Antiprotozoal Germacranolide Sesquiterpene Lactones from *Tanacetum sonbolii*

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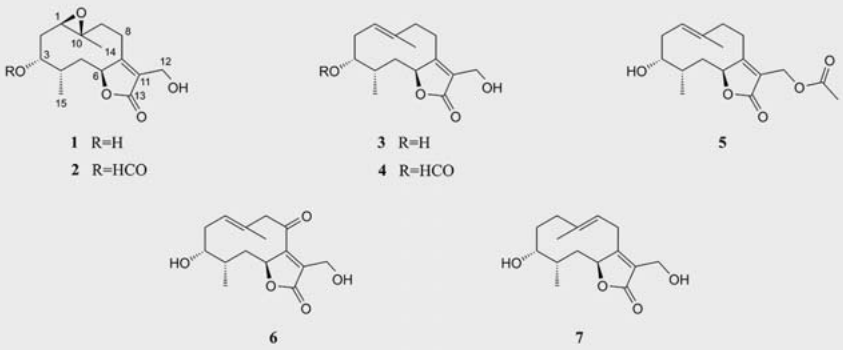
ABSTRACT

A phytochemical investigation of extracts from flowers and aerial parts of *Tanacetum sonbolii* afforded 7 new germacranolide sesquiterpene lactones. The structures were established by a combination of 1- and 2-dimensional nuclear magnetic resonance spectroscopy, high-resolution mass spectrometry, and electronic circular dichroism. The *in vitro* antiprotozoal activity of the compounds against *Trypanosoma brucei rhodesiense* and cytotoxicity against rat myoblast (L6) cells were determined. Compounds **4** and **5** showed IC_{50} values of 5.1 and 10.2 µM and selectivity indices of 3.9 and 4.0, respectively.

Introduction

Protozoan parasites cause serious public health problems in many parts of the world, particularly in Sub-Saharan Africa [1]. No vaccines are currently available for protozoan diseases, and antiprotozoal strategies rely on vector control and pharmacotherapy for infected patients. The treatment options for sleeping sickness (human African trypanosomiasis) remain insufficient, in particular for the late stage of the disease [2, 3]. Given the successful track record of natural product leads such as quinine and artemisinin [4], a continued search for new antiprotozoal compounds from higher plants is warranted.

The genus *Tanacetum* (Asteraceae) comprises approximately 200 species that occur in many regions of the northern hemisphere [5]. Of the 36 *Tanacetum* species growing in Iran, a total of 16 are endemic [6]. *Tanacetum sonbolii* Mozaff. grows in Takab, West Azerbaijan Province of Iran, and has been recently identified as an endemic species [7–9]. α-Cadinol, globulol, and 1,8-cineole were identified as major compounds in the essential oil [10], which was shown to possess antioxidant, anti-seizure, and pain re-



▶ **Fig. 1** Structures of compounds **1**–**7**.

▶ **Table 1** ^{13}C NMR (125 MHz) spectroscopic data of **1**–**7** in $CDCl_3$ (δ in ppm).

Position	1	2	3	4	5	6[a]	7
1	61.7	61.2	126.1	124.7	126.2	125.9	29.7
2	27.4	25.6	37.2	37.2	27.4	36.8	35.5
3	74.1	75.0	73.8	75.0	74.0	75.4	73.9
4	37.1	34.4	41.1	34.5	37.1	38.7	36.8
5	37.2	36.8	38.4	36.8	35.5	23.1	37.2
6	82.0	81.0	82.1	81.4	81.7	81.8	82.1
7	166.0	166.0	168.6	168.0	168.6	168.6	166.9
8	23.0	22.8	23.0	25.6	23.8	210.0	23.0
9	35.6	35.4	35.5	35.4	37.2	37.3	125.7
10	60.7	60.7	134.3	136.0	132.4	133.9	134.3
11	126.3	126.0	125.7	126.2	122.7	125.5	126.3
12	54.6	53.7	54.2	54.3	54.9	53.5	54.1
13	173.8	171.0	174.2	173.9	170.8	173.9	174.1
14	17.0	16.9	17.0	17.0	17.0	17.9	17.0
15	19.9	19.6	20.1	19.9	19.8	19.7	19.9
HCO	–	161.0		160.1			
CH_3CO					20.8		
CH_3CO					169.7		

[a] ^{13}C NMR data of compound 6 was extracted from HSQC-DEPT and HMBC spectra due to a low amount of sample.

lieving properties [10–13]. In contrast, nonvolatile constituents of the species have not been studied so far. In a continued effort to discover new bioactive secondary metabolites from endemic Iranian plants [14–16], we here report on the isolation, structure elucidation, and *in vitro* antitrypanosomal activity of 7 new germacranolide sesquiterpene lactones from the aerial parts and flowers of *T. sonbolii*.

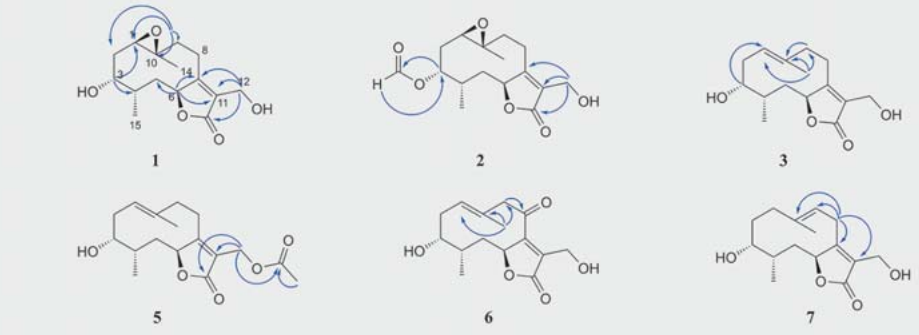
Results and Discussion

Sesquiterpene lactones **1**, **3**, and **5** were isolated from the flowers and **2**, **4**, **6**, and **7** from the aerial parts of *T. sonbolii* (▶ **Fig. 1**). The structure elucidation was achieved using extensive NMR spectroscopy and HRESI-TOFMS, and the absolute configuration was established by a comparison of experimental and calculated electronic circular dichroism (ECD) spectra.

Compound **1** was obtained as a white amorphous powder. The HRESIMS of **1** showed a molecular ion at *m/z* 283.1543 $[M+H]^+$ (calcd. 283.1540), indicating a molecular formula of $C_{15}H_{22}O_5$ and 5 indices of hydrogen deficiency. The ^{13}C NMR spectrum (▶ **Table 1**) showed 15 carbon resonances, which were assigned with the aid of HSQC and DEPTQ spectra as 1 methyls, 5 methylenes, 4 methines, and 4 quaternary carbons. The ^{1}H NMR spectrum (▶ **Table 2**) displayed the characteristic signals of 2 aliphatic methyl groups at δ_H 0.94 and 1.43, 3 oxygenated methine protons at δ_H 3.48, 4.11, and 5.78, and 1 oxygenated methylene proton at δ_H 4.33. The NMR data of **1** were similar to those of germacranolide sesquiterpene lactones that had been reported previously

▶ **Table 2** ^{1}H NMR (500 MHz) data spectroscopic of **1–7** in $CDCl_3$ (δ in ppm, J in Hz).

Position	1	2	3	4	5	6	7
1	3.48 d (10.5)	3.26 d (11.5)	5.73 br s	5.60 br s	5.86 br s	5.75 br s	1.22 m
2β	1.60 ddd (15.0, 10.5, 1.0)	1.74 dd (16.0, 11.5)	2.24 m	2.55 m	2.46 m	2.27 m	2.25 m
2α	2.29 m	2.19 dd (16.0, 5.5)	2.42 m			2.44 m	2.45 m
3	4.11 br d (5.5)	5.06 br s	3.77 br s	4.94 br s	3.82 br s	3.77 m	3.77 m
4	2.37 m	1.25 m 2.25 m	2.12 m	2.24 m	2.18 m	2.12 m	2.12 m
5β	1.38 m	1.25 m	1.07 m	1.17 m	1.33 m	2.38 m	1.10 m
5α	1.90 dd (15.0, 12.0(	1.86 m	1.85 m	1.90 m		3.10 m	1.83 m
6	5.78 d (12.0)	5.36 d (9.0)	5.6 br s	5.10 br s	5.70 m	5.70 br s	5.24 s
8α	2.44 dd (16.5, 13.0)	2.51 m	2.42 m	2.45 m	2.40 m	–	2.40 m
8β	3.14 ddd (16.5, 7.0, 1.5)	3.14 ddd (16.5, 7.5, 1.0)	2.92 dd (11.5, 3.5)	2.99 m	2.97 m		2.92 m
9α	1.31 m	1.27 m	2.31 m	2.13 m	1.19 m	1.12 m	5.75 br s
9β	2.35 m	2.27 m	2.40 m	2.38 m	1.87 m	1.85 m	
12	4.33 d (4.0)	4.10 br s	4.22 br s	4.14 br s	4.70 br s	4.20 br s	4.2 br s
14	1.43 s	1.38 s	1.64 s	1.69 s	1.70 s	1.68 s	1.66 s
15	0.94 d (7.0)	0.99 d (7.0)	0.89 d (6.5)	0.95 d (7.5)	0.93 d (6.5)	0.91 d (6.5)	0.88 d (6.5)
HCO		8.00 s		8.04 s			
CH_3CO					1.99 s		

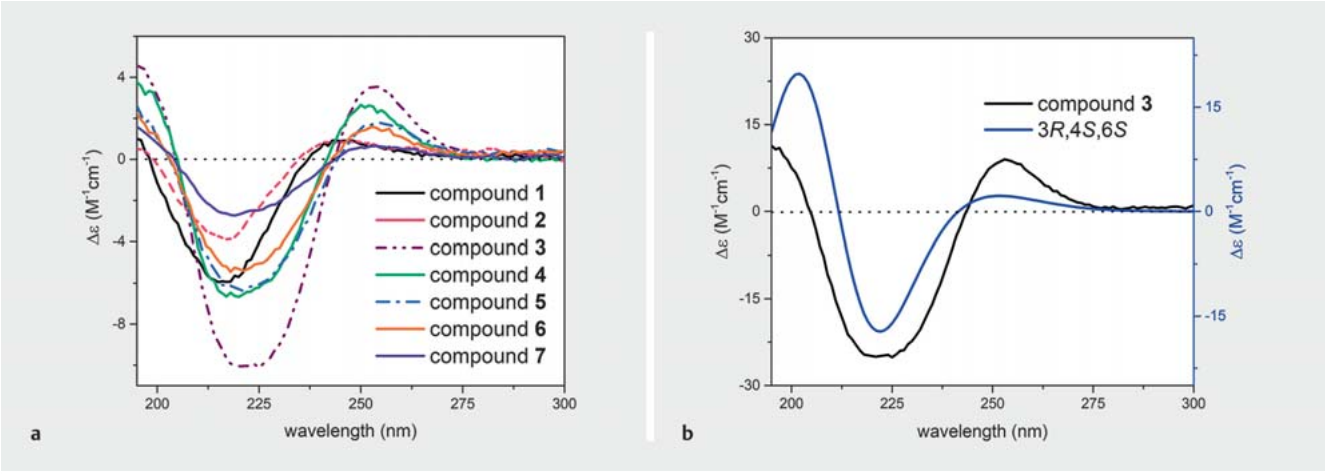


▶ **Fig. 2** Selected HMBC correlations.

from *Tanacetum* species [17]. The ^{1}H-^{1}H COSY and HSQC spectra indicated the presence of 3 contiguous fragments: -CH(1)-CH_2(2)-CH(3)-CH(4)-CH_2(5)-CH(6)-, -CH(4)-CH_3(15)-, and -CH_2(8)-CH_2(9)-. HMBC correlations from H_2-12 (δ_H 4.33) to the carbonyl carbon (C-13, δ_C 173.8) and to olefinic carbons C-11 (δ_C 126.3) and C-7 (δ_C 166.0), from H_2-8 (δ_H 3.14) to C-7, C-11, C-6 (δ_C 82.0), C-9 (δ_C 35.6), and C-10 (δ_C 60.7), and from H-6 (δ_H 5.78) to C-5 (δ_C 37.2), C-7, and C-11 confirmed the presence of an α-methyl-γ-butyrolactone moiety with a hydroxyl group at C-12. Thus, the double bond was located in the lactone ring instead of the exocyclic methylene group that is usually seen in germacranolides. HMBC correlations from H_3-14 (δ_H 1.43) to C-1, C-9, and C-10 confirmed the attachment of the methyl at C-10 next to an epoxy moiety. HMBC correlations from H-3 (δ_H 4.11) to C-1 and C-4 (δ_C 37.1) corroborated the hydroxyl group at C-3 (δ_C 74.1). The HMBC correlations from H-1 (δ_H 3.48) to C-2 (δ_C 27.4) and C-3 (δ_C 74.1) and from H_3-15 (δ_H 0.94) to C-4 and C-3 corroborated the planar structure of **1** (▶ **Fig. 2**).

Analysis of the NOESY spectrum established the relative configuration of **1**. Cross-peaks between H-6 (δ_H 5.78) and H-1 (δ_H 3.48) and between H-1, H_3-14, and H_α-2 (δ_H 2.29) indicated that they were all located on the same face of the molecule. The cross-peak between H_β-2 (δ_H 1.60) and H-3 confirmed the α orientation of the hydroxyl group at C-3, and the NOESY correlation between H_β – 5 (δ_H 1.90) and H-4 (δ_H 2.37) confirmed the α orientation of the methyl group at C-4. The configuration of stereogenic centers of **1** was established by ECD spectroscopy as 3*R*,4*S*,6*S* given that the experimental ECD spectrum of **1** was in good agreement with the spectrum calculated for the 3*R*,4*S*,6*S* stereoisomer (▶ **Fig. 3**).

Compound **2** had a molecular formula of $C_{16}H_{22}O_6$, as deduced from the HRESIMS (m/z 311.1490 $[M+H]^+$; calcd. 311.1489), corresponding to 6 indices of hydrogen deficiency. The ^{1}H NMR data of **2** (▶ **Table 2**) were similar to those of **1**. The



▶ **Fig. 3** **a** Experimental ECD spectra of compounds **1**–**7** in MeOH. **b** Comparison of experimental and TDDFT calculated ECD spectra of compound **3**.

major differences were in the presence of an additional aldehyde proton (δ_H 8.0) attached to the carbonyl carbon at δ_C 161.0. Also, the carbinolic methine proton at C-3 exhibited a paramagnetic shift from 4.11 ppm in **1** to 5.06 ppm in **2** and showed an HMBC correlation to the formyl group. Thus, compound **2** was the formyl ester of **1**. The experimental ECD spectrum of **2** was similar to that of **1**, and the configuration of stereogenic centers was thus established as 3*R*,4*S*,6*S*.

The HRESIMS spectrum of compound **3** showed a molecular ion at *m/z* 267.1588 $[M+H]^+$ (calcd. 267.1591), corresponding to a formula of $C_{15}H_{22}O_4$. 1D and 2D NMR spectra suggested a germacranolide scaffold. Compared to **1**, the structural differences of **3** were in the presence of 2 additional olefinic carbons at δ_C 125.7 and 134.3 instead of 2 oxygenated carbons in **1**. HMBC correlations from H_3-14 (δ_H 1.64) to C-9 (δ_C 36.5), C-10 (δ_C 134.3) and C-1 (δ_C 126.1) confirmed the location of this double bond between C-1 and C-10. The relative configuration of **3** was established by NOESY data. A cross-peak between H-3 (δ_H 3.77) and H-4 (δ_H 2.12) indicated that they were cofacial with *syn* orientation. The experimental ECD spectrum of **3** showed 2 positive cotton effects (CE) at 255 and 205 nm along with a negative CE at 225 nm which were due to π–π* transition of the lactone moiety. The ECD spectra for all possible stereoisomers of **3** were calculated, and the spectrum of the 3*R*,4*S*,6*S* stereoisomer showed excellent match with the experimental spectrum (▶ **Fig. 3**). Thus, the absolute configuration of **3** was established as 3*R*,4*S*,6*S*.

Compound **4** had a molecular formula of $C_{16}H_{23}O_5$ (HRESIMS *m/z* 295.1542 $[M+H]^+$, calcd. 295.1540). The NMR data (▶ **Tables 1** and **2**) of **4** closely reassembled those of **3** except for the presence of a formyl residue (δ_H 8.03 and δ_C 160.1) as in compound **2**. The HMBC correlation from the formyl hydrogen to C-3 (δ_C 75.0) confirmed the attachment at C-3. The relative configuration was established by an NOESY spectrum and corresponded with that of **3**. The ECD spectrum of **4** corresponded to that of **3**, and the absolute configuration of **4** was thus established as 3*R*,4*S*,6*S*.

HRESIMS data of compound **5** (*m/z* 309.1698 $[M+H]^+$, calcd. 309.1697) indicated a molecular formula of $C_{17}H_{24}O_5$. The NMR data (▶ **Tables 1** and **2**) closely reassembled to those of **3**. However, an extra methyl signal (δ_H 1.99 and δ_C 20.8) and an extra carbonyl resonance (δ_C 169.7) were present in **5**. HMBC correlations from the additional methyl (δ_H 1.99) and from H_2-12 (δ_H 4.7) to the carbonyl group at δ_C 169.7, together with the downfield shift of H_2-12 (δ_H 4.7 in **5** vs. 4.22 in **3**), confirmed that **5** was an acetyl derivative of **5**. The NOESY and ECD spectra of **5** closely resembled those of **3,** and the absolute configuration was established as 3*R*,4*S*,6*S*.

A molecular formula of $C_{15}H_{20}O_5$ (HRESIMS *m/z* 303.1205 $[M+Na]^+$, calcd. 303.1203) was assigned to compound **6**. The NMR data were reminiscent of a germacranolide skeleton bearing a double bond between C-1 and C-10. Compared to **3**, diagnostic differences were in the presence of an additional carbonyl (δc 210.0) and in the absence of a methylene group. In the HMBC spectrum, a correlation of one of the diastereotopic methylene protons (δ_H 1.12) at C-9 to this carbonyl group observed. Hence, the carbonyl group was at C-8. The relative configuration was established with the aid of the NOESY spectrum. The ECD spectrum (▶ **Fig. 3**) closely resembled that of **3**, and the absolute configuration of compound **6** was established as 3*R*,4*S*,6*S*.

Compound **7** had the same molecular formula as **3** ($C_{15}H_{22}O_4$; HRESIMS *m/z* 267.1588 $[M+H]^+$, calcd. 267.1591). The NMR data closely resembled those of **3**, In the COSY spectrum a correlation of H_2-8 (δ_H 2.40 and 2.92) to H-9 (δ_H 5.75) was observed, and in the HMBC spectrum these 2 diastereotopic protons correlated to 4 olefinic carbons (δ_C 125.7, 126.3, 134.3, and 166.9) (▶ **Fig. 2**). Thus, the double bond was located between C-9 and C-10. The relative configuration of **7** was determined from NOESY correlations and was in accordance with that of **3**, and the ECD spectrum also matched with that of **3** (▶ **Fig. 3**). Thus, the absolute configuration was established as 3*R*,4*S*,6*S*.

The *in vitro* antiprotozoal activity and cytotoxicity of compounds **1**–**7** was evaluated against *Trypanosoma brucei rhodesiense* and rat myoblast (L6) cells, respectively, and the selectivity indices (SI) for tested compounds were calculated (▶ **Table 3**). Compounds **4** and **5** showed the most potent inhibitory activity

against *T. b. rhodesiense* with IC_{50} values 5.1 µM (SI 3.9) and 10.2 µM (SI 4.0), respectively. Epoxides **1** and **2** exhibited only marginal activity, while **3, 6**, and **7** were inactive at the highest test concentration. From the limited number of compounds tested here, only preliminary observations on structural requirements for activity are possible. When comparing active compound **4** with marginally active epoxide **2**, it seems that the conformation of the 10-membered ring affects the degree of activity. A formyl or acetyl moiety as in **4** and **5** appears to enhance activity when compared to parent compound **3**. Numerous sesquiterpene lactones with different scaffolds reportedly show *in vitro* activity against *T. b. rhodesiense* [18–21]. However, these compounds all bear an *α,β*-unsaturated carbonyl group that may react with biological nucleophiles such as thiol groups of cysteine residues in proteins [18]. In contrast, the *α,β*-unsaturated carbonyl group in active compounds **4** and **5** cannot act as a Michael acceptor. Additional, structurally related germacranolides need to be tested to unravel the features that are relevant for activity.

NMR spectra (1D and 2D) of compounds **1–7** as well as the dose-response curves leading to the IC_{50} determination of compounds **1, 2, 4**, and **5** are available as Supporting Information.

▶ **Table 3** *In vitro* activity of compounds **1–7** against *T. b. rhodesiense* (STIB 900) and cytotoxicity in L6 cells.

Compound	T. b. rhodesiense IC_{50}[a]	L6 cells
1	50.3 (44.1–56.2); 3.0[b]	150.9 (170.6–194.1)
2	88.1 (55.5–123.6); 3.3[b]	287.0 (282.3–291.7)
3	> 200	–
4	10.2 (9.5–10.9); 4.0[b]	40.9 (39.7–42.1)
5	5.1 (1.7–8.5); 3.9[b]	19.9 (17.9–21.9)
6	> 200	–
7	> 200	–
Melarsoprol	0.003	
Podophyllotoxin		0.005

[a] Values are expressed in µM. Each value corresponds to the mean (CI 95 %) from 2 independent assays. [b] SI: IC_{50} in L6 cells divided by IC_{50} in the titled parasitic strain.

Materials and Methods

General experimental procedures

The JASCO P-2000 digital polarimeter was used for measuring optical rotation. UV spectra were recorded using a Shimadzu UV-2501PC spectrophotometer. ECD spectra were recorded in MeOH on a Chirascan (Applied Photophysics) spectrometer with 1-mm path precision cells (110 QS, Hellma Analytics), and data were analyzed with Pro-Data V2.4 software. NMR spectra were recorded at a target temperature of 18 °C on a Bruker Avance III 500-MHz spectrometer operating at 500.13 MHz for ^{1}H and 125.77 MHz for ^{13}C. A 1-mm TXI-microprobe with z-gradient was used for ^{1}H-detected experiments. ^{13}C NMR spectra were recorded with a 5-mm BBO probe head with z-gradient. Spectra were analyzed using Bruker TopSpin 3.5 software. Deuterated solvents for NMR (100 % D) were purchased from Armar Chemicals. HRESIMS data were recorded in positive ion mode on an Agilent 1290 Infinity system with an Agilent 6540 UHD Accurate-Mass Quadrupole Time-of-Flight detector (G6540A). HPLC separations were performed on a Knauer HPLC system consisting of a mixing pump with degasser module, photodiode array (PDA) detector, and an autosampler. Knauer Eurospher Π 100–5 RP C18 (5 µm, 4.6 × 250 mm i. d.) and SunFire Prep C18 ODB (5 µm, 19 × 50 mm i. d.) columns were used for analytical and semipreparative separations, respectively. Solvents used for extraction and column chromatography were of technical grade and were distilled before use. HPLC-grade solvents (Merck) were used for HPLC. Silica gel (70–230 mesh) for column chromatography, precoated silica gel F_{254} (20 × 20 cm) plates, anisaldehyde, glacial acetic acid, and sulfuric acid were all from (Merck).

Plant material

Aerial parts and flowers of *T. sonbolii* were collected from Takab, Baderloo village, West Azerbaijan Province, Iran, in June 2015 and identified by Dr. Ali Sonboli. A voucher specimen (MPH-2556) has been deposited at the Herbarium of the Medicinal Plants and Drugs Research Institute of Shahid Beheshti University, Tehran, Iran.

Extraction and preparative isolation

Air-dried flowers of *T. sonbolii* (190 g) were powdered and macerated with acetone (5 × 2 L). The combined extracts were concentrated to dryness, and the residue (7 g) was subjected to silica gel column chromatography (70–230 mesh, 45.0 × 300 mm, 230 g) using a step gradient of *n*-hexane–EtOAc (100 : 0 → 0 : 100), followed by EtOAc/MeOH (100 : 0 → 50 : 50). A total of 55 fractions of 250 mL each were collected and pooled after TLC analysis (detection at 254 nm and after spraying with anisaldehyde-sulfuric acid reagent) to 12 fractions. Fraction 8 (110 mg; eluted with *n*-hexane–EtOAc [50 : 50]) was separated by preparative HPLC ($MeCN/H_2O$, 60 : 40, v/v) to yield **5** (5.6 mg, t_R 10.3 min). Fraction 11 (200 mg; eluted with *n*-hexane–EtOAc [25 : 75]) was separated by preparative HPLC ($MeCN/H_2O$, 40 : 60, v/v) to yield compounds **1** (2 mg, t_R 12.3 min) and **3** (4 mg, t_R 16.7 min).

The air-dried aerial parts of *T. sonbolii* (1.6 kg) were milled and macerated with ethyl acetate (5 × 7 L). The extract was concentrated *in vacuo* to afford 60 g of a dark gummy residue. The residue was separated on a silica gel column (70–230 mesh, 60.0 × 1180 mm, 850 g) with a step gradient of *n*-hexane–EtOAc (100 : 0 → 0 : 100) as eluent, followed by EtOAc containing increasing concentrations of MeOH (up to 100 %). A total of 21 fractions were collected on the basis of TLC analysis. Fraction 12 (1.8 g; eluted with *n*-hexane–EtOAc [30 : 70]) was separated on a silica gel column (70–230 mesh, 25 × 450 mm, 150 g) eluted with $CH_2Cl_2/(CH_3)_2CO$ (70 : 30). Fractions were pooled on the basis of TLC patterns to give 5 subfractions ($F12_1$–$F12_5$). Subfraction $F12_3$ (60 mg) was separated on a silica gel column (70–230 mesh, 20 × 600 mm, 90 g) using $CH_2Cl_2/(CH_3)_2CO$ (80 : 20) as mobile phase, and 5 subfractions ($F12_{3.1}$–$F12_{3.5}$) were obtained. The precipitate of subfraction $F12_{3.3}$ was recrystallized from $(CH_3)_2CO$ to afford compound **4** (1.2 mg). $F12_4$ was separated by preparative

HPLC (MeCN/H_2O, 50 : 50, v/v) to yield compounds **2** (1.5 mg, t_R 13.1 min) and **6** (2.5 mg, t_R 17.5 min). Fraction 14 (120 mg; eluted with *n*-hexane–EtOAc [45 : 55]) was separated by preparative HPLC (MeCN/H_2O, 60 : 40, v/v) to yield compounds **7** (2.5 mg, t_R 14.3 min). The purity of the isolated compounds was assessed by ^{1}H NMR and HPLC analyses and found to be > 95% for compounds **1**–**5** and > 90% for **6**–**7**.

Compound ***1***: White powder; $[\alpha]_D^{25}$ = + 25.9 (*c* 1.0, MeOH); UV (MeOH) λ_{max} (log ε) 218 (4.03) nm; ECD (MeOH) 217 nm ($\Delta\varepsilon$ − 13.89) and 247 nm ($\Delta\varepsilon$ + 2.08). For ^{1}H and ^{13}C NMR spectroscopic data, see ▶ **Tables 1** and **2**; HRESITOFMS *m/z* 283.1543 $[M + H]^+$ (calcd. for 283.1540).

Compound ***2***: White powder; $[\alpha]_D^{25}$ = + 3.3 (*c* 1.0, MeOH); UV (MeOH) λ_{max} (log ε) 220 (3.33) nm; ECD (MeOH) 217 nm ($\Delta\varepsilon$ − 2.06) and 250 nm ($\Delta\varepsilon$ + 0.55). For ^{1}H and ^{13}C NMR spectroscopic data, see ▶ **Tables 1** and **2**; HRESITOFMS *m/z* 311.1490 $[M + H]^+$ (calcd. for 311.1489).

Compound ***3***: White powder; $[\alpha]_D^{25}$ = + 9.9 (*c* 1.0, MeOH); UV (MeOH) λ_{max} (log ε) 211 (4.04) nm; ECD (MeOH) 224 nm ($\Delta\varepsilon$ − 24.71) and 254 nm ($\Delta\varepsilon$ + 8.81). For ^{1}H and ^{13}C NMR spectroscopic data, see ▶ **Tables 1** and **2**; HRESITOFMS *m/z* 267.1588 $[M + H]^+$ (calcd. for 267.1591).

Compound ***4***: White powder; $[\alpha]_D^{25}$ = + 13.7 (*c* 1.0, MeOH); UV (MeOH) λ_{max} (log ε) 267 (5.13) nm; ECD (MeOH) 220 nm ($\Delta\varepsilon$ − 14.95) and 252 nm ($\Delta\varepsilon$ + 5.88). For ^{1}H and ^{13}C NMR spectroscopic data, see ▶ **Tables 1** and **2**; HRESITOFMS *m/z* 295.1542 $[M + H]^+$ (calcd. for 295.1540).

Compound ***5***: White powder; $[\alpha]_D^{25}$ = + 13.2 (*c* 1.0, MeOH); UV (MeOH) λ_{max} (log ε) 265 (5.13) nm; ECD (MeOH) 221 nm ($\Delta\varepsilon$ − 13.64) and 255 nm ($\Delta\varepsilon$ + 3.78). For ^{1}H and ^{13}C NMR spectroscopic data, see ▶ **Tables 1** and **2**; HRESITOFMS *m/z* 309.1698 $[M + H]^+$ (calcd. for 309.1697).

Compound ***6***: White powder; $[\alpha]_D^{25}$ = + 16.2 (*c* 1.0, MeOH); UV (MeOH) λ_{max} (log ε) 217 (3.87) nm; ECD (MeOH) 218 nm ($\Delta\varepsilon$ − 12.82) and 253 nm ($\Delta\varepsilon$ + 3.73). For ^{1}H and ^{13}C NMR spectroscopic data, see ▶ **Tables 1** and **2**; HRESITOFMS *m/z* 303.1205 $[M + Na]^+$ (calcd. for 303.1203).

Compound ***7***: White powder; $[\alpha]_D^{25}$ = + 14.929 (*c* 1.0, MeOH); UV (MeOH) λ_{max} (log ε) 214 (3.19) nm; ECD (MeOH) 218 nm ($\Delta\varepsilon$ − 12.82) and 253 nm ($\Delta\varepsilon$ + 3.73). For ^{1}H and ^{13}C NMR spectroscopic data, see ▶ **Tables 1** and **2**; HRESITOFMS *m/z* 267.1588 $[M + H]^+$ (calcd. for 267.1591).

ECD computational details

Conformational analysis of **3** was performed with MacroModel 9.8 software (Schrödinger LLC) employing the OPLS 2005 (Optimized Potential for Liquid Simulations) force field in H_2O. Conformers occurring within a 2 kcal/mol energy window from the global minimum were selected for geometrical optimization and energy calculation using DFT-B3LYP/6-31G** in MeOH with the Gaussian 09 program package [22]. The vibrational evaluation was done at the same level to confirm minima. Excitation energy (denoted by wavelength in nm), rotatory strengths dipole velocity (R_{vel}), and dipole length (R_{len}) were calculated in MeOH by TD-DFT/B3LYP/6-31G** using the selfconsistent reaction field (SCRF) method with the conductor-like polarizable continuum model (CPCM). ECD curves were obtained by rotatory strengths with a half-band of 0.25 eV and UV shift using SpecDis v1.64 [23]. ECD spectra were calculated from the spectra of individual conformers according to their contribution calculated by Boltzmann weighting.

In vitro biological testing

The *in vitro* activities against the protozoan parasites *T. b. rhodesiense* (STIB900) bloodstream forms and cytotoxicity in L6 cells (rat skeletal myoblasts) were determined at the Swiss Tropical and Public Health Institute as reported previously [24, 25]. Assays were run in singleton and repeated at least 2 times. The positive control podophyllotoxin was purchased from Sigma, and Melarsoprol was received from the World Health Organization. Their purity was > 95% according to the suppliers.

Activity against *T. b. rhodesiense* STIB900

The stock was isolated in 1982 from a human patient in Tanzania and after several mouse passages cloned and adapted to axenic culture conditions [26]. Minimum essential medium (MEM) (50 µL) supplemented with 25 mM HEPES, 1 g/L additional glucose, 1% MEM nonessential amino acids (100 ×), 0.2 mM 2-mercaptoethanol, 1 mM Na-pyruvate, and 15% heat inactivated horse serum was added to each well of a 96-well microtiter plate. Compounds were dissolved in DMSO (10 mg/mL) and stored at − 20 °C until testing. Final test concentrations did not exceed a 1%. DMSO and assays were done at least 3 times independently. Serial dilutions of compounds prepared to cover in a range from 100 to 0.001 µg/mL were prepared. Then 104 bloodstream forms of *T. b. rhodesiense* STIB 900 in 50 µL of medium were added to each well and the plate incubated at 37 °C under a 5% CO_2 atmosphere for 72 h. Alamar blue solution (10 µL, 12.5 mg resazurin dissolved in 100 mL distilled water) were then added to each well and incubation continued for a further 2–4 h [27]. After that he plate was read in a Spectramax Gemini XS microplate fluorometer (Molecular Devices Corporation) using an excitation wavelength of 536 nm and emission wavelength of 588 nm. The IC_{50} values were calculated by linear regression [28] from the sigmoidal dose inhibition curves using SoftmaxPro software (Molecular Devices Cooperation). Melarsoprol (received from WHO) is used as control.

In vitro cytotoxicity with L-6 cells

Assays were performed in 96-well microtiter plates, each well containing 100 µL of RPMI 1640 medium supplemented with 1% L-glutamine (200 mM) and 10% FBS and 4000 L-6 cells (a primary cell line derived from rat skeletal myoblasts) [29, 30]. Serial drug dilutions of eleven 3-fold dilution steps covering a range from 100 to 0.002 µg/mL were prepared. After 70 h of incubation, the plates were inspected under an inverted microscope to assure growth of the controls and sterile conditions. Ten microliters of alamarBlue was then added to each well and the plates incubated for another 2 h. Then the plates were read with a Spectramax Gemini XS microplate fluorometer (Molecular Devices Cooperation) using an excitation wave length of 536 nm and an emission wave length of 588 nm. The IC_{50} values were calculated by linear regression [28] from the sigmoidal dose inhibition curves using SoftmaxPro software (Molecular Devices Cooperation).

Supporting information

NMR spectra (1D and 2D) of compounds **1**–**7** as well as the dose-response curves leading to the IC_{50} determination of compounds **1**, **2**, **4**, and **5** are available as Supporting Information.

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Conflict of Interest

The authors declare no conflict of interest.

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