated in ► **Fig. 3** and it confirms the proposed structure. The characterization of formulated nanoparticles by SEM analysis and X-ray diffraction reports were confirm the formation of copper-drug nanoparticles that of less than 30 nm diameter in size ► **Fig. 8**.

Anticancer activity

The in silico modeling report was encouraging us to perform in vitro anticancer activity and it was screened against MCF7 cancer cell lines by using MTT assay. In a dose-dependent manner the 4-(4-hydroxybenzyl)-2-amino-6-hydroxypyrimidine-5-carboxamide nanoparticles was having capability to decrease viability of cells. The observed results with 5 µg/mL concentration was decreases 21 % viability of cell death as shown in, 25 µg/mL concentration was decreases 54 % viability of cell death and 72 % cell death was observed with 50 µg/mL concentration of nanoparticles. The findings of above results were directly indicating that the increasing concentration of nanoparticles was directly decreases viability of cell death ► **Table 5**.

Quantitative proteomics studies

Quantitative proteomics was performed for our drug to understand the molecular mechanism at its functional level. After applying the filtering criteria of greater than 2 unique peptides with 5 % FDR and a Q-value of zero 34 proteins were differentially expressed in treatment groups. Among them, 21 proteins were up regulated with intensity ratio > 1.5 ► **Table 6** and 13 proteins were down regulated

proteins with intensity ratio <0.05 ▶ **Table 7**. Gene ontology and interactive pathway maps were represented in the Further, KEGG pathway enrichment analysis was performed and key pathways and its regulatory proteins were represented in the ▶ **Table 8**. Over all, the identified regulatory pathways and differentially expressed proteins on drug treatment were well established and linked with the major regulatory networks in carcinogenesis, tumor progression and metastasis in multiple cancer types.

Conclusion

A general method has been developed for the synthesis of 4-(4-hydroxybenzyl)-2-amino-6-hydroxypyrimidine-5-carboxamide. This process involved in the addition of ethyl cyanoacetate, 4-hydroxy benzaldehyde and guanidine hydrochloride with various temperatures and rotation per minute (rpm). On the basis of optimization measurement, the synthesis of 4-(4-hydroxybenzyl)-2-amino-6-hydroxypyrimidine-5-carboxamide has been proposed for achieve high yield with accuracy of physiochemical parameters like melting point and R_f value. The optimization behavior of title compound was studied which showed that more positive potential value of the yield 82 % as compared to those for corresponding synthesis. The in vitro biocompatibility study was highlighting that 4-(4-hydroxybenzyl)-2-amino-6-hydroxypyrimidine-5-carboxamide nanoparticles was having significant cell viability and proliferation and 92 % MCF7 cellular death was confirming that it was aligned with in silico modeling study report. Therefore the formulated nanoparticles might be an exclusive carrier in the connection of drug delivery against cancer activity and we introduced graph theoretical analysis, density functionality theory and proteomics reports are having a great impact and it will open a new resource in the identification of alternative treatment of various cancers.

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Conflict of Interest

Authors declare there is no conflict of interest.

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