

Comparison of Maternal Serum Levels and Placental mRNA Levels of Dickkopf-1 in Preeclamptic and Normal Pregnant Women at Delivery

Vergleich von Dickkopf-1-Konzentrationen in mütterlichem Serum und placentärem mRNA in Frauen mit Präeklampsie und Frauen mit normaler Schwangerschaft bei der Geburt




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ABSTRACT

Background Preeclampsia remains a major cause of perinatal and maternal mortality and morbidity worldwide. Wnt/ β -catenin signaling is known to be critically involved in placenta development processes. Dickkopf-1 (DKK1) is a key regulator of this transduction pathway. The aim of this study is to compare maternal serum DKK1 levels and placental mRNA levels of *DKK1* and β -catenin in preeclamptic and normal pregnant women at delivery.

Methods The present study included 30 women with preeclampsia and 30 women with normal pregnancy. Maternal serum DKK1 levels were measured by ELISA. Placental mRNA levels of *DKK1* and β -catenin were detected using RT-PCR.

Results Decreased maternal serum DKK1 levels were associated with worse maternal and fetal complications including HELLP syndrome, determination of one or more pathological symptom and IUGR diagnosis. No significant difference in maternal serum DKK1 levels was reported between preeclamptic women and women with normal pregnancy. Placental mRNA *DKK1* levels were lower in preeclamptic women compared with normal pregnant women. Placental mRNA β -catenin levels showed no significant difference between two groups.

Conclusions Our findings reported the aberrant placental mRNA *DKK1* levels in patients with preeclampsia. In addition, worse preeclampsia features were associated with decreased maternal serum DKK1 levels. Hence, aberrant Wnt/ β -catenin signaling might present a plausible mechanism in preeclampsia pathogenicity. Dysregulated expression of DKK1 at gene level in the placenta but not at protein level in the maternal serum might confirm the notion that preeclampsia is a type of placenta-derived disease.

ZUSAMMENFASSUNG

Hintergrund Präeklampsie ist immer noch eine der wesentlichen Ursachen für die perinatale und mütterliche Mortalität und Morbidität weltweit. Wnt/ β -Catenin-Signalisierung spielt bekanntlich eine wichtige Rolle bei der Entwicklung der Pla-

zenta, und Dickkopf-1 (DKK1) ist ein zentraler Regulator dieses Transduktionswegs. Ziel dieser Studie war es, die DKK1-Konzentrationen im mütterlichen Serum und die plazentären Konzentrationen von *DKK1* und β -Catenin in Frauen mit Präeklampsie und in Frauen mit normaler Schwangerschaft bei der Geburt zu vergleichen.

Methoden Es wurden insgesamt 30 Frauen mit Präeklampsie und 30 Frauen mit normalem Schwangerschaftsverlauf in die Studie aufgenommen. DKK1-Konzentrationen im mütterlichen Serum wurden mit ELISA gemessen. Die plazentären mRNA-Konzentrationen von *DKK1* und β -Catenin wurden mithilfe von RT-PCR ermittelt.

Ergebnisse Niedrige DKK1-Konzentrationen im mütterlichen Serum waren mit schlechteren mütterlichen und fötalen Komplikationen assoziiert, darunter HELLP-Syndrom, einem oder mehreren pathologischen Symptomen bzw. der Diagnose einer intrauterinen Wachstumsretardierung. Es wurde kein signifikanter Unterschied in den DKK1-Konzentrationen im mütterlichen Serum zwischen Frauen mit Präeklampsie und

Frauen mit normaler Schwangerschaft festgestellt. Die plazentären mRNA-*DKK1*-Konzentrationen waren niedriger bei Frauen mit Präeklampsie verglichen mit den Konzentrationen bei Frauen mit normaler Schwangerschaft. Es fanden sich aber keine signifikanten Unterschiede in den plazentären mRNA- β -Catenin-Konzentrationen zwischen den beiden Gruppen.

Schlussfolgerungen Unsere Ergebnisse weisen auf abnormale plazentäre mRNA-*DKK1*-Konzentrationen bei Patientinnen mit Präeklampsie hin. Dazu kommt noch, dass ein schlechterer Verlauf der Präeklampsie mit verminderten DKK1-Konzentrationen im mütterlichen Serum assoziiert ist. Dies weist darauf hin, dass eine abnormale Wnt/ β -Catenin-Signalisierung einen plausiblen Mechanismus bei der Pathogenität von Präeklampsie darstellen könnte. Die Dysregulierung der Expression von DKK1 auf der genetischen Ebene in der Plazenta, nicht aber auf der Ebene des Proteins im mütterlichen Serum könnte die These untermauern, dass Präeklampsie eine Art plazentabedingte Erkrankung ist.

Background

Preeclampsia (PE) is a major cause of maternal and perinatal morbidity and mortality, affecting 2–8% of all pregnancies worldwide [1]. PE is characterized by hypertension, proteinuria and other organ dysfunction, occurring at any time after the 20th week of pregnancy [2]. Different theories have been proposed to explain the PE etiology. Abnormal implantation, aberrant trophoblastic invasion of spiral arteries of the myometrium, which causes deficient vascular placental development are the most discussed pathophysiology of PE [3]. However, the trigger for poor placental development and the subsequent cascade of events is multifactorial and still not well understood [4].

It has been shown that placentation is regulated by different factors including multiple signaling pathways such as Wnt/ β -catenin cascade [5]. Wnt/ β -catenin signaling is evolutionarily conserved. It controls a wide range of developmental processes. Activation of this cascade is initiated when a Wnt ligand protein binds to Frizzled receptors and their co-receptors (LRP5/6) resulting in transmitting the signal into the cytoplasm. This signal leads to β -catenin stabilization and its cytoplasmic accumulation. The accumulated β -catenin is translocated into the nucleus to activate the transcription of numerous genes controlling cell cycle, differentiation and invasion such as c-myc and cyclin D1 [6]. Dickkopf (DKK) family proteins are extracellular regulators of Wnt/ β -catenin signaling. DKK1 is a secreted protein that has been shown to modulate Wnt/ β -catenin pathway activity via degradation of cytosolic β -catenin and reduction of nuclear β -catenin-dependent gene transcription [7, 8].

Wnt/ β -catenin signaling is involved in endometrial hemostasis and normal uterine function [9–12], as well as in different gestational disorders such as aberrant trophoblastic invasion and placental organoid [13–14]. Wnt ligands and DKK1 were found to be expressed in endometrial cells in a menstrual cycle-dependent manner [15]. In addition, Sonderegger and colleagues iden-

tified different expression pattern of many Wnt ligands and frizzled receptors in human placenta and its different trophoblast subtypes. Their results suggested that several different Wnt signalling pathways could be active in human placenta depending on the cell-specific expression and distribution of individual Wnts and FZD receptors [16].

Abnormal Wnt/ β -catenin signaling revealed to be involved in the pathogenesis of various diseases including cancer [17]. Trophoblasts share some biological characteristics with cancer cells such as migration and invasion, yet in a highly organized and precisely controlled spatiotemporal manner [18]. Therefore, the role of Wnt/ β -catenin signalling in trophoblast development has been investigated and results proved the involvement of this cascade in the etiology of PE [19].

Aberrant Wnt/ β -catenin signaling could serve as a pathologic mechanism for PE inception and progression. This study was designed to explore the role of DKK1 as a negative regulator of Wnt/ β -catenin signaling in PE. Therefore, we have investigated the expression levels of secreted DKK1 and placental mRNA levels of *DKK1* and β -catenin in women with PE comparing to women with normal pregnancy.

Material and Methods

Ethical approval

This study was carried out at the Department of Gynecology and Reproductive Medicine of University Medical School of Saarland in Germany. Our Study was approved by the local Ethic Committee of the Medical Association of the Saarland (Decision number 249/13) and was in accordance with the Declaration of Helsinki. All study participants were properly informed and provided a written consent prior to their participation.

Study cases

We designed a prospective cohort study and included 60 pregnant women. The patient collective was divided into two equal sub-collectives: 30 women with PE and 30 women with normal pregnancy (NP) as a control group.

Diagnosis of PE, definition of PE severity, HELLP syndrome and intrauterine growth retardation (IUGR) determination were assessed following the German national guideline for hypertensive diseases during pregnancy [20]. PE was defined as blood pressure $\geq 140/90$ mmHg with proteinuria > 300 mg/24 h occurring after completing 20th pregnancy week. Severe PE is defined as PE with the presence of at least one of the following criteria: blood pressure $\geq 160/110$ mmHg, progressive renal insufficiency (creatinine ≥ 0.9 mg/dl), impaired liver function (elevated transaminases, upper abdominal pain), thrombocytopenia, pulmonary edema, central nervous system dysfunction abnormalities (severe headache, blurred vision), or IUGR. IUGR was determined by the presence of fetal estimated weight < 10 th percentile and pathological Doppler of the umbilical and/or uterine artery. HELLP syndrome represents a type of PE with severe features in which elevated liver enzymes (transaminases elevation > 35 U/l), low platelet count (thrombocytes $< 100\,000/\mu\text{l}$) and hemolysis (low serum haptoglobin [≤ 25 mg/dl] or lactate dehydrogenase [LDH] ≥ 2 times the upper level of normal range [0–262 U/l]) were the predominant features. PE patients with gestational age ≥ 34 weeks were classified as late onset PE.

The control group consisted of age-matched women without PE symptoms according to the German national guideline for hypertensive diseases during pregnancy [20].

All participants were older than 18 years. The exclusion criteria included known chronic systemic diseases (such as cardiac, renal or lung disease), inflammatory or infectious disease, gestational diabetes mellitus or type 2 diabetes mellitus, and gestations complicated by chromosomal or fetal anomalies. Giving birth ≥ 37 gestational week was defined as term birth; delivery < 37 weeks was considered as preterm birth.

Clinical data and laboratory test results were collected from each patient's electronic medical record using Viewpoint version 5, SAP, and GE Voluson E8 Ultrasound systems.

Sample collection

Blood samples were collected at the time of delivery. A total of 5 ml venous blood was drawn in S-Monovette serum-gel tubes (Sarstedt, Germany). The samples were centrifuged at 1000 g for 10 min at 4 °C. The supernatant serum was transferred in aliquots to 1.5 ml Eppendorf tubes and frozen at -80 °C until analysed.

Placental tissues were obtained within 15 min after delivery. Samples (approximately 1 cm³ each) were dissected from three different sites of the maternal side under aseptic conditions. Biopsy cores were snap frozen in liquid nitrogen and further stored at -80 °C until analysis.

Serum analysis

Maternal secreted levels of DKK1 were measured using a sandwich enzyme-linked immunosorbent assay (ELISA) kit (Human DKK1 Quantikine ELISA Kit, catalog no. DKK100B, R & D Systems, Germany). Measurement procedures were done following the manu-

facturer's instruction manual. All measurements were performed in duplicate. Optical density was measured at 450 nm and referenced to 570 nm on a 96-well microplate reader (Sunrise-Tecan, Life Science) and protein levels were obtained with a four-parameter logistic curve fitted against a standard curve and multiplied by the dilution factor using Magellan 7.2 Ink Data Analysis Software (Life Science-Tecan).

Tissue analysis

RNA extraction and cDNA synthesis

A maximum amount of 20–25 mg frozen placenta tissues were distributed and homogenized in 300 μl of RNeasy lysis buffer (Qiagen, Valencia, CA, USA) using TissueLyser LT Adapter and stainless steel beads from Qiagen. RNA extraction was performed according to the manufacturer's instructions using RNeasy MiniKit (Qiagen, Valencia, CA, USA). Thereafter, Ambion TURBO DNA-free DNase kit (Life Technologies GmbH, Darmstadt, Germany) was used to remove contaminating DNA and to subsequently remove the DNase and divalent cations from extracted RNA samples. RNA purity and concentration were measured using Thermo Scientific NanoDrop 2000. In addition, further detecting of RNA integrity was done using Agilent RNA 6000 Nano Reagents Part I (Agilent Technologies, Waldbronn, Germany) and Bioanalyser Agilent 2100 from Agilent Technologies. Reverse transcribed complementary DNA (cDNA) was synthesized using High Capacity cDNA Reverse Transcription Kit (Applied Biosystem, Foster City, CA, USA) as described by the manufacturer.

RT-PCR test

PCR efficiency between 90% and 100% was established to confirm that RT-PCR results will not be influenced by amplification of contaminating DNA in the RNA samples. All samples were diluted to obtain a concentration of 10 ng cDNA. Samples were tested using TaqMan Gene Expression Assays by Life Technologies (Dickkopf WNT signaling pathway inhibitor 1 [DKK1], [catalog no. Hs00183740_m1], Catenin beta1 [CTNNB1] [catalog no. Hs00355045_m1], and Actin beta [ACTB] [catalog no. Hs03929097_m1]) and Applied Biosystems 7500 Fast Real-Time PCR System. All samples were tested in triplicate and no template control (NTC) was included in each run. Results were analyzed using the 7500 Software v.2.0.5 (Life Technologies GmbH, Darmstadt, Germany). A relative quantification $2^{-\Delta\Delta C_t}$ method was used to normalize the samples with respect to the reference gene [ACTB] and obtain fold change values of PE cohort using normal pregnant women as a control group [21]. The mean value of mRNA transcripts of each participant was set as one for each tested gene.

Statistical analysis

Data were analyzed using SPSS version 21 (IBM, Armonk, NY, USA). Median and range were used for the descriptive analysis of non-parametric continuous variables and absolute and relative frequencies were used for the categorical variables. The null hypothesis was tested against its alternative using either non-parametric Mann-Whitney-U-Test for continuous variables or χ^2 Test for categorical variables. Correlations between two continuous variables were evaluated using the non-parametric Spearman cor-

► **Table 1** Clinical characteristics of study subjects.

Characteristic	Women with NP (n = 30)	Women with PE (n = 28)	p-value
Maternal age (years) [†]	32 (23–42)	32 (24–42)	NS
Gestational age at sampling (weeks) [†]	38 + 3 ([37 + 1]–[39 + 4])	35 + 3 ([26 + 4]–[40 + 2])	< 0.0001
Gravida ^{*#}	Primigravida: n = 5 Secundigravida: n = 13 Multigravida n = 12	Primigravida: n = 19 Secundigravida: n = 6 Multigravida n = 3	< 0.0001
Para ^{*#}	Nulliparous: n = 0 Primiparous: n = 6 Multiparous: n = 24	Nulliparous: n = 1 Primiparous: n = 21 Multiparous: n = 6	< 0.0001
BMI (kg/m ²) [†]	31.8 (20.2–47.9)	33.8 (24.8–58.1)	0.028
Systolic blood pressure (mmHg) [†]	112 (85–144)	147 (126–240)	< 0.0001
Diastolic blood pressure (mmHg) [†]	66 (50–89)	90 (69–120)	< 0.0001
Urine protein (mg/24 h) [†]	0	800 (180–9020) [Ⓟ]	< 0.0001
Symptoms ^{*,**}	Absent: n = 15 Existent: n = 15	Absent: n = 9 Existent: n = 19	NS
HELLP syndrome (n) (%) [‡]	0 (0%)	5 (18%)	0.021
Birthweight (g) [†]	3375 (2135–4550)	2290 (480–3880)	< 0.0001
Preterm vs. term infants [*]	Preterm: n = 0 Term: n = 30	Preterm: n = 17 Term: n = 11	< 0.0001
1-min Apgar [*]	7–10: n = 28 < 7: n = 2	7–10: n = 19 < 7: n = 8	0.013
5-min Apgar [*]	7–10: n = 30 < 7: n = 0	7–10: n = 26 < 7: n = 1	NS
10-min Apgar [*]	7–10: n = 30 < 7: n = 0	7–10: n = 26 < 7: n = 1	NS
IUGR neonates, n (%) [‡]	0 (0%)	9 (32%)	0.001

Data present the number of gravida and para of study cases excluding the investigated delivery. ** Existence of one or more of the following signs: headache, severe edema, vaginal bleeding, epigastric pain, and vision changes. † Results are shown as (Median [Range]) and p-value is calculated using Mann-Whitney-U-Test. * Results are shown as categories and p-value is calculated using χ^2 test. ‡ Results are shown as number (%) and p-value is calculated using χ^2 test. Ⓟ Data were available from 25 PE cases. n: Number of cases. NS: Not significant.

relation coefficient and the corresponding test of the null hypothesis that the correlation coefficient is equal to zero. All statistical tests were performed at a two-sided comparison-wise significance level of 5%. As appropriate for exploratory studies no adjustments for multiple testing was performed.

Results

The data of two PE patients had to be removed from the analysis after withdrawal of their consent forms. Clinical characteristics of study participants are summarized in ► **Table 1**. Briefly, cases with NP had decreased BMI and increased gravidity and parity rate compared with PE cases. Moreover, PE was associated with higher incidence of HELLP syndrome and poorer birth outcomes including lower birthweight, preterm birth, and IUGR in 32% (9/28) of included cases. In addition, PE cases with symptoms included 14 cases with edema, 2 cases with epigastric pain, 2 cases with headache, 1 case with headache and edema and 1 case with headache and epigastric pain. On the other hand, NP cases included 15 women with mild edema and 15 women with no symptom.

Maternal serum DKK1 levels

DKK1 levels did not correlate with maternal age or BMI in both study groups and in the complete study cohort (all $p > 0.05$). We found no significant difference in maternal serum DKK1 levels between PE patients and women with NP (Mean [Average]: 2320 [985–4161] pg/ml vs. [1070–4717] pg/ml respectively, $p > 0.05$) (► **Fig. 1**). Similar observations were made comparing maternal serum DKK1 levels in PE cases and NP cases of matched gestational age (≥ 35 weeks) (Mean [Range]: 2344 [1764–3470] pg/ml in 15 PE cases vs. 2403 [1070–4717] pg/ml in 30 NP cases, $p > 0.05$). However, we found that PE women diagnosed with one or more symptoms (headache, severe edema, vaginal bleeding, epigastric pain, and vision changes) had decreased maternal serum DKK1 levels compared with PE cases with no clinical symptoms (► **Fig. 2a**). NP women showed no differences in maternal serum DKK1 levels comparing cases diagnosed with one or more symptoms and cases with no symptom (► **Fig. 2b**).

We also correlated between serum DKK1 levels and PE features (► **Table 2**). Decreased serum DKK1 levels were associated with HELLP syndrome diagnosis (Mean [Range]: 1710 [985–2565] pg/

ml in five cases with HELLP vs. 2361 [1644–4161] pg/ml in 23 cases without HELLP, $p = 0.027$). In addition, cases with IUGR neonates had lower serum DKK1 levels compared with cases with normal IUG neonates (Mean [Range]: 1884 [985–2872] pg/ml vs. 2377 [1764–4161] pg/ml respectively, $p = 0.027$). We found no significant correlation between serum DKK1 levels and PE severity or onset (► **Table 2**).

Placental mRNA levels of *DKK1* and β -catenin

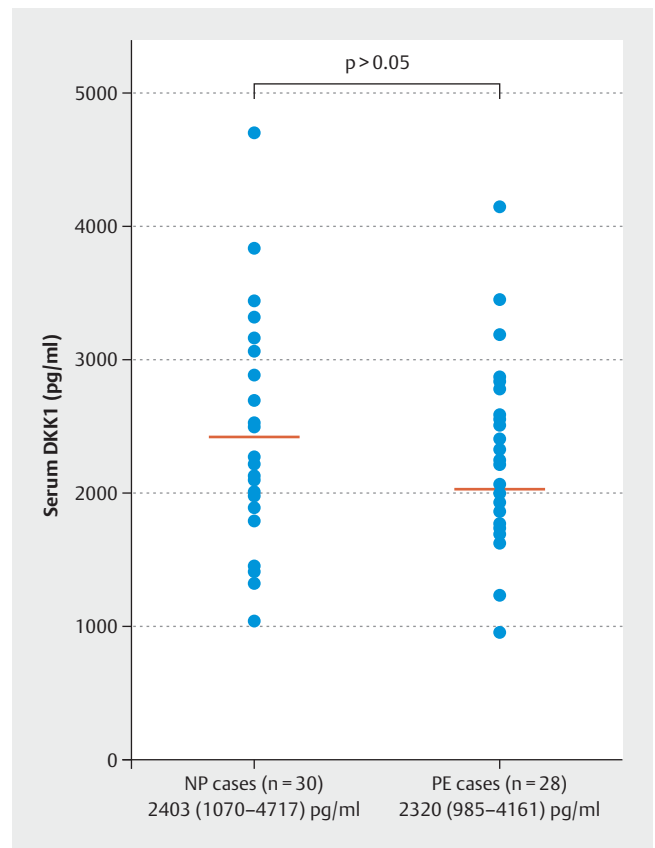
Our RT-PCR results showed that PE cases had significantly lower placental *DKK1* mRNA levels compared with NP cases (Mean [Range] ΔC_t : 6.29 [3.48–7.71] vs. 5.43 [2.67–7.54] respectively, $p = 0.033$) (► **Fig. 3 a**). Using $2^{-\Delta\Delta C_t}$ equation, this reduction was estimated by 38%. By comparing *DKK1* mRNA levels in PE cases and NP cases of matched gestational age (≥ 35 weeks), we reported also that PE cases had significantly lower placental *DKK1* mRNA levels compared with NP cases (Mean [Range] ΔC_t : 6.37 [3.48–7.71] in 15 PE cases vs. 5.43 [2.67–7.54] in 30 NP cases, $p = 0.046$).

In addition, placental β -catenin mRNA levels in PE cases were also lower than values in NP cases, but the difference was not statistically significant (Mean [Range] ΔC_t : 0.9 [– 2.36–1.83] vs. 0.62 [0.01–1.89] respectively, $p = 0.054$) (► **Fig. 3 b**).

Moreover, we tested the correlation of placental *DKK1* mRNA levels with maternal serum DKK1 levels and with placental β -catenin mRNA levels in PE and NP cases and results reported no significant results.

Discussion

The clinical diagnostic and prognostic value of serum levels of DKK1 in human cancers is well documented in the literature [22, 23]. Up to now, however, little is known about circulating levels of DKK1 in PE. In the current study, we investigated maternal serum DKK1 levels in PE and NP women. The results revealed no significant differences between the two groups. In addition, we reported no differences between women with early-onset PE (11 cases) and women with late-onset PE (17 cases). Our results are in line with the findings of Ueland et al., who documented no significant differences in circulating DKK1 levels in a cohort consist-



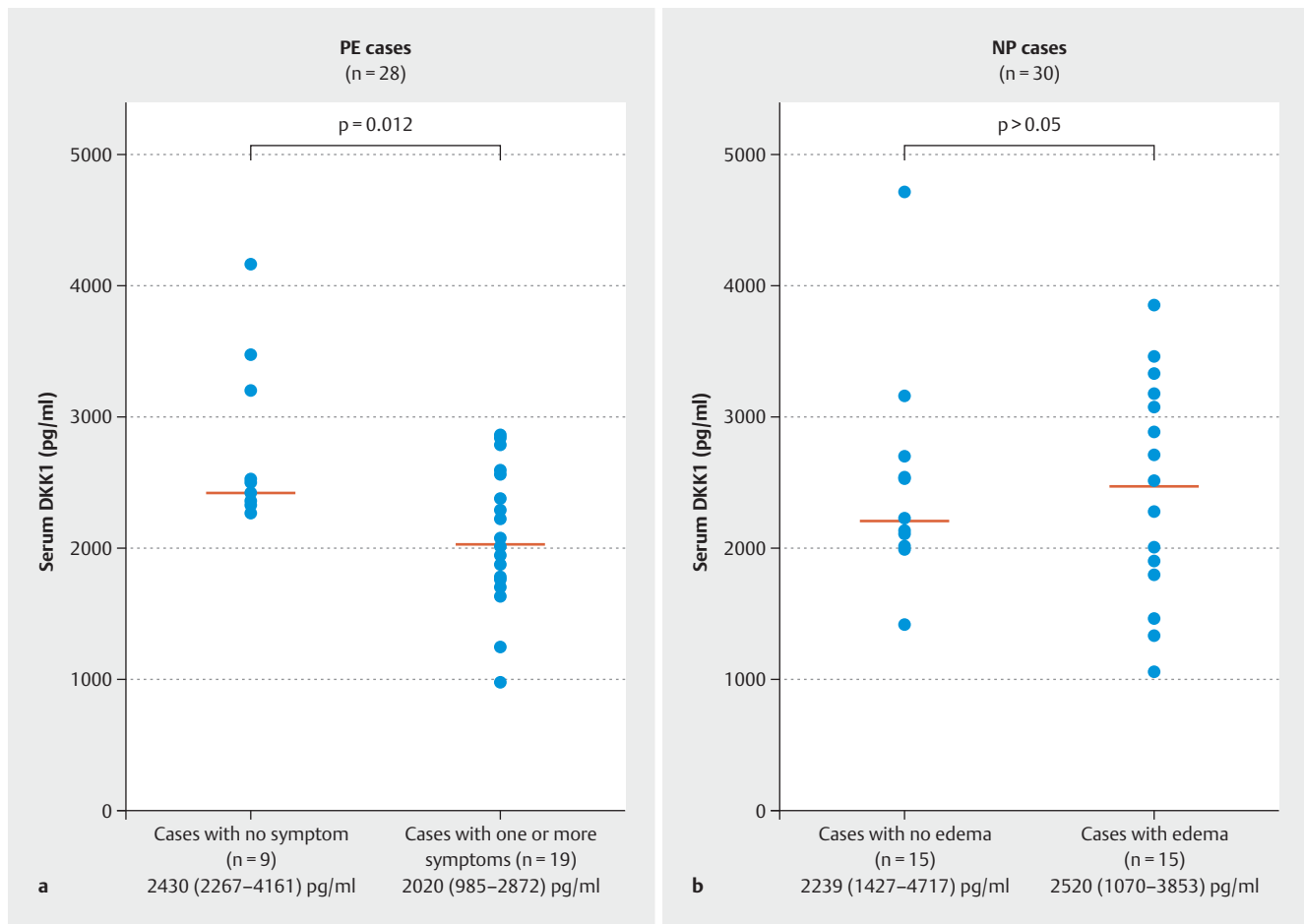
► **Fig. 1** Maternal serum DKK1 levels in NP and PE cases. No statistically significant differences in maternal serum DKK1 levels were found between PE cases and NP cases. Results are presented as Mean (Range). p -value is tested using Mann-Whitney-U-test. n: number of cases.

ing of 61 women with NP and 34 women with PE at term (weeks 35–36) [24]. On the contrary, Tayyar and colleagues showed increased serum DKK1 levels in women with early-onset PE (27 cases) compared with NP women (27 cases). Nevertheless, late-onset PE women (26 cases) showed no differences compared with

► **Table 2** Correlation of maternal serum DKK1 levels with preeclampsia features.

Feature	Cases	Serum DKK1 (pg/ml)	p-value
PE severity	Mild PE (n = 14)* Severe PE (n = 14)	2321 (1256–3470) 2306 (985–4161)	NS
HELLP syndrome	Cases without HELLP (n = 23) Cases with HELLP (n = 5)	2361 (1644–4161) 1710 (985–2565)	0.027
PE onset	Late onset (n = 17) Early onset (n = 11)	2267 (985–3470) 2430 (1256–4161)	NS
IUGR	IUGR neonates (n = 9) Normal IUG neonates (n = 19)	1884 (985–2872) 2377 (1764–4161)	0.048

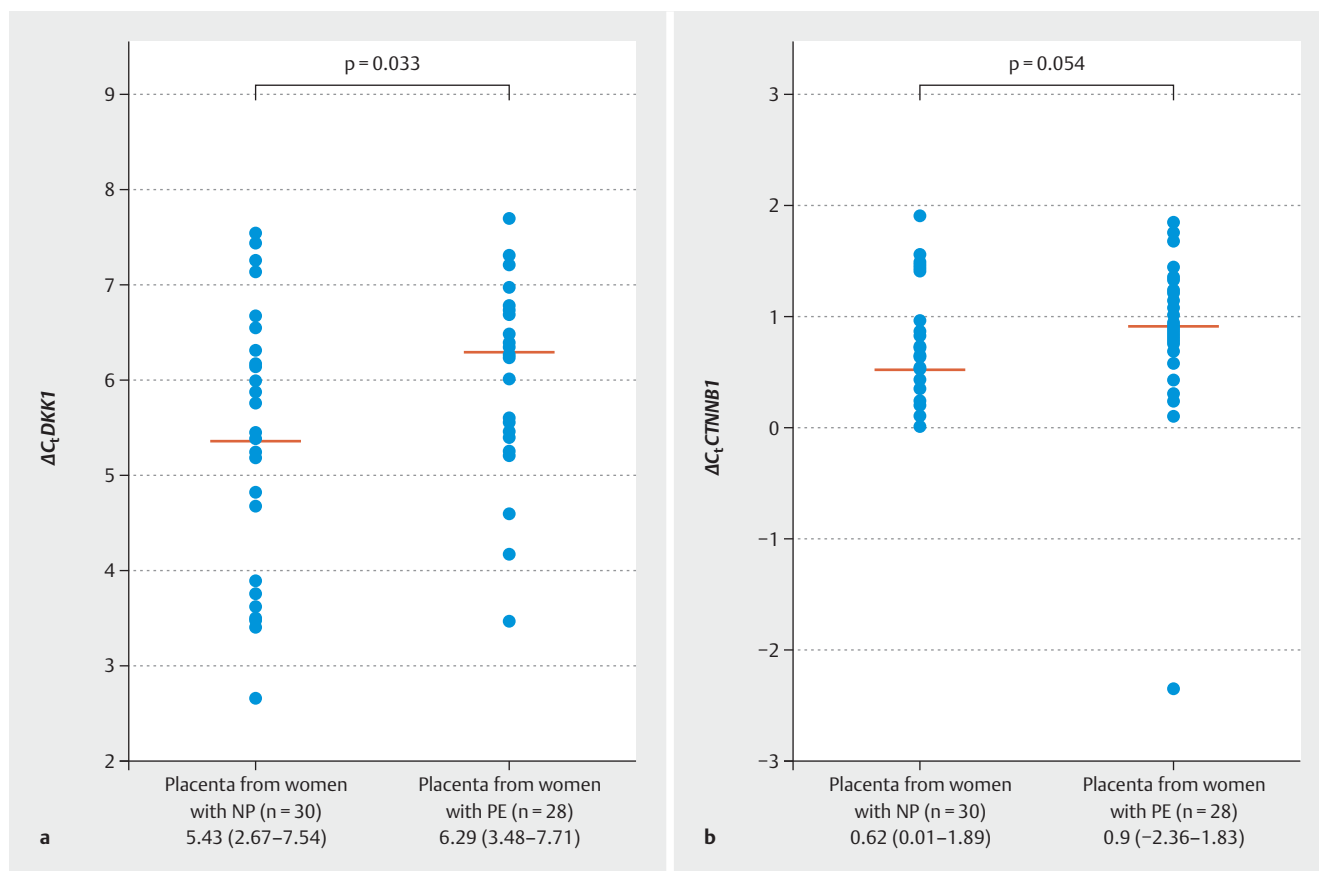
* Mild PE: diagnosis of PE with systolic blood pressure 140–160 mmHg and proteinuria < 5 g/24 h. Results are shown as (Median [Range]) and p -value is calculated using Mann-Whitney-U-Test. n: Number of cases. NS: Not significant.



► **Fig. 2** Correlation between maternal DKK1 serum levels and existence with symptoms in PE and NP women. **a** PE women with diagnosis of one or more symptoms had decreased maternal serum DKK1 levels compared with PE cases with no symptom. **b** NP women showed no differences in maternal serum DKK1 levels between cases diagnosed with edema and cases without edema. Results are presented as Mean (Range). p-value is tested using the non-parametric Mann-Whitney-U-test. n: number of cases.

NP women or early-onset PE women [25]. Moreover, their results revealed that maternal serum DKK1 levels had significant negative correlation with age but not BMI or gestational age at the time of blood sampling. Our data reported no significant correlation between serum DKK1 levels and any of these three parameters. In the study of Tayyar and colleagues, authors did not specify which cases were included for the correlation analysis between serum DKK1 levels and clinical parameters. In our study, the analysis was performed separately, for all study cases as one cohort, for NP cohort, and for PE cohort. None of these analyses revealed statistically significant correlations. Inconsistency across studies could be due to the differences in study parameters including number of tested cases, clinic-pathological characteristics of study subjects, and time of sample drawing. In addition, we assume that the notable overlapping in serum DKK1 values between tested groups enables defining a clear cut-off value of this parameter, which may in turn affect its potential as a screening biomarker in PE. An apparent dichotomy in DKK1 activity as a clinical diagnostic and prognostic biomarker in various types of cancer was also reported by different studies [26, 27].

Vascular endothelial dysfunction is responsible for the clinical signs observed in women with PE. Impaired trophoblastic invasion of spiral arteries leads to chronic placental ischemia and oxidative stress, which in turn cause fetal complications and induce release of different circulating substances resulting in defective hepatic endothelium and cerebral endothelium responsible for onset of HELLP syndrome and cerebral edema respectively [28]. In kidneys, aberrant endothelial function induces vascular hyperpermeability associated with low serum albumin causes edema [29]. DKK1 has been shown to play an important role in endothelial dysfunction in different diseases [30, 31]. Therefore, DKK1 could be linked to the clinical symptoms resulting from the endothelial dysfunction in PE. In this study, we found that the diagnosis of one or more symptoms was associated with decreased maternal serum DKK1 levels in PE women. On the other hand, NP women with mild edema showed no differences in serum DKK1 levels compared with NP women without edema. Moreover, decreased maternal serum DKK1 levels were associated with fetal complication presented by IUGR diagnosis.



► **Fig. 3** Placental gene expression levels of *DKK1* and β -catenin in NP and PE cases. **a** PE cases had significantly lower placental *DKK1* mRNA levels compared with NP cases. **b** Placental β -catenin mRNA levels in PE cases were also lower than those in NP cases but the difference was not statistically significant. Results are presented as Mean (Range). p-value is tested using the non-parametric Mann-Whitney-U-test. n: number of cases.

We also found that maternal serum DKK1 levels were changed in regard to HELLP syndrome diagnosis but not to the degree of disease severity. Women with gestational hypertension progress to mild preeclampsia and severe preeclampsia at rates of 46% and 9.6%, respectively [32]. HELLP syndrome is regarded as a more severe form of PE occurring in 10 to 20% of cases with severe PE and in 0.5 to 0.9% of all pregnancies [33]. The relationship between severe PE and HELLP syndrome remains controversial. Kinay et al. showed that severe PE and HELLP syndrome cases presented different maternal outcomes and similar perinatal outcomes at <34 and \geq 34 weeks of gestation [34]. In our study, the comparison of maternal serum DKK1 levels between severe PE and HELLP syndrome cases was not performed because of the small number of cases diagnosed with HELLP syndrome without extreme high blood pressure or elevated proteinuria concentration.

Further, we detected placental gene expression levels of *DKK1* in both study groups and found that PE cases had significantly 38% lower placental *DKK1* transcript levels compared with NP women. We also detected placental transcript levels of β -catenin in order to investigate the influence of the reduction in *DKK1* transcript levels on Wnt signal activity. We found that placental transcript levels of β -catenin in PE women were lower than levels

in NP women but the difference was not statistically significant. Few studies have examined placental gene and/or protein expression levels of *DKK1* and β -catenin in PE. Zhang and co-workers analysed *DKK1* and β -catenin expression in third trimester placentas from NP women and women with severe PE. They found that mRNA and protein expression levels of β -catenin were decreased and those of *DKK1* were increased in PE placentas compared with NP placentas [35]. These observations were later confirmed by Wang et al. Their study showed that women with severe PE had increased placental *DKK1* expression and decreased placental β -catenin expression at protein level compared with NP women. In addition, placental β -catenin mRNA levels were also reduced in PE cases [36]. Placental gene and/or protein expression levels of *DKK1* and β -catenin in PE were also detected according to disease onset. In a recent comparative study, placental mRNA *DKK1* levels were found to be significantly decreased in preterm NP women compared to term NP women and early-onset PE women. The last two groups showed no significant differences [37]. On the other hand, Na and colleagues demonstrated decreased placental protein expression of *DKK1* and increased placental protein expression of β -catenin in early-onset PE women and in late-onset PE women compared with NP women [38]. The incompatibility between the findings of all the previously mentioned studies includ-

ing ours could be due to the different timing of sample acquisition and/or to the absence of normalization using trophoblast sub-type-specific/stromal markers as placenta from women with PE may have a different composition of different trophoblast subtypes versus stroma.

Moreover, our results showed no causal relationship between gene and protein expression levels of DKK1 in both study groups. DKK1 as a secretory protein could undergo different types of posttranslational modifications in it or in its ligands, which in turn might affect its function. DKK1 is a downstream target of Wnt/ β -catenin cascade performing a regulatory effect through negative feedback loop [39]. However, we found no significant correlation between placental *DKK1* mRNA levels and placental β -catenin mRNA levels in our study cohort. This might be due, in part, to the small number of tested cases and the lack of detecting placental protein localization and expression levels of DKK1 and β -catenin. Concurrent detection of gene and protein expression levels of target molecules enable more insight into the molecular basis of Wnt/ β -catenin in the reciprocal interactions between the mother and the fetus during normal and preeclamptic pregnancy.

Conclusions

This study aimed to detect the role of DKK1 in PE. Based on a quantitative analysis of maternal serum levels and placental gene levels in PE women as well as in NP women, we conclude that DKK1 could be involved in PE pathogenicity. Dysregulated expression of DKK1 at gene level in the placenta but not at protein level in the maternal serum might confirm the notion that preeclampsia is a type of placenta-derived disease. More serious PE features were associated with decreased maternal serum DKK1 levels. However, further large-scale and multicenter studies using blood samples at different times (at pregnancy diagnosis, at PE diagnosis in affected women, and at delivery) are warranted to demonstrate the potential of serum DKK1 levels in PE.

Conflict of Interest

The authors declare that they have no conflict of interest.

References

- Townsend R, O'Brien P, Khalil A. Current best practice in the management of hypertensive disorders in pregnancy. *Integr Blood Press Control* 2016; 9: 79–94. doi:10.2147/IBPC.S77344
- Peres GM, Mariana M, Cairrão E. Pre-Eclampsia and Eclampsia: An Update on the Pharmacological Treatment Applied in Portugal. *J Cardiovasc Dev Dis Actions* 2018; 5: 3. doi:10.3390/jcdd5010003
- Meekins JW, Pijnenborg R, Hanssens M et al. A study of placental bed spiral arteries and trophoblast invasion in normal and severe pre-eclamptic pregnancies. *Br J Obstet Gynaecol* 1994; 101: 669
- Knöfler M, Haider A, Saleh L et al. Human placenta and trophoblast development: key molecular mechanisms and model systems. *Cell Mol Life Sci* 2019; 76: 3479–3496
- Knöfler M, Pollheimer J. Human placental trophoblast invasion and differentiation: a particular focus on Wnt signaling. *Front Genet* 2013. doi:10.3389/fgene.2013.00190
- Clevers H. Wnt/ β -Catenin Signaling in Development and Disease. *Cell* 2006; 127: 469–480
- Niehrs C. Function and biological roles of the Dickkopf family of Wnt modulators. *Oncogene* 2006; 25: 7469–7481
- Matoba K, Mihara E, Tamura-Kawakami K et al. Conformational freedom of the LRP6 ectodomain is regulated by N-glycosylation and the binding of the Wnt antagonist Dkk1. *Cell Rep* 2017; 18: 32–40
- Wang Y, van der Zee M, Fodde R et al. Wnt/ β -catenin and sex hormone signaling in endometrial homeostasis and cancer. *Oncotarget* 2010; 1: 674–684
- Fan X, Kriegl S, Hwang JY et al. Dynamic regulation of Wnt7a expression in the primate endometrium: implications for postmenstrual regeneration and secretory transformation. *Endocrinology* 2012; 153: 1063–1069
- Atli MO, Guzeloglu A, Dinc DA. Expression of wingless type (WNT) genes and their antagonists at mRNA levels in equine endometrium during the estrous cycle and early pregnancy. *Anim Reprod Sci* 2011; 125: 94–102
- Dunlap KA, Filant J, Hayashi K et al. Postnatal deletion of Wnt7a inhibits uterine gland morphogenesis and compromises adult fertility in mice. *Biol Reprod* 2011; 85: 386–396
- Meinhardt G, Haider S, Haslinger P et al. Wnt-dependent T-cell factor-4 controls human extravillous trophoblast motility. *Endocrinology* 2014; 155: 1908–1920
- Turco MY, Gardner L, Kay RG et al. Trophoblast organoids as a model for maternal-fetal interactions during human placentation. *Nature* 2018; 564: 263–267
- Tulac S, Overgaard MT, Hamilton AE et al. Dickkopf-1, an inhibitor of Wnt signaling, is regulated by progesterone in human endometrial stromal cells. *J Clin Endocrinol Metab* 2006; 91: 1453–1461
- Sonderregger S, Husslein H, Leisser C et al. Complex expression pattern of Wnt ligands and frizzled receptors in human placenta and its trophoblast subtypes. *Placenta* 2007; 28: S97–S102
- Ng LF, Kaur P, Bunnag N et al. WNT Signaling in Disease. *Cells* 2019; 8: 826
- Soundararajan R, Rao AJ. Trophoblast 'pseudo-tumorigenesis': Significance and contributory factors. *Reprod Biol Endocrinol* 2004; 2: 15
- Zhang Z, Wang X, Zhang L et al. Wnt/ β -catenin signaling pathway in trophoblasts and abnormal activation in preeclampsia (Review). *Mol Med Rep* 2017; 16: 1007–1013
- Stepan H, Kuse-Fohl S, Klockenbusch W et al. Diagnosis and Treatment of Hypertensive Pregnancy Disorders. Guideline of DGGG (S1-Level, AWMF Registry No. 015/018, December 2013). *Geburtshilfe Frauenheilkd* 2015; 75: 900–914
- Livak KJ, Schmittgen TD. Analysis of relative gene expression data using real-time quantitative PCR and the 2⁻(Delta Delta C(T)) Method. *Methods* 2001; 25: 402–408
- Wei R, Rodríguez RA, Mullor MDMR et al. Analyzing the prognostic value of DKK1 expression in human cancers based on bioinformatics. *Ann Transl Med* 2020; 8: 552
- Li J, Gao Y, Yue W. The Clinical Diagnostic and Prognostic Value of Dickkopf-1 in Cancer. *Cancer Manag Res* 2020; 12: 4253–4260
- Ueland T, Estensen ME, Grindheim G et al. Elevated levels of the secreted wingless agonist R-spondin 3 in preeclamptic pregnancies. *J Hypertens* 2020; 38: 1347–1354
- Tayyar AT, Karakus R, Eraslan Sahin M et al. Wnt signaling pathway in early- and late-onset preeclampsia: evaluation with Dickkopf-1 and R-Spondin-3 glycoproteins. *Arch Gynecol Obstet* 2019; 299: 1551–1556
- Xu H, Wu J, Chen B et al. Serum Dickkopf-1 (DKK1) is significantly lower in patients with lung cancer but is rapidly normalized after treatment. *Am J Transl Res* 2014; 22: 850–856
- Menezes ME, Talukdar S, Wechman SL et al. Dickkopf1: A tumor suppressor or metastasis promoter? *Adv Cancer Res* 2018; 138: 213–237

- [28] Sibai BM. Diagnosis, controversies, and management of the syndrome of hemolysis, elevated liver enzymes, and low platelet count. *Obstet Gynecol* 2004; 103: 981–991
- [29] Karumanchi SA, Maynard SE, Stillman IE et al. Preeclampsia: a renal perspective. *Kidney Int* 2005; 67: 2101–2113
- [30] Pontremoli M, Brioschi M, Baetta R et al. Identification of DKK-1 as a novel mediator of statin effects in human endothelial cells. *Sci Rep* 2018; 8: 16671
- [31] Jiang SJ, Li W, Li Y et al. Dickkopf-related protein 1 induces angiogenesis by upregulating vascular endothelial growth factor in the synovial fibroblasts of patients with temporomandibular joint disorders. *Mol Med Rep* 2015; 12: 4959–4966
- [32] Barton JR, O'Brien JM, Bergauer NK et al. Mild gestational hypertension remote from term: progression and outcome. *Am J Obstet Gynecol* 2001; 184: 979–983
- [33] Tranquilli AL, Dekker G, Magee L et al. The classification, diagnosis and management of the hypertensive disorders of pregnancy: A revised statement from the ISSHP. *Pregnancy Hypertens* 2014; 4: 97–104
- [34] Kinay T, Küçük C, Kayıkçıoğlu F et al. Severe Preeclampsia versus HELLP Syndrome: Maternal and Perinatal Outcomes at <34 and ≥34 Weeks' Gestation. *Balkan Med J* 2015; 32: 359–363
- [35] Zhang Z, Li H, Zhang L et al. Differential expression of beta-catenin and dickkopf-1 in the third trimester placentas from normal and preeclamptic pregnancies: a comparative study. *Reprod Biol Endocrinol* 2013. doi:10.1186/1477-7827-11-17
- [36] Wang X, Zhang Z, Zeng X et al. Wnt/β-catenin signaling pathway in severe preeclampsia. *J Mol Histol* 2018; 49: 317–327
- [37] Zhang L, Leng M, Li Y et al. Altered DNA methylation and transcription of *WNT2* and *DKK1* genes in placentas associated with early-onset preeclampsia. *Clin Chim Acta* 2019; 490: 154–160
- [38] Li N, Huang L, Li Y et al. Lin28B/miR-92b Promote the Proliferation, Migration, and Invasion in the Pathogenesis of Preeclampsia via the DKK1/Wnt/β-Catenin Pathway. *Reprod Sci* 2020; 27: 815–822
- [39] Niida A, Hiroko T, Kasai M et al. DKK1, a negative regulator of Wnt signaling, is a target of the β-catenin/TCF pathway. *Oncogene* 2004; 23: 8520–8526