

Ruscus Genus: A Rich Source of Bioactive Steroidal Saponins

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Key words

- *Ruscus* genus
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- chronic venous insufficiency

Abstract

▼
The genus *Ruscus* (Asparagaceae family) is native to the Mediterranean, Southern and Western Europe and is represented by perennial, rhizomatous, and evergreen shrubs. Among the approximately seven species spread throughout Europe up to Iran, *Ruscus aculeatus* L. (butcher's broom) is the most widely distributed and appreciated. This review provides an overview of the traditional use of *Ruscus* spp., the current knowledge of the chemistry of this genus, and the pharmacological studies carried out on *Ruscus* spp. extracts. The underground parts of *Ruscus* plants are a source of steroidal saponins that can be classified into two structural classes: the hexacyclic spiro-

stanol saponins and the pentacyclic furostanol saponins. The main aglycones are ruscogenin and neoruscogenin. From the pharmacological point of view, the most studied *Ruscus* species is undoubtedly *R. aculeatus*, a very ancient phlebotherapeutic agent. Pharmacological investigations since the discovery of the vasoconstrictive and venotonic properties of ruscogenin and neoruscogenin in the underground parts of *R. aculeatus* are discussed. Preparations based on *Ruscus* species are currently used for the treatment of chronic venous insufficiency, varicose veins, haemorrhoids, and orthostatic hypotension. Finally, analytical techniques for the quality control of *R. aculeatus* extracts are reported.

Botany of *Ruscus* Genus

▼
The genus *Ruscus* has shuttled between various families including Ruscaceae [1], Convallariaceae, and Liliaceae, but currently, in the APG III classification system, it has been included in the Asparagaceae family [2]. The genus is native to the Mediterranean, Southern and Western Europe [3]. The species are represented by perennial, rhizomatous, and evergreen shrubs with multiple stems arising from a creeping, thick, sympodially branched rhizome to form an oval, pyramidal bush. Stems appear striate, green, erect, and much branched, growing to 1 m tall. Leaves are reduced to triangular scarious scales up to 5 mm long and replaced functionally by rigid cladodes (flattened, leaf-like stem tissue also known as asphylloclades) 2–18 cm long and 1–8 cm broad, each arising from a leaf axil; cladodes are ovate, entire, dark green, and spine-pointed. Flowers are small, arising from the axil of a small scarious bract in the center of the upper surface of a cladode, each with a short pedicel. Perianth is greenish-white, approximately 3 mm long, and made

up of two whorls of three segments bearing papillae. Some species are monoecious while others are dioecious. In the latter, male and female plants are very similar in appearance; female flowers show a cup formed from fused stamen filaments around the superior, unilocular ovary, which has a subsessile capitate stigma, while male flowers possess three stamens and green or violet filaments fused into a tube around an undeveloped ovary. Fruits are bright red globose berries of 8–14 mm in diameter with 1–4 large seeds [2]. The genus includes approximately seven species spread throughout Europe up to Iran (Yeo 1968) including *Ruscus aculeatus* L., *Ruscus colchicus* Yeo, *Ruscus hypoglossum* L., *Ruscus hypophyllum* L., *Ruscus hyrcanus* Woronow, *Ruscus x microglossus* Bertol., and *Ruscus streptophyllum* Yeo. *R. aculeatus* is known with the common English name “butcher's broom”. Jekyll described the derivation of its name as “In country places where it abounds, butchers use the twigs tied in bunches to brush the little chips of meat off their great chopping-blocks, that are made of solid sections of elm trees, standing three and half feet high

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and about two and half feet across" (● Fig. 1). The common Italian name *pungitopo* and the German *Mausedorn* signify "mouse-stinger" and are related with the old practice to put the well-armed branches around stored food to protect it from pests [4]. The plant has also a nutritional use in the Mediterranean countries, where the young shoots are eaten like asparagus spears [5], and the seeds were once used to make a coffee substitute.

R. aculeatus is the most widely distributed *Ruscus* species, while *R. hypoglossum*, *R. hypophyllum* and *R. x microglossus* are representatives of the Mediterranean area. *R. streptophyllum*, a rare species, is endemic of Maidera, and *R. colchicus* and *R. hyrcanus* are endemic of Caucasus and Azerbaijan, respectively [4].

R. hypophyllum presents the greater morphological variation within the genus. Its distribution range is throughout the western Mediterranean region, principally Northern Africa as far east as Tunisia. *R. hypoglossum* represents a gentler version of *R. aculeatus* for the presence of larger and softer cladodes. It can be distinguished by its large foliaceous inflorescence bracts. Its Italian common name *bislingua*, meaning "double-tongue", is due to one tongue being the bract and the other the cladode. *R. x microglossus* is probably a hybrid between *R. hypoglossum* and *R. hypophyllum* [4].

Ruscus ponticus Woronow is a synonym of *R. aculeatus* L. and is widespread in Crimea and Caucasus, particularly in the forests of West and East Georgia [6]. *R. ponticus* is well known in this country for the preparation of ruscoptonin, obtained from the underground parts of the plant [6].

Traditional Medicine

Although the aerial parts of *Ruscus* species are edible, the rhizomes and roots of the plants are used in traditional medicine as phytotherapeutic products. During the Middle Ages, the young shoots of *R. aculeatus* were not only used as food, but also as a medicinal agent for the treatment of heavy legs, urinary disorders, and abdominal pain [7]. The hydroalcoholic extract of *R. aculeatus* rhizomes is traditionally used as a vascular preventive and tonic in preparations for disorders involving the venous system, including venous fragility and varicose veins [8,9]. The underground parts of *R. aculeatus* have been also used as diuretic and anti-inflammatory agents, as well as for the treatment of hemorrhoids and atherosclerosis. As a remedy for diseases of the circulatory system, *R. aculeatus* has a long tradition of proven success in Europe. A written record of its use as a phlebotherapeutic agent dates back at least 2000 years. In the middle of the 20th century, the main active substances, the steroidal saponins ruscoegenin and neoruscoegenin, were identified [7]. Indeed, *R. aculeatus* preparations are widely distributed in Europe, and have been used for more than 40 years to treat chronic venous insufficiency and vasculitis [10,11].

In European tradition, both the aerial parts (leaves) and the rhizome of *R. aculeatus* are considered to be diuretic and mildly laxative [12]. In the same way, in the folk medicine of Turkey, a decoction of the roots of *R. aculeatus* is widely used internally as a diuretic and for the treatment of urinary system disorders, such as nephritis, and towards eczema as well as kidney stones [13].

In Palestinian folk medicine, the rhizome extract is used externally against skin diseases [14], while in central Italy it is used in the treatment of warts and chilblains [15]. In some parts of Italy the same preparation is used for colitis and diarrhea [16]. It is also used locally against inflammation and arthritis [16].



Fig. 1 *R. aculeatus* L. (Color figure available online only.)

Not only rhizomes but also other parts of the plants have been used in traditional medicines. Indeed, the aerial parts of *R. aculeatus* are traditionally used as diuretics, mostly in Mediterranean and Middle East countries [16]. In Turkey, the decoction of berries of *R. hypoglossum* is applied externally for boils and warts, while its fresh leaves are used for livestock against cold and mastitis [16]. The leaves of *R. colchicus* are used by the local population for feeding livestock in order to yield more milk and to increase its fat content [17].

In Iran, *R. hyrcanus* is traditionally used as a diuretic, appetizer, antilaxative, vasoconstrictor, antibleeding, antinephritis, anti-infection, aperient, antivaricose, and laxative remedy [18].

Chemistry

The underground parts of *Ruscus* plants are a source of steroidal saponins. Steroidal saponins consist of a C27 skeleton, typically an oxidized cholesterol derivative, bearing varying numbers of sugar residues at different positions. The steroidal saponins isolated from *Ruscus* genus can be classified into two structural classes, both of them bearing a sugar chain linked mostly to C-1: the hexacyclic spirostanol saponins, which possess a bicyclic ketal at C-22 and the pentacyclic furostanol saponins, which usually contain a hemiketal function at C-22, and a glycosidic linkage (typically to a single β -D-glucose residue) at C-26 [19].

The diol aglycones (25*R*)-spirost-5-en-1 β ,3 β -diol, named ruscoegenin (1), and spirost-5,25(27)-dien-1 β ,3 β -diol, named neoruscoegenin (2), were first isolated from the subterranean parts of *R. aculeatus* and described in 1955–1957 [20]. These aglycones and derived spiro- and furostanol glycosides were subsequently also isolated from other species of the genus.

Moreover, a series of cholestane glycosides have been isolated only from *R. colchicus*, *R. hypoglossum*, and *R. hypophyllum* [21–23], and few examples of pregnane glycosides have been reported in *R. aculeatus*, *R. hypoglossum*, and *R. ponticus* [6,24].

Generally, the fresh underground parts of the plant material were extracted with methanol, and sometimes the obtained crude extract was partitioned between water and *n*-butanol. The fractionation of the selected extract was performed by a combination of column chromatographic methods over a porous polymer resin (Diaion HP-20), silica gel and octadecylsilanized (ODS) silica gel, Sephadex LH-20, or by droplet countercurrent chromatography (DCCC) as well as HPLC.

The first characterization of the structures of deglucoside and ruscoside was obtained in 1971 by acid and enzymatic hydrolysis and degradation studies. Successively, the structure elucidation of steroidal saponins isolated from the *Ruscus* genus was obtained by spectroscopic methods including 1D- (^1H , ^{13}C and TOCSY) and 2D-NMR (DQF-COSY, HSQC, HMQC, HMBC, NOESY and ROESY) experiments as well as ESIMS and HRMS analyses. Acid hydrolysis of saponins was carried out to afford the sugar moieties, which were successively identified by GC-MS.

Spirostanol saponins from *Ruscus* spp.

The spirostanol saponins contain ruscogenin (1) and neoruscogenin (2) as aglycones (Fig. 2). The structure of ruscogenin was compared with that of the typical plant sapogenin (25*R*)-spirost-5-en-3 β -ol, also known as diosgenin [25], revealing that it differed from that of diosgenin only for the occurrence of a β -hydroxy group at C-1, being identical for all other structural features, including orientation of the C-3 oxygen atom (3 β -equatorial) and ring junctions (B/C *trans*, C/D *trans*, D/E *cis*) [25]. Three saponins named ruscin (also known as ruscoponticosides D) (3), desglucoscin (also known as ruscoponticosides C) (4), and desglucodesrhamnoscin (5), containing neoruscogenin as the aglycone, have been firstly described in 1971 from *R. aculeatus* [26]. These compounds, described among the main constituents, are accompanied by the corresponding (25*R*)-25,27-dihydro derivatives, of which the respective structures are 6, 7, and 8 (Fig. 2). Like all of the spirostanol prosapogenins, all of these take origin from furostanol saponins carrying a glucose residue at C-26.

Since 1971, from the underground parts of *R. aculeatus*, several spirostane derivatives have been isolated (9–28) (Fig. 3) [25, 27–32]. All isolated neoruscogenin derivatives are characterized by the presence of a sugar chain only at C-1. Among these, compounds 9–19 possess an α -L-rhamnopyranosyl-(1 \rightarrow 2)- α -L-arabinopyranosyl sugar chain at C-1. Substitution with acetyl groups, sulphate ions, and 2-hydroxy-3-methylpentanoyl moieties are present on the sugar units. Hydroxylation at positions 23 and 24 of the spiro system (15–19) [29, 32] and also the presence of a glucopyranosyl unit at C-23 (15–16) can occur [29].

Aculeoside A (17) represents an unusual bisdesmosidic spirostanol saponin, having 6-deoxy-D-glycero-L-threo-4-hexosulose linked to the C-24 hydroxyl group of the aglycone. Aculeoside A was active as a cyclic AMP phosphodiesterase inhibitor [30]. Other unusual structures are spilacleosides A and B (18 and 19) having spiro 1,3-dioxolan-4-one structures, made up of aculeoside A and (2*R*,3*S*)-2,3-dihydroxy-3-methylpentanoic acid [32]. These were reported as rare examples of natural compounds containing a spiro structure with unusual features: they are the first examples where both segments intramolecularly construct a 1,3-dioxolan-4-one ring with a spiro system at C-2 and where the naturally derived 2,3-dihydroxy-3-methylpentanoic acid has the opposite configuration at C-3 with respect to the same acid produced by a mutant strain of *Neurospora crassa* and by *Heliotropium strigosum* showing a (2*R*,3*R*) configuration [32]. Few natural products intramolecularly incorporating a spiro 1,3-dioxolan-4-one ring are known and the usefulness of this moiety as a building block in asymmetric organic synthesis has been often reported [32]. The structural peculiarities of aculeoside B (20) are the presence of three acetyl esters at the inner galactose unit and the glucosyloxy group attached to C-23 of the aglycone [31]. Other examples of ruscogenin derivatives isolated from the underground parts of *R. aculeatus* are represented by a saponin

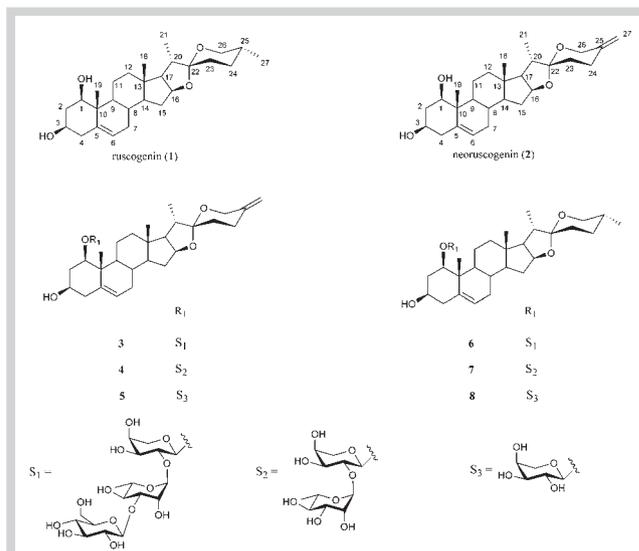


Fig. 2 The aglycones ruscogenin (1) and neoruscogenin (2) and their derived saponins (3–8).

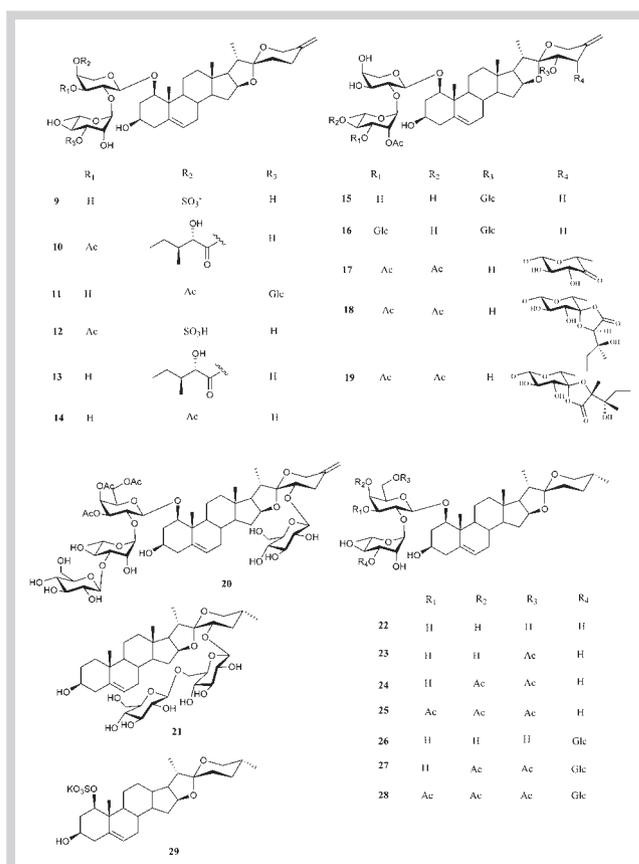


Fig. 3 Spirostanol saponins isolated from *R. aculeatus*.

having a diglycoside moiety at C-23 of the aglycone (21) [25] and by a series of ruscogenin glycosides characterized by a diglycoside moiety made up of rhamnose and galactose units linked to C-1, with the presence of an additional glucose unit or acetyl groups on the sugar chain (22–28) [33]. A sulphated derivative

at C-1 of ruscogenin (**29**) has been isolated from *R. aculeatus* rhizomes [34].

From the rhizomes of *R. colchicus* and *R. hypoglossum*, the two sapogenins **1** and **2** along with the spirostane saponins **3–8** were isolated [19].

An HPLC-ESIMSⁿ method, based on high-performance liquid chromatography coupled to electrospray positive ionization multistage ion trap mass spectrometry, has been used as an effective tool to rapidly identify and guide the isolation of saponins from the leaves of *R. colchicus*, among which there are four spirostanol derivatives (**30–33**) [17] (● Fig. 4).

From the cladophylls of *R. hypoglossum*, four neoruscogenin derivatives (**34–37**) have been isolated [35] (● Fig. 4). Another six compounds (**38–43**), some of which characterized by the presence of a primary alcoholic function at C-27 (**41–42**), and **44** have been isolated from the roots and rhizomes of *R. hypoglossum* (● Fig. 4) [21, 36].

The first phytochemical investigation on the rhizomes of *R. hypophyllum* was performed in 1971 with the isolation and identification of diosgenin and the two isomers (25*R* and 25*S*) of ruscogenin [36]. In the last years ruscoponticoside C (**4**), two spirostanol derivatives (**22–23**) [22], and five spirostanol derivatives {(**45–47**) [22] and (**48, 49**) [23]} have been detected in the rhizomes of *R. hypophyllum* (● Fig. 5).

A phytochemical investigation of the roots of *R. ponticus* led to the isolation of diosgenin and neoruscogenin (**1**) [37], along with ruscoponticosides C (**4**) and D (**3**) [38].

The ¹³C NMR data of the aglycone moieties of spirostanol derivatives **3, 15, 21, 22, 29, 30, and 45–47** are reported in ● Table 1.

Furostanol saponins from *Ruscus* spp.

Pentacyclic furostanol saponins can be divided into those that possess either a hydroxy or methoxy moiety at C-22 and those that possess a $\Delta^{20(22)}$ -unsaturation. Two furostanol saponins, containing a double bond in the C25–C27 position, have been described in *R. aculeatus*, desglucoruscoside (also known as ruscoponticoside E) (**50**), and ruscocide (**51**). The corresponding (25*R*)-25,27-dihydro derivatives (**52** and **53**) are also present as minor constituents [19].

From the underground parts of *R. aculeatus*, several furostane derivatives have been isolated (**54–62**) [39], (**63–70**) [27], (**71–75**) [29] (● Figs. 6 and 7). Also, sulphated compounds **76–79** have been identified [34, 40]. As regards to these compounds, Oulad-Ali et al. reported the isolation of compound **76**, which is constitutionally identical to compound **79** [34] (● Fig. 7). The stereochemistry at C-22 was left unassigned, but comparison between the ¹³C-NMR data of the two compounds evidenced some small but not insignificant differences, pointing to a stereoisomeric relationship [40]. Moreover, De Marino et al. described the isolation of a furostanol saponin with a double bond between C20–C22 (**80**), reported for the first time in the underground parts of *R. ponticus* [40] (● Fig. 7).

Rhizomes of *R. colchicus* and *R. hypoglossum* were found to contain the furostanol saponins **50–53**, along with compound **81** [19] (● Fig. 8).

From the leaves of *R. colchicus*, ruscoponticoside E (**50**) and ceparoside B (**52**) along with compounds **82–96** [17] were identified (● Fig. 8).

Furostanol compounds **50, 51, 54, 63, 80, and 97–98** were isolated from *R. ponticus* underground parts, and compounds **99–107** from *R. ponticus* leaves [6] (● Fig. 9).

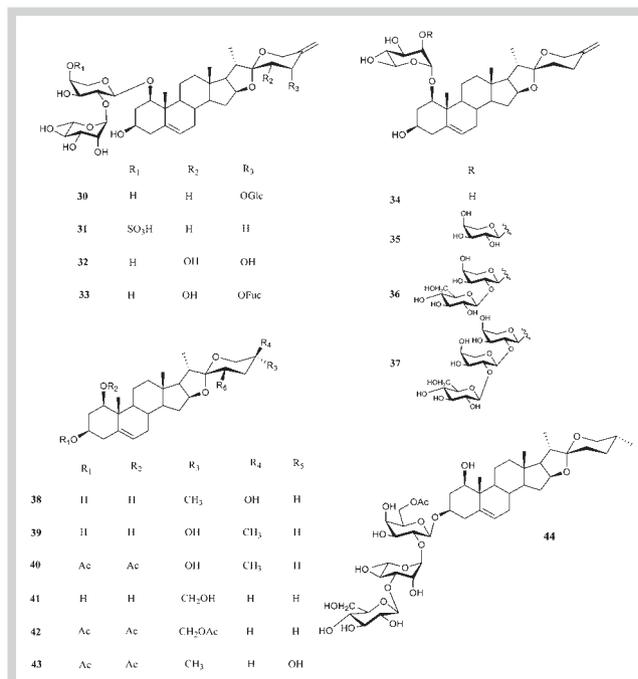


Fig. 4 Spirostanol saponins isolated from *R. hypoglossum* (**30–44**).

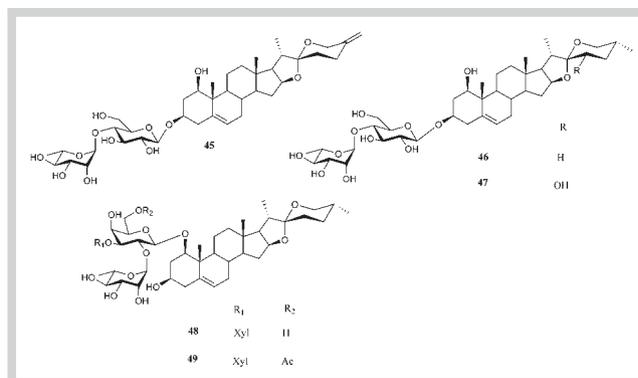


Fig. 5 Spirostanol saponins isolated from *R. hypophyllum* (**45–49**).

The ¹³C NMR data of the aglycone moieties of furostanol derivatives **55, 56, 61, 77–80, 82–85, 95, 96, 98, and 100** are reported in ● Table 2.

Stereochemistry of furostanol and spirostanol saponins

In both furostanol and spirostanol saponins, C-25 is found naturally with either the *R* or *S* configuration. Further structural diversity is generated in this class of natural products by differences in the stereochemistry of C-22 and in the *cis* or *trans* fusion of the steroid A and B rings as well as differences in the steroid hydroxylation and glycosylation patterns. This structural variation may account for the wide range of bioactivities reported for steroidal saponins [41].

By NMR spectroscopy, the stereochemistry of C-25 in spirostanol saponins may be determined thanks to the proton vicinal coupling constants between H-24 and H-26 as well as by comparison of the ¹H and ¹³C NMR chemical shifts of the steroid F ring with literature values [42–44]. Infrared spectroscopy has been used for

Table 1 ^{13}C NMR data of the aglycone moieties of spirostanol derivatives **3**, **15**, **21**, **22**, **29**, **30**, and **45–47**.

	3 ^a [27]	15 ^b [29]	21 ^a [25]	22 ^a [33]	29 ^a [34]	30 ^c [17]	32 ^a [79]	45 ^a [22]	46 ^a [80]	47 ^a [22]
1	84.8	84.5	37.8	84.2	84.1	84.6	83.8	77.6	37.3	37.4
2	38.1	37.4	32.6	38.0	39.8	37.1	37.3	40.8	30.0	30.2
3	68.2	68.2	71.2	68.1	68.0	69.1	68.1	74.9	78.2	78.8
4	43.8	43.9	43.5	43.8	40.1	43.3	43.6	39.6	39.1	39.3
5	139.6	139.5	141.9	139.5	139.0	139.8	139.3	139.0	140.7	140.8
6	124.8	124.7	121.1	124.7	125.2	126.0	124.6	125.0	121.6	121.8
7	32.0	32.2	32.4	31.9	33.0	32.5	31.8	32.1	32.0	32.2
8	33.1	33.0	31.6	33.0	31.9	33.8	32.8	32.7	31.5	31.6
9	50.5	50.4	50.4	50.5	49.8	51.4	50.3	51.0	50.1	50.2
10	43.0	42.9	37.0	42.8	43.7	43.5	42.7	43.5	36.9	37.0
11	23.9	23.9	21.2	24.0	23.6	24.6	23.9	24.0	21.0	21.1
12	40.3	40.7	40.4	40.4	41.9	41.2	40.4	40.3	40.0	39.9
13	40.2	40.8	41.0	40.1	40.6	41.5	40.5	40.1	40.9	40.5
14	56.9	56.8	56.7	57.1	56.7	57.9	56.7	56.7	56.5	56.6
15	32.4	32.1	32.0	32.4	32.8	32.5	32.1	32.2	32.1	31.8
16	81.5	81.8	81.3	81.1	81.1	83.0	83.1	81.3	81.5	81.1
17	63.0	62.3	62.1	62.9	63.1	63.0	61.2	63.0	62.3	62.9
18	16.7	17.3	17.3	16.8	16.6	16.6	16.7	16.4	16.4	16.4
19	15.1	15.0	19.5	15.0	14.8	15.1	14.9	13.6	19.2	19.4
20	41.8	35.7	35.7	41.9	43.1	37.0	36.9	41.7	35.6	42.0
21	15.0	14.6	14.7	14.9	15.0	14.1	14.5	14.9	14.6	15.0
22	109.5	110.7	110.8	109.2	109.2	113.8	112.5	109.3	111.6	109.3
23	33.2	77.2	76.2	31.7	31.9	32.5	69.5	33.1	67.3	31.8
24	29.0	37.7	37.6	29.2	29.2	73.2	74.0	28.8	38.6	29.3
25	144.5	143.9	31.4	30.6	30.7	147.0	146.1	144.3	31.4	30.6
26	65.0	64.1	65.8	66.8	66.8	64.3	60.6	64.9	65.8	66.9
27	108.6	109.8	16.8	17.2	17.3	107.8	112.4	108.6	16.8	17.3

^a Pyridine- d_5 ; ^b pyridine- d_5 - CD_3OD (11:1); ^c CD_3OD

distinguishing between spirostanol and furostanol saponins, as the former possesses characteristic absorption bands at around 980, 920, 900, and 860 cm^{-1} . The relative intensities of the 920 and 900 cm^{-1} bands are predictive of the C-25 configuration in spirostanols (920 > 900 in 25S; 900 > 920 in 25R) [41,45].

The determination of C-25 stereochemistry is more challenging in furostanol saponins, which do not possess the rigid bicyclic system present in their spirostanol analogues. There are reports in which the intensities of IR absorption bands have been used to assign the C-25 configuration in furostanols [46,47], as well as in spirostanols, but there is no basis for the former application [48,49]. The most reliable method for determination of the C-25 configuration in furostanol saponins is conversion to the corresponding spirostanol form, either through enzymatic cleavage of the C-26 glucose residue specifically to effect ring closure of the side chain [50] or by complete hydrolysis to yield the free spirostanol aglycone.

In most of the reported furostanol saponins, the absolute configuration of C-25 has been assigned directly by NMR spectroscopy. Empirically, it has been found that both the chemical shifts and resolution of the geminal proton resonances of H_2 -26 (δ_a and δ_b , $\Delta_{ab} = \delta_b - \delta_a$) are dependent on the configuration of C-25. The resolution of the H_2 -26 resonances is normally used to assign C-25 stereochemistry, as these signals are more separated in 25S ($\Delta_{ab} \geq 0.57$ ppm) than 25R furostanols ($\Delta_{ab} \leq 0.48$ ppm) [48,49]. In addition, the chemical shift of H_3 -27 occurs at a slightly higher field in 25R (δ_{H} 0.98–1.03 ppm) compared with 25S saponins (δ_{H} 1.01–1.05 ppm). The initial reports gave examples in both pyridine- d_5 and methanol- d_4 [48,49], and this predictor of C-25 stereochemistry has since been widely applied in both of these solvents [41]. Challinor et al. observed that the chemical shifts of

H_2 -26 are, in general, more resolved in 25S than 25R compounds, and that this empirical rule in different solvents led to conflicting assignments of stereochemistry. An experimental survey revealed that, while the chemical shifts of H_2 -26 exhibit a dependence on C-25 configuration, it is less pronounced in methanol- d_4 than in pyridine- d_5 solvent, and thus the general rule derived for pyridine- d_5 fails when NMR spectra are acquired in methanol- d_4 . A modified empirical method for the direct assignment of C-25 stereochemistry in furostanol saponins in methanol- d_4 ($\Delta_{ab} = 0.45$ – 0.48 ppm for 25S; $\Delta_{ab} = 0.33$ – 0.35 ppm for 25R) has been provided [41]. Therefore, the stereochemistry of C-25 of furostanol saponins isolated before this modified empirical method was deduced could be reversed.

Other steroidal saponins from *Ruscus* spp.

Cholestane glycosides have been reported in *R. hypophyllum*, *R. hypoglossum* and *R. colchicus*. The first report was compound **108** in *R. hypoglossum* [21]. The six cholestane saponins **109–113** [22] and **114** have been isolated from *R. hypophyllum* [23]. Moreover, a sulphated cholestane glycoside (**115**) has been reported in *R. colchicus* [17] (● Fig. 10).

In 1972, two pregnane glycosides (**116** and **117**) were identified in the rhizomes of *R. aculeatus* [51]. Compound **116** was also isolated from *R. hypoglossum* [24], while more recently compounds **117** and **118** were identified in the underground parts of *R. ponticus* [6] (● Fig. 10). Compound **118** represents an example of pregnane with the presence of a five-membered lactone ring between C-22 and C-16. These kinds of lactone-type saponins and their glycosides might be biosynthetically derived from the genuine 23,26-oxygenated spirostane or furostane type [6]. The

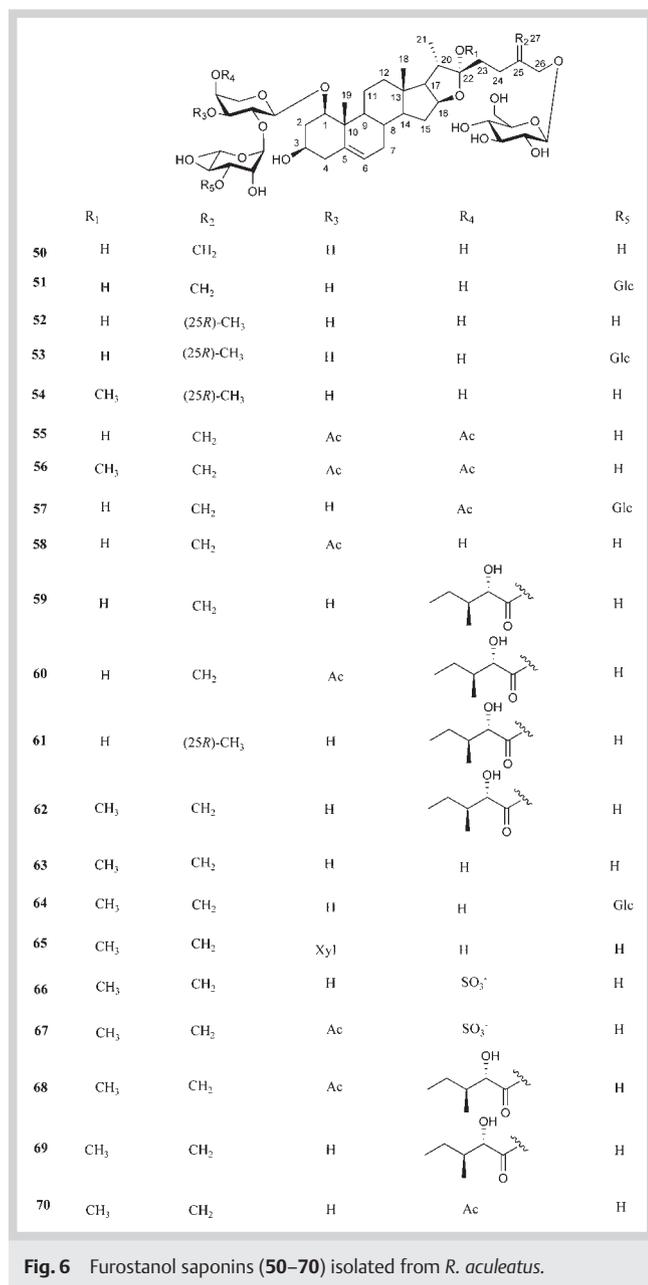


Fig. 6 Furostanol saponins (50–70) isolated from *R. aculeatus*.

¹³C NMR data of the aglycone moieties of compounds **109**, **111**, **112**, **115**, and **118** are reported in **Table 3**.

Pharmacological Activity

From the pharmacological point of view, the most studied *Ruscus* species is undoubtedly *R. aculeatus*, a very ancient phlebotherapeutic agent. Since the discovery of the vasoconstrictive and venotonic properties of ruscogenin and neoruscogenin, *R. aculeatus* L. was extensively used, especially in Germany and France, for the treatment of chronic venous insufficiency, varicose veins, haemorrhoids, and orthostatic hypotension [7].

In 1972, sapogenins isolated from *R. aculeatus* displayed anti-inflammatory activity on rat paw edema, but were inactive on rat capillary fragility. Vasoconstrictor effects were also observed on the vessels of isolated rabbit ears, but only ruscogenin derivatives

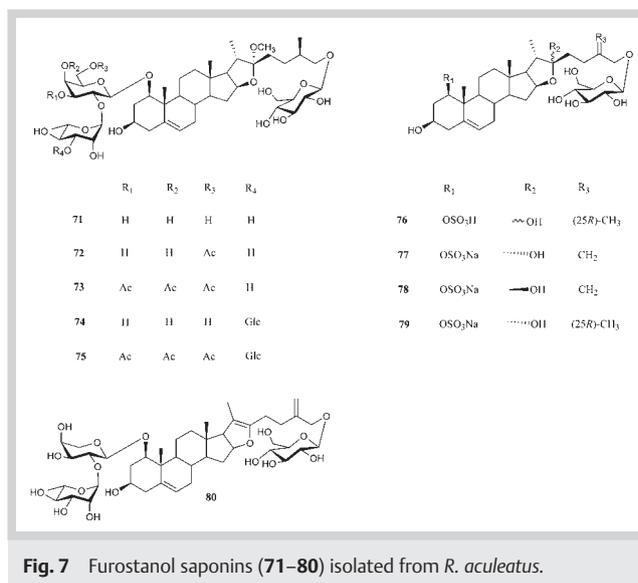


Fig. 7 Furostanol saponins (71–80) isolated from *R. aculeatus*.

significantly decreased capillary permeability in the rabbit. The acute oral and parenteral toxicities were low in mice and rats, and prolonged oral administration of high doses was well tolerated in rats [52].

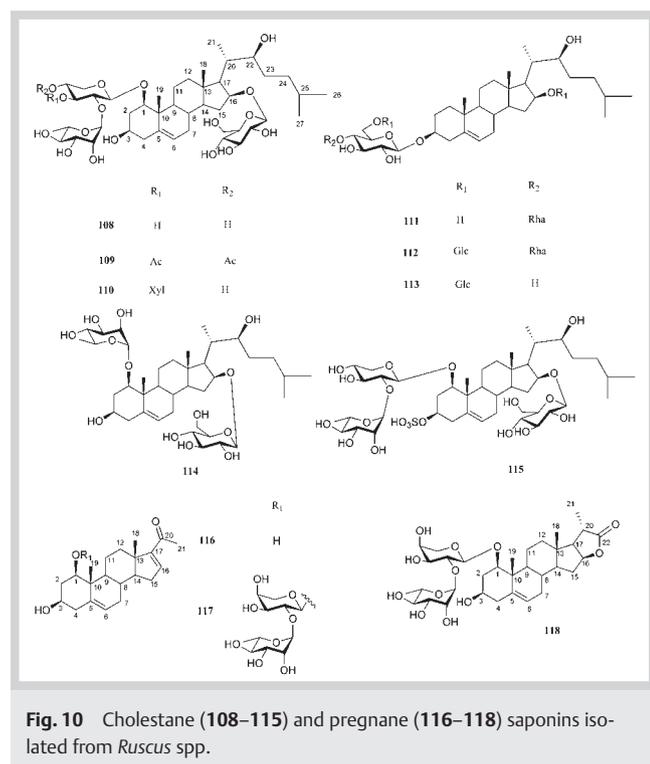
Rudofsky reported a decrease of about 10% in venous capacity 2 h after oral administration of *Ruscus* hydroalcoholic extract in healthy volunteers. Patients with chronic venous insufficiency treated with *Ruscus* extract maintained constant venous tone and had improved venous emptying, unlike placebo-treated patients [53].

In isolated cutaneous veins, *Ruscus* extract caused contractions owing to activation of postjunctional α -1 and α -2 adrenergic receptors by releasing endogenous norepinephrine (NE) from adrenergic nerve endings and through a direct action on the venous smooth muscle [54,55]. The contractions caused by the extract were increased by local warming and reduced by cooling [9,56]. Intravenous administration of *Ruscus* extract (5 mg/kg) was also tested in the microcirculation *in vivo*, investigating its effects on the diameter of arterioles and venules of the hamster cheek pouch and the influence of temperature on the observed effects [8]. Moreover, the effects of *Ruscus* extract and the flavonoid hesperidine methylchalcone were investigated in the hamster cheek pouch on increased microvascular permeability induced by various agents, such as bradykinin, histamine, and leukotriene B₄.

A study aimed at evaluating the factors contributing to the efficacy of *R. aculeatus* in the treatment of venous insufficiency showed that ruscogenins were ineffective on hyaluronidase activity, but exhibited remarkable anti-elastase activity [57]. Their anti-edematous effects have also been demonstrated [58]. Another study reported a comparison of the effects of *R. aculeatus* extract and norepinephrine on the contractions in veins. Contractions to norepinephrine were greater in control veins than in varicose tributaries. Contractions to the extract were greater in varicose tributaries than in greater saphenous veins from varicose patients. Contractions to norepinephrine were reduced similarly by α -1 and α -2-adrenergic agonists in control and varicose veins, but to a greater extent by α -2-blockade in greater saphenous veins from varicose patients. Contractions to *Ruscus* extract were not reduced by α -1-adrenergic blockade in control veins, but were reduced by α -2-adrenergic blockade in varicose veins.

Table 2 ^{13}C NMR data (CD_3OD) of the aglycone moieties of furostanol derivatives **55**, **56**, **61**, **77–80**, **82–85**, **95**, **96**, **98**, and **100**.

	55 [40]	56 [40]	61 [39]	77 [40]	78 [40]	79 [40]	80 [81]	82 [17]	83 [17]	84 [17]	85 [17]	95 [17]	96 [17]	98 [6]	100 [6]
1	83.9	83.7	84.2	85.5	85.4	85.6	83.5	78.8	78.9	78.8	78.8	83.8	83.9	84.1	38.3
2	36.8	37.0	37.2	38.8	38.7	38.8	37.4	40.4	40.4	40.3	40.2	34.4	34.4	37.3	30.5
3	68.6	69.0	68.9	68.6	68.6	68.7	68.2	75.9	75.8	76.0	75.7	75.9	75.9	69.0	79.7
4	43.0	43.2	43.2	43.2	43.2	43.0	43.9	39.7	39.9	39.8	39.7	40.6	40.5	43.2	39.6
5	139.2	139.2	139.3	138.7	139.0	138.8	139.6	139.0	138.7	139.0	138.8	139.0	138.9	139.4	142.1
6	125.4	125.6	125.5	126.5	126.4	126.5	124.7	126.0	126.2	126.1	126.0	126.5	126.5	125.7	122.5
7	32.2	32.6	32.5	32.8	33.0	32.6	32.1	32.6	32.7	32.5	32.6	32.5	32.5	32.6	32.9
8	33.6	33.8	33.9	33.8	33.8	33.8	32.9	33.6	33.6	33.6	33.7	33.4	33.5	33.8	32.5
9	50.9	51.1	51.0	50.6	50.7	50.7	50.4	52.0	51.8	51.2	51.7	51.0	51.1	51.2	51.6
10	43.5	43.4	43.0	43.7	43.6	43.7	43.1	43.7	43.6	43.7	43.8	43.2	43.1	43.3	38.0
11	24.7	25.0	24.9	24.0	24.0	24.0	24.2	24.3	24.5	24.2	24.6	24.0	24.1	24.6	21.8
12	40.7	41.1	41.0	40.8	40.9	40.9	40.2	40.7	40.9	40.8	41.0	40.9	40.9	41.0	40.7
13	41.6	41.4	40.8	41.2	41.4	41.4	42.9	41.0	41.2	41.0	41.3	41.0	41.0	44.1	43.4
14	57.2	57.5	57.6	57.4	57.5	57.5	55.1	57.8	57.6	57.9	57.7	57.8	57.7	56.2	56.2
15	32.5	32.6	32.6	32.7	32.6	32.7	34.7	32.8	32.8	32.7	32.8	32.6	32.5	34.9	34.9
16	81.8	82.4	82.4	81.8	82.2	82.0	84.5	82.3	82.4	82.1	82.3	82.3	82.3	85.2	85.2
17	63.7	65.0	64.9	64.0	64.1	64.1	64.6	65.2	65.3	65.2	65.3	64.8	65.1	65.5	65.5
18	16.7	17.1	16.6	16.8	16.9	16.9	14.5	16.6	16.8	16.5	16.7	16.7	16.7	14.6	14.6
19	14.9	15.1	14.7	14.5	14.5	14.6	15.1	13.3	13.2	13.4	13.2	14.7	14.7	15.1	19.6
20	40.4	41.0	40.9	40.8	40.9	41.1	104.2	40.8	41.0	40.8	41.0	41.0	41.0	105.5	105.3
21	15.4	15.9	15.7	15.8	16.0	16.1	11.8	15.8	15.7	15.7	15.8	16.3	15.8	11.5	11.6
22	111.7	113.3	113.9	111.1	113.7	111.3	151.5	113.3	113.2	113.0	113.2	113.3	113.3	152.7	152.7
23	37.4	32.1	31.3	37.5	37.3	31.4	31.1	32.0	31.1	32.1	31.1	31.1	32.5	24.0	23.8
24	28.3	28.5	28.9	28.5	28.4	28.6	24.7	24.8	28.8	28.4	28.7	28.8	28.4	31.3	31.1
25	147.4	147.1	147.1	147.0	147.3	147.3	146.3	146.5	146.5	146.6	146.6	146.8	146.8	146.8	146.8
26	72.4	72.4	75.9	72.6	72.4	75.6	71.7	72.5	75.2	72.4	75.3	75.2	72.4	75.7	75.7
27	112.1	112.2	16.9	111.9	111.9	17.1	11.6	112.0	17.0	112.1	17.1	17.0	112.0	17.1	17.1

**Fig. 10** Cholestane (**108–115**) and pregnane (**116–118**) saponins isolated from *Ruscus* spp.

dependent reduction in hyperpermeability to 71.8% at 100 μM [62].

Studies dealing with the pharmacological properties of *R. aculeatus* extracts showed that compounds exhibiting these properties were the steroidal glycosides ruscin and ruscoside and their hydrolysis products desglucoruscin, desglucodesrhamnoruscin, and desglucoruscoside. The pharmacological activity seems to increase with the decrease of the number of the sugar residues [63]. For these reasons, a number of methods for enzymatic or chemical hydrolysis of *R. aculeatus* have been developed. A bioconversion process to produce the monoglycoside desglucodesrhamnoruscin from dry extracts of the rhizome of *R. aculeatus* has been developed using enzyme preparations containing a β -glucopyranosidase and an α -rhamnopyranosidase [63]. Furthermore, aculeoside A exhibited inhibitory activity on cell growth of leukemia HL-60 cells with an IC_{50} value of 0.48 $\mu\text{g}\cdot\text{mL}^{-1}$, while aculeoside B was inactive [31]. Also compounds **4**, **5**, **9–11**, **22–28**, and **63–75** were tested for their cytotoxic activity against HL-60 cells. Among the tested compounds, **10**, **25**, **68** and **73** showed an IC_{50} ranging from 3.0 μM to 3.7 μM [27,33].

Formulations Containing *Ruscus aculeatus* Extracts

A number of products based on *R. aculeatus* acting on the venous system have been developed over the last 50 years. Recent studies have also demonstrated the efficacy of a combination of Rusci rhizome with hesperidin methylchalcone and ascorbic acid against vascular disease [64,65]. Indeed, some products have proven to be highly successful, such as Cyclo 3 Fort (Fabroven) and Phlebodril [66].

In particular, Cyclo 3 Fort is a formulation of the root extract of *R. aculeatus* (150 mg per capsule), hesperidin methyl chalcone

	109 ^a [22]	111 ^a [22]	112 ^a [22]	115 ^a [17]	118 ^c [6]
1	84.1	37.4	37.3	84.3	84.3
2	37.0	30.1	30.1	34.6	37.1
3	67.9	78.2	78.2	76.0	68.8
4	43.5	39.2	39.2	40.7	43.2
5	138.9	140.9	140.7	138.7	139.4
6	125.0	121.8	121.9	127.2	125.3
7	31.6	32.1	32.0	32.5	32.4
8	33.1	31.8	31.6	34.0	33.8
9	50.2	50.4	50.3	51.3	51.2
10	42.6	36.9	36.9	43.0	43.2
11	24.0	21.0	21.0	24.6	24.0
12	40.5	40.3	39.9	41.5	39.1
13	42.0	42.9	42.3	42.8	41.9
14	51.1	54.7	55.0	56.4	55.7
15	37.1	37.6	36.8	37.8	34.0
16	82.5	71.3	82.5	83.2	84.5
17	57.9	59.6	57.8	59.1	60.1
18	13.7	13.3	13.4	13.7	14.3
19	14.8	19.3	19.3	14.9	14.9
20	35.8	37.4	35.8	36.2	37.5
21	12.5	13.4	12.5	11.8	17.9
22	73.0	73.2	73.0	74.5	184.1
23	33.6	29.9	33.7	33.0	
24	36.6	36.4	36.6	37.0	
25	28.8	28.4	28.8	29.4	
26	22.9	22.6	22.9	23.3	
27	23.0	23.2	23.0	23.3	

^a Pyridine-d₅ ^c CD₃OD

Table 3 ¹³C NMR data of the aglycone moieties of cholestane derivatives **109**, **111**, **112**, **115**, and of pregnane derivative **118**.

(150 mg), and ascorbic acid (100 mg) used to increase the venous tone in patients with venous disease. Studies of the effects of oral Cyclo 3 Fort on capillary permeability were carried out in hamsters with moderate diabetes and compared with hamsters treated with a placebo, showing that oral administration of Cyclo 3 Fort inhibited histamine-induced plasma exudation in hamsters with mild diabetes without affecting the glycemia [67]. Another study on the effects of Cyclo 3 Fort in cheek pouch preparations from diabetic hamsters showed that the venoarteriolar reflex, studied by measuring the internal diameter of arterioles during venular occlusion, was reversed by treatment with Cyclo 3 Fort, 10 and 50 mg/kg [68].

Janssens showed the effect of venotropic drugs, including Cyclo 3 Fort, on the respiratory activity of isolated mitochondria and in endothelial cells. The results showed that Cyclo 3 Fort protected human endothelial cells against the hypoxia-induced decrease in ATP content. In addition, it induced a concentration-dependent increase in respiratory control ratio (RCR) of liver mitochondria preincubated with the drug for 60 min. Cyclo 3 inhibited the carbonyl cyanide *m*-chlorophenyl hydrazone (*m*CCP)-induced uncoupling of mitochondrial respiration. The results suggested that the protective effects on mitochondrial respiration activity by Cyclo 3 Fort may explain its protective effect on the cellular ATP content in ischemic conditions and some of its beneficial therapeutic effect in chronic vascular diseases [69].

Nonsteroidal anti-inflammatory drugs such as Cyclo 3 Fort are associated with lymphocytic colitis. The mechanisms include immunological activation or attenuated immunological defences [70].

Clinical Studies



In a key study, Rauwald and Janssen demonstrated that steroidal glycosides from *Ruscus rhizoma* are absorbed and can be detected in their unmodified form in human plasma after oral administration [71]. A study showed the treatment of secondary lymphedema of the upper limb with Cyclo 3 Fort or placebo according to a double-blind protocol. With Cyclo 3 Fort, the reduction in the volume of arm edema, the main assessment criteria, was 12.9% after 3 months of treatment compared with a placebo ($p = 0.009$). Decreased edema tended to be more marked in the forearm compared with the upper arm where an excess of fat deposition seemed to dominate over the excess of fluid accumulation. Cyclo 3 Fort was well tolerated with minimal adverse reaction [58].

Other clinical studies showed the efficacy and safety of Cyclo 3 Fort versus hydroxyethyl rutoside in chronic venous lymphatic insufficiency [72]. The comparative efficacy of a single daily dose of two capsules of Cyclo 3 Fort in the morning versus a repeated dose of one capsule in the morning and another at noon was evaluated [73]. Thus, a single dose of two capsules of Cyclo 3 Fort in the morning appeared as effective and as well tolerated as a twice daily dose of one capsule of Cyclo 3 Fort in the morning and at noon [73]. Formulations such as Cyclo 3 Fort have been tested in clinical trials for their action on the symptoms of chronic venous insufficiency (CVI), leg pain, and edema. The effects on venous distensibility, rheological disorders, the prevention of wall dystrophy, and action on the microcirculation were examined [66]. A meta-analysis of clinical trials of Cyclo 3 Fort in the treatment of CVI demonstrated the clinical efficacy of Cyclo 3 Fort in treating patients with CVI, for which it significantly reduced the severity of the symptoms compared to the placebo. An open-label

clinical trial conducted in 65 women with CEAP class C2 s to C3 s showed that the improvement in subjective functional signs under treatment with Cyclo 3 Fort was correlated with objective plethysmographic parameter improvement.

As its efficacy has been demonstrated in several clinical studies, the ESCOP (European Scientific Cooperative on Phytotherapy) recommends a daily intake of *Ruscus rhizoma* corresponding to a dose of 7–11 mg of ruscogenins (ESCOP, 2003) [65].

Analytical Techniques for the Quality Control of *Ruscus aculeatus* Extracts

Frequently, medicinal herbs are commercialized as food supplements. Safety, quality, and composition assessments of food supplements based on botanical ingredients are of major concern, as they have usually not been checked through a rigorous testing process as required for the approval of therapeutic phytopreparations [74]. The variability in the content and concentrations of constituents of plant material, together with the range of extraction techniques and processing steps used by different manufacturers, results in marked variability in the quality of commercially available herbal products. Thus, quality control of herbal products is needed to ensure their consistency, safety, and efficacy [75]. *R. aculeatus* appears in a great number of dietary supplement patents [39], and several preparations are on the market as food supplements. As cited above, the mixture of the two spirostane aglycones, neoruscogenin and ruscogenin, is considered the active ingredient of *R. aculeatus* commercial drugs [19]. Several methods based on the properties of saponins, such as hemolytic and surface activity, have been used in the past for the quantitative determination of saponins in *Ruscus* extracts. These methods have been replaced by TLC, using a number of visualization reagents, and, more recently, by HPLC [18]. Detection of ruscogenin in 5 *Ruscus* taxa in Turkey by ultra-performance liquid chromatography (UPLC) was performed [13].

The detection of steroidal glycosides using UV is well known for its insensitivity because of the absence of chromophoric groups. Thereby, liquid chromatography coupled to mass spectrometry (HPLC-ESIMS) methods may work well for the identification and quantification of the steroidal saponins. HPLC-ESIMS is one of the most powerful techniques in phytochemical analysis because of its high sensitivity and specificity. HPLC-ESIMS provides structural information by analyzing the fragmentation patterns produced in HPLC-ESI/MSⁿ experiments.

de Combarieu et al. developed an HPLC-UV method for the analysis of saponins in *R. aculeatus*, and, moreover, the saponins were identified by HPLC-ESI-MS by TurbolonSpray on a single quadrupole mass spectrometer [19].

An HPLC-ESIMS analytical method based on the positive ionization mode by atmospheric pressure chemical ionization (APCI) on an ion trap instrument operating in the MSMS scan mode was developed [76]. Kite et al. observed that the use of methanol as the mobile phase under acidic conditions determined furostanol saponins hydroxylated at C-22 to chromatograph as broad peaks, while the use of methanol as mobile phases without the addition of acid determined the elution of furostanol saponins with a good peak shape, but each C-22 hydroxylated furostanol saponin was accompanied by a second chromatographic peak identified as its C-22 methyl ether. The C-22 methyl ether of deglucoscoside was found to convert to deglucoscoside during chromatography in acidified aqueous acetonitrile, or by dissolv-

ing in water. An LC/MS/MS method was developed for the quantification of ruscogenin and neoruscogenin in hydrolyzed extracts of *R. aculeatus*. The detection was performed in the multiple reaction monitoring mode using an ion trap mass spectrometer with an electrospray ionization source operating in the positive ionization mode [77].

A study of *Ruscoven* gocce (drops), a preparation of *Hamamelis virginiana* and *Vitis vinifera* (hydroalcoholic leaf extract), *R. aculeatus* (freeze-dried root extract), *Centella asiatica* (freeze-dried leaf extract), and *Ginkgo biloba* (freeze-dried leaf extract) used to improve functionality of blood microcircles, has been carried out by electrospray mass spectrometry analysis, suggesting that either ESI(+) or ESI(-) spectra, coupled to a multivariate analysis, leads to a satisfactory and quick characterization of complex mixtures of herbal-based products [78].

More recently, to allow a rapid analysis of *R. aculeatus* species utilized as ingredients of food supplements, as well as to identify and guide the isolation of target saponins, a study of the saponin profile of *R. aculeatus* rhizomes was carried out by employing an analytical method based on high-performance liquid chromatography coupled to electrospray positive ionization tandem ion trap mass spectrometry (HPL-ESI/ITMS²), so improving the reported HPLC-ESIMS methods available [39]. In this study, a semi-preparative reversed-phase HPLC-UV approach, resulting in being able to achieve a good chromatographic separation for *Ruscus* saponins, was also proposed as an alternative and suitable analytical method for the purification of *Ruscus* steroidal saponins.

An efficient multi-targeted method based on liquid chromatography coupled with a hybrid triple quadrupole linear ion trap analyzer was developed to screen selected botanicals, among which *R. aculeatus* was characterized by means of its appropriate biomarker ruscin [74].

Conclusion

R. aculeatus (butcher's broom) represents the best-known and appreciated *Ruscus* species. Since the Middle Ages, the young shoots of *R. aculeatus* were used as medicinal agents for disorders involving the venous system, anticipating the proven success of its hydroalcoholic extract in Europe. Steroidal saponins represent the main class of chemical compounds isolated from rhizomes and roots of *R. aculeatus* and are considered to be the active compounds of *R. aculeatus* commercial products, in particular, a mixture of two spirostane aglycones, neoruscogenin and ruscogenin. A chemical investigation revealed the presence of steroidal saponins also in other *Ruscus* spp. Nevertheless, the scientific production focused mostly on *R. aculeatus*. A biological evaluation of *R. aculeatus* extracts were mainly focused on investigating and exploring deeper its use for venous diseases. Besides these investigations, a few other reports on the biological activity of *R. aculeatus* extracts or pure isolated compounds have been carried out so far.

Due to its benefits demonstrated in several clinical studies, oral supplementation with *R. aculeatus* is recommended. So, *R. aculeatus* appears in several commercial herbal products. Thanks to the innovation in analytical technology, identification, and detection of chemical markers in *R. aculeatus* extracts, food supplements have been strongly improved. In particular, hyphenated techniques based on HPLC coupled to MS analysis have proved to be the most suitable for the rapid identification of com-

pounds in the plant matrix. Moreover, taking into account that not only the main compounds but also the low-concentration compounds can affect the sample quality, fingerprint analysis might be a simple and rapid way for the identification and quality control of *R. aculeatus* commercial preparations.

Conflict of Interest

The authors declare no conflict of interest.

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