Cystic pancreatic lesions: Can we diagnose them accurately what to look for? FNA marker molecular analysis resection, surveillance, or endoscopic treatment?

W. R. Brugge

GI Unit, Blake 452, Massachusetts General Hospital, Boston

Introduction

Cystic lesions of the pancreas consist of a spectrum of benign, pre-malignant, and malignant malignancies. In the past, cystic neoplasms of the pancreas were thought to be relatively rare, but the widespread use of cross sectional imaging has dramatically increased ability to detect these lesions. Although the vast majority of pancreatic cysts are discovered incidentally, large or invasive lesions may produce sufficient symptoms to cause the patient to seek medical attention.

Cystic neoplasms are often confused or misdiagnosed as pseudocysts or peripancreatic collections of inflammatory fluid that may morphologically mimic cystic neoplasms. Furthermore, the presenting symptoms of pseudocysts may be identical to the symptoms associated with cystic neoplasms.

Cystic neoplasms of the pancreas are traditionally organized by the type of lining epithelium since this feature dominates the risk of malignancy and management [1] (Table 1). There are three types of mucinous lesions, benign mucinous cystadenomas, malignant mucinous cystic lesions, and intra-ductal papillary mucinous neoplasms (IPMNs). The non-mucinous lesions include serous cystadenomas, cystic endocrine tumors and other rare lesions.

Prevalence

The prevalence of pancreatic cysts has been examined with autopsy examinations of the pancreas in adults without known pancreatic disease. The prevalence of pancreatic cysts found at autopsies in Japan was approximately 73 of 300 autopsies (24.3%) cases [2]. The prevalence of cysts increased with increasing age of the patient. The cysts were located throughout the pancreatic parenchyma and were not related to chronic pancreatitis. The epithelium of the cysts displayed a spectrum of neoplastic change, including atypical hyperplasia (16.4%); carcinoma in situ (3.4%).

Correspondence: William R. Brugge, M.D. · GI Unit · Blake 452 · Massachusetts General Hospital · Boston, MA 02114 · Phone: 001-617-724-3715 · Fax: 001-617-724-5996 · E-mail: brugge.william@mgh.harvard.edu

Bibliography: Endoscopy 2006; 38 (S1): S40–S47 © Georg Thieme Verlag KG Stuttgart · New York · ISSN 0013-726X · DOI 10.1055/s-2006-946650

Table 1

<i>Tumor Type</i>	<i>Gender</i>	<i>Age</i>	<i>Morphology</i>	<i>Type of Epithelium</i>	<i>Risk of Malignancy</i>
Mucinous Cystadenoma	Female	Middle-Aged	Unilocular	Mucinous	Moderate
Mucinous Cystic Neoplasm	Female	Middle-Aged	Associated mass	Malignant Mucinous	High
Intra-ductal Papillary Mucinous Tumor	Mixed	Elderly	Unilocular, septated, associated dilated ducts	Papillary Mucinous	Moderate
Serous Cystadenoma	Female	Middle-Aged	Microcystic	Serous (PAS positive for glycogen)	Low
Cystic Endocrine Tumor	Mixed	Middle-Aged	Associated mass	Endocrine	Low
Solid Cystic Pseudopapillary Tumor	Female	Young	Mixed solid and cystic	Endocrine-like	Low

The prevalence of pancreatic cysts in the United States has been estimated in patients undergoing MRI for a number of various medical problems [3]. This review of MRI images revealed about 15–20% of 1,444 patients had at least one pancreatic cyst. Older patients are more likely to have a cyst than younger patients. Screening abdominal ultrasound in a younger population revealed that (0.21%) of the 130,951 adults had a pancreatic cystic lesion [4].

Clinical epidemiology

Mucinous cystic neoplasms account for approximately 2% to 5% of all exocrine pancreatic tumors and are the more common type of cystic tumor. Women are affected far more commonly than men (9:1 ratio), with a mean age at diagnosis in the fifth decade.

Intraductal papillary mucinous neoplasms are similar to mucinous cystic neoplasms. Their true incidence is uncertain, but estimates range from 1% to 8%. IPMNs affect men predominantly, depending on the reported series, and they tend to occur in an older age group than MCNs.

Serous cystadenomas have been estimated to account for about 25% of all cystic neoplasms of the pancreas [5]. Estimates of the incidence and prevalence vary. Using surgical pathology studies, it has been estimated that serous cystadenomas account for about 1% to 2% of all exocrine pancreatic neoplasms.

Serous cystadenomas occur only in adults with a median age in the sixth or seventh decade. The vast majority of patients with serous cystadenomas are female [6]. Traditionally about 1/2 of tumors are discovered as incidental findings during abdominal imaging or surgery or at autopsy.

Risk factors for cystic lesions

In the vast majority of patients with a cystic lesion, no risk factor is apparent. Von Hippel Lindau (VHL) syndrome is the best-described inherited disorder associated with cystic lesions [7]. In the largest series to date, pancreatic involvement was observed in 122/158 patients (77.2%) and included true cysts (91.1%), serous cystadenomas (12.3%), neuroendocrine tumors (12.3%), or combined lesions (11.5%).

Pathogenesis

The pathogenesis of cystic neoplasms of the pancreas is poorly understood. Serous cystadenomas are strongly associated with mutations of the VHL gene, located on chromosome 3p25 [8]. The mutations in the VHL gene probably affect most commonly the centro-acinar cell and result in hamartomatous proliferation of these small cuboidal cells. In one study, 70% of the sporadic serous cystadenomas studied demonstrated loss of heterozygosity (LOH) at 3p25 with a VHL gene mutation in the remaining allele [9].

Mucinous cystic neoplasms and intraductal papillary mucinous neoplasms (IPMN) probably have a similar pathogenesis. K-ras mutations are present only in mucinous cystic neoplasms and not in serous microcystic adenomas. In addition, LOH at 3p25, the chromosomal location of VHL gene, was present in 57% (8/14) of serous microcystic adenomas compared with only 17% (2/12) of mucinous cystic neoplasms [10]. Mucinous cystic neoplasms frequently contain mutations of the K-ras oncogene and p53 tumor suppressor gene, and the frequency of these mutations increases with increasing degrees of dysplasia in the neoplasm [11]. The frequency of K-ras mutation in mucinous cystic neoplasms is linearly related to the grades of atypia [12]. However, the degree of atypia in IPMT does not seem to correlate with the presence of k-ras mutations. LOH of the p16 gene was observed with increasing degrees of histological atypia in IPMN, whereas LOH of the p53 gene was seen only in invasive carcinomas.

Pathology

Serous cystadenomas (Fig. 1)

Serous cystadenomas are benign, solitary, cystic tumors that lined by small, bland, centro-acinar cells. Although the majority of serous cystadenomas have a microcystic morphology, there are two other variants based on growth pattern: macrocystic and solid. Microcystic serous cystadenomas are composed of multiple small thin-walled cysts with a honeycomb-like appearance on cross section. Microcystic serous cystadenomas may grow to a large diameter over the long-term and the large lesions often have a fibrotic or calcified central scar. Macrocystic serous cystadenomas are composed of far fewer cysts, and the diameter of each cyst varies from microcystic to large cavities [13]. The pres-

Fig. 1 Gross photograph of a serous cystadenoma.

Fig. 2 Gross photograph of a mucinous cystic neoplasm.

ence of discrete, large cystic cavities mimics the appearance of mucinous lesions, particularly IPMT. However, the cyst fluid from serous cystadenomas is non-viscous and may contain blood as a result of the vascular nature of the lesions.

Mucinous cystic neoplasms (Fig. 2)

Mucinous cystic neoplasms (MCNs) are composed of discrete individual locules that vary in diameter. MCNs are lined by mucin-producing cells in a columnar epithelium. The World Health Organization classification catalogues MCNs into three types, based on the degree of epithelial dysplasia: benign, borderline, and malignant. The degree of atypia of the tumor is classified according to the most advanced degree of dysplasia/carcinoma present.

Mucinous cystic neoplasms of the pancreas often contain a unique, highly cellular (so-called “ovarian”) stroma that often contain estrogen and progesterone receptors. It occurs almost exclusively in female patients, although rare cases of MCNs with ovarian stroma in male patients have been encountered. Many authorities have restricted the very definition of MCNs to include only those cystic mucinous tumors that contain ovarian stroma. The cyst fluid from MCNs is often viscous and clear.

IPMNs are similar to MCNs in that they are cystic tumors that secrete mucin. However, IPMNs are characterized by a unique pap-

illary epithelium and arise from ductal epithelium. The presence of a papillary neoplasm causes the pancreatic duct to dilate. The degree of ductal ectasia produced varies with degree of mucin production, but duct dilatation great enough to be seen on imaging studies or gross pathologic examination is a diagnostic feature of the diagnosis. Mucin production may be so excessive that mucin will be spontaneously secreted out of the ampulla. The degree of dysplasia exhibited by the epithelium may range from mild to moderate to severe (carcinoma in situ), and the foci of early malignancy may be evident by the presence of mural nodules [14]. The solid malignancies that arise from IPMN are more likely to have papillary features, as compared to typical pancreatic malignancies that arise from the main pancreatic duct [15].

Cystic endocrine neoplasms

Cystic neoplasms are composed of neuroendocrine tissue. The cystic lesions are not true cysts since they are composed of centralized tumor necrosis. The lesions are rare and comprise 0.5–4% of all primary pancreatic neoplasms. The classic neuroendocrine cystic tumor is populated with a characteristic small, granular population of cells that are stainable for immunoreactive hormones, chromogranin and synaptophysin [16]. It is rare for the cystic endocrine tumors to produce sufficient hormones to be clinically active. Cystic endocrine tumors are seen in association with Von Hippel-Lindau syndrome [17]. A related cystic lesion, the solid pseudopapillary tumor contains cells with neuroendocrine characteristics [18]. The tumor often contains areas of hemorrhage and necrosis as well as cystic components. These neoplasms exhibit low grade malignant behavior and have an excellent prognosis when resected, but a small percent metastasize.

Clinical presentation

Most patients with a pancreatic cystic lesion have non-specific symptoms [19]. The cystic lesion is usually found with CT or US imaging performed for the evaluation of another condition. When symptoms are present, the most common presentation is recurrent abdominal pain, nausea, and vomiting as result of mild pancreatitis [19]. Cystic lesions that cause duct compression or involvement of the main pancreatic duct are prone to cause pancreatitis. Chronic abdominal pain and jaundice are a rare presentation of a cystic lesion and suggests a malignancy or a pseudocyst. Patients with a cystic malignancy will present with symptoms and signs similar to pancreatic cancer, i.e. pain, weight loss, and jaundice [20]. Pseudocysts may arise after an episode of acute pancreatitis or insidiously in the setting of chronic pancreatitis and are associated with chronic abdominal pain. It is common for cystic lesions associated with pancreatitis to be diagnosed as pseudocysts and be confused with cystic neoplasms that also cause pancreatitis [21].

Differential diagnosis (Table 2)

The differential diagnosis of a cystic lesion of pancreas is very wide and often causes confusion. Since the treatment of a pseudocyst and cystic neoplasm are so different, it is incumbent on the clinician to first differentiate between these major categories of lesions. Although it is unusual for a patient with a pseudocyst to present without preceding symptoms, it may occur in mild

Table 2

Type of Lesion	Morphology	Location	Cytology	CEA (ng/ml)
Serous	Microcystic	Evenly distributed	Bland PAS +	< 0.5
Mucinous	Macrocytic	Tail	Mucinous	> 200
IPMT	Mixed	Head	Mucinous	> 200
Pseudocyst	Unilocular	Evenly distributed	Pigmented histiocytes	< 200

chronic pancreatitis. Evidence of inflammatory changes or calcifications in the pancreas is suggestive of a pancreatic pseudocyst. However, in the initial setting of mild pancreatitis it may be difficult to differentiate between a cystic neoplasm that has caused pancreatitis and a small pseudocyst that has formed as a result of pancreatitis. If a cystic lesion has been present for many years, it is highly likely that the lesion represents a cystic neoplasm. Congenital cysts of the pancreas are rare [22].

Once a pancreatic pseudocyst has been excluded, attention should be focused on the differential between the types of cystic neoplasms. The principal differentiation is between mucinous and serous lesions because the fundamental difference in management is based on the neoplastic potential of mucinous lesions. The non-neoplastic serous cystadenomas may be diagnosed on the initial imaging test because their typical microcystic morphology. Once a serous lesion has been confidently diagnosed, the lesion may be followed by serial imaging, looking for evidence of growth or organ impingement. In contrast, the approach to mucinous lesions is quite different. The underlying risk of malignancy or the development of malignancy often results in resection of the lesion. Under some clinical circumstances, such as patients who are at high risk for complications of pancreatotomy, differentiation between benign and grossly malignant mucinous lesions is important. The risk of surgery must be weighed against the risk of malignancy in the decision-making process. The risk of surgery must also take into account the variation in risk that is inherent in the location of the lesion.

Diagnostic methods

CT is an excellent test for cystic lesions of the pancreas because of its widespread availability and ability to detect cysts [23] (Fig. 3). MR imaging is used increasingly because of its ability to determine if there is involvement of the main pancreatic duct [24]. Ultrasonography whether performed transabdominally or intraoperatively is generally not helpful [25]. Recently PET scanning has been shown to be positive in a high percentage of malignant cystic lesions [26].

Although seen in less than 20 percent of lesions, demonstration of a central scar by CT or MR is a highly diagnostic feature of a serous cystadenoma [27]. The honey-combed or microcystic appearance of the lesion is commonly used to provide a diagnosis. However, macrocystic serous cystadenomas are difficult to diagnose with cross-sectional imaging because of the morphologic similarities with mucinous lesions [13,28]. The presence of mul-

tiple small thin-walled cysts is suggestive of Von Hippel Lindau syndrome [29]. Mucinous cystic neoplasms, in contrast, are commonly diagnosed with CT based on the unilocular or macrocystic characteristics [30]. Although not frequently seen, the finding of peripheral calcification by CT is specific for a mucinous cystic neoplasm. Intraductal papillary mucinous neoplasms (IPMN) may involve the main pancreatic duct exclusively, a side-branch or both. MRCP can demonstrate the diagnostic findings of pancreatic duct dilation, mural nodules, and ductal connection better than ERCP [31]. However, ERCP will can demonstrate the intraductal filling defects, mucin extrusion, and cystic side branches that are associated with IPMN in 70–90% of patients [32,33].

Despite these imaging features, the ability to accurately diagnose a specific cystic lesion and to determine whether malignancy is present by CT and MR remains uncertain (Fig. 4). The diagnosis of a pancreatic pseudocyst is more dependent upon the clinical history and the associated findings of chronic pancreatitis. Pancreatic pseudocysts appear as unilocular fluid-filled cavities associated with parenchymal changes such as calcifications and atrophy.

Recently endoscopic ultrasound (EUS) has been used to diagnose cystic lesions of the pancreas and guide fine needle aspiration (FNA) [34]. The detailed imaging features of cystic neoplasms by EUS do not appear to be sufficiently accurate to differentiate between benign and malignant cystadenomas unless there is evidence of a solid mass or invasive tumor [35] (Fig. 5). EUS is also very sensitive for detecting IPMN lesions, but imaging alone may not be sufficient for differentiating between benign and ma-

Fig. 3 CT scan of a mucinous cystic neoplasm in the tail of the pancreas.

Fig. 4 CT scan of a mucinous cystadenocarcinoma.

Fig. 5 EUS image of a mucinous cystadenocarcinoma: note the mass in the wall of the cyst.

lignant lesions [36,37]. The strength of EUS is its ability to detect and aspirate small cystic lesions with a high level of safety [38]. EUS is more accurate in the evaluation of lesions less than 3 cm in diameter [39]. The macrocystic variant of serous cystadenomas can be diagnosed with EUS using a combination of a thick cyst wall, the presence of microcysts, and a low cyst fluid CEA [40].

The fluid contents of cystadenomas are often analyzed for cytology [41]. However, the low cellular content of cyst fluid has hampered the use of the cytologic analysis of cyst fluid. Small, cuboidal cells in cytologic specimens are diagnostic of serous cystadenomas. In contrast, mucinous cystadenoma may have large secretory epithelial cells with evidence of mucin secretion or atypia [42]. Only inflammatory cells should be present in the fluid aspirated from pseudocysts.

Fig. 6 ERCP of IPMT: Note the dilated and tortuous main pancreatic duct.

A variety of cyst fluid tumor markers have been studied to help differentiate between the major types of cystic neoplasms. Serum 19-9 is elevated in frank cystic malignancies [43]. A number of glycoproteins are present in the epithelium of mucinous neoplasms and are secreted into the cyst fluid [44]. The presence of extracellular mucin in aspirated cyst fluid is moderately predictive of a mucinous neoplasm [45]. Several studies suggest that carcinoembryonic antigen (CEA) or CA 72-4 are useful for identifying mucinous lesions [46,47]. These carbohydrate antigens are secreted by the epithelium lining mucinous lesions and may be present in high concentrations. Cyst fluid concentrations of CEA and CA 72-4 are very low in serous cystadenomas [48]. Despite considerable overlap between mucinous and non-mucinous cysts, cyst fluid CEA is the most accurate marker [47,49]. Cyst fluid CEA of less than 5 ng/ml is highly diagnostic of serous cystadenomas and values greater than 800 are predictive of mucinous lesions [49]. Recently molecular studies of cyst fluid DNA have revealed *KRAS*, tumor suppressor gene mutations, telomerase activity in mucinous cystic lesions [11,50].

Intra-ductal papillary mucinous neoplasms can be evaluated with endoscopic retrograde cholangio-pancreatography (ERCP) or EUS [51]. Prior to ERCP or EUS, the endoscopic finding of a patulous ampulla filled with mucin is diagnostic of an intraductal papillary mucinous neoplasm. Contrast retrograde pancreatography will demonstrate the characteristic findings of mucinous filling defects within the duct, diffuse ductal dilation, and cystic dilation of side branches [52]. MRCP may be more sensitive for detecting the sidebranch lesions of IPMN [53]. EUS may assist in the detection of malignancy arising from intraductal papillary mucinous neoplasms by demonstrating focal nodules, invasive lesions and guiding fine needle aspiration of suspicious lesions

Table 3 Use of FNA samples: prioritizing the use of samples

Type of lesion	First priority	Second priority	Third priority	Experimental
Serous	CEA	Imaging	Fluid cytology	VHL gene testing
Mucinous	CEA	Cytology	Subjective assessment of viscosity	kRAS LOH analysis
Malignant	Tissue cytology	Fluid cytology	CEA	kRAS LOH analysis

[54]. EUS can also be used to monitor IPMN lesions, looking for increases in the size of cysts and the diameter of the main pancreatic duct [55].

Diagnostic evaluation

Patients suspected of having a cystic neoplasm of the pancreas should undergo a CT scan with contrast as the initial test. If no lesion is seen in the pancreas, it is very unlikely that a clinically significant neoplasm is present. MR imaging, particularly MRCP, may be substituted for CT scanning. If the CT scan demonstrate a diagnostic finding, such as a classic microcystic serous cystadenoma, a malignant cystic mass, or pancreatitis with a fluid collection, no further evaluation is necessary. If the patient is a young woman and there is a solitary, unilocular cystic lesion in the tail of the pancreas, the patient should undergo surgical resection. Indeterminate lesions should undergo EUS with FNA. Aspirated cyst fluid should be analyzed for CEA, cytology, and amylase (Table 3). Preference should be given to sending the fluid for CEA, over cytology in small mucinous lesions. Cytology should be used preferentially in malignant appearing cystic lesions. IPMN lesions should have FNA of cystic lesions, an enlarged pancreatic duct, and focal mass lesions. Cytology should be used preferentially.

Treatment

Surgical resection is the treatment of choice for pre-malignant cystic neoplasms. The decision to resect a lesion, however, is based on the presence or absence of symptoms, the risk of malignancy, and the surgical risk of the patient. High risk patients with low grade cystic neoplasms may be monitored with periodic CT/MRI scanning or EUS-FNA [56]. Experimentally, high risk patients have been treated with EUS-guided ethanol lavage which safely produces ablation of the cyst epithelium [57]. Small cystic lesions in the elderly can be safely monitored [58].

The increasing safety of surgical resection has prompted the use of surgery for a wider range of lesions [59]. However, serous cystadenomas do not require resection except for relief of symptoms [60]. Since most mucinous cystic neoplasms are located in the tail of the pancreas, a distal pancreatectomy is sufficient for these pre-malignant lesions. Since intraductal papillary mucinous neoplasms invade the pancreas along ductal structures, it is important that frozen section histology be used during surgery to assure negative margins [61,62].

Prognosis

Cystic neoplasms are slow growing and 19% will demonstrate an increase in diameter at 16 months [63]. Surgical resection is associated with a morbidity of 27.9%, with a reoperation rate of 7.3% and a very low mortality rate [6]. The overall 5-year survival for patients having IPMNs without invasive cancer was 77%, compared with 43% in those patients with an invasive component [64]. The overall post-operative recurrence rate varies from 7% to 43% [65]. Approximately 50% of patients will have evidence of malignancy in the resected specimen [66]. Similar survival rates are seen in patients with mucinous cystic neoplasms [67]. Side branch lesions arising from IPMN have a better prognosis than main duct IPMN [68]. The worst prognosis is for advanced, transmural adenocarcinomas arising from mucinous lesions; the 5-year survival is only 30% for resected lesions.

References

- Brugge WR, Lauwers GY, Sahani D et al. Cystic neoplasms of the pancreas. *N Engl J Med* 2004; 351: 1218–1226
- Kimura W, Nagai H, Kuroda A et al. Analysis of small cystic lesions of the pancreas. *Int J Pancreatol* 1995; 18: 197–206
- Zhang XM, Mitchell DG, Dohke M et al. Pancreatic cysts: depiction on single-shot fast spin-echo MR images. *Radiology* 2002; 223: 547–553
- Ikeda M, Sato T, Morozumi A et al. Morphologic changes in the pancreas detected by screening ultrasonography in a mass survey, with special reference to main duct dilatation, cyst formation, and calcification. *Pancreas* 1994; 9: 508–512
- Compton CC. Serous cystic tumors of the pancreas. *Semin Diagn Pathol* 2000; 17: 43–55
- Bassi C, Salvia R, Molinari E et al. Management of 100 consecutive cases of pancreatic serous cystadenoma: wait for symptoms and see at imaging or vice versa? *World J Surg* 2003; 27: 319–323
- Hammel P. [Diagnostic value of cyst fluid analysis in cystic lesions of the pancreas: current data, limitations, and perspectives]. *J Radiol* 2000; 81: 487–490
- Moore PS, Zamboni G, Brighenti A et al. Molecular characterization of pancreatic serous microcystic adenomas: evidence for a tumor suppressor gene on chromosome 10q. *Am J Pathol* 2001; 158: 317–321
- Vortmeyer AO, Lubensky IA, Fogt F et al. Allelic deletion and mutation of the von Hippel-Lindau (VHL) tumor suppressor gene in pancreatic microcystic adenomas. *Am J Pathol* 1997; 151: 951–956
- Ahmad NA, Kochman ML, Brensinger C et al. Interobserver agreement among endosonographers for the diagnosis of neoplastic versus non-neoplastic pancreatic cystic lesions. *Gastrointest Endosc* 2003; 58: 59–64
- Khalid A, McGrath KM, Zahid M et al. The role of pancreatic cyst fluid molecular analysis in predicting cyst pathology. *Clin Gastroenterol Hepatol* 2005; 3: 967–973
- Yoshizawa K, Nagai H, Sakurai S et al. Clonality and K-ras mutation analyses of epithelia in intraductal papillary mucinous tumor and mucinous cystic tumor of the pancreas. *Virchows Arch* 2002; 441: 437–443

- 13 Khurana B, Morteale KJ, Glickman J et al. Macrocystic serous adenoma of the pancreas: radiologic-pathologic correlation. *AJR Am J Roentgenol* 2003; 181: 119–123
- 14 Tanaka M. Intraductal papillary mucinous neoplasm of the pancreas: diagnosis and treatment. *Pancreas* 2004; 28: 282–288
- 15 Furuta K, Watanabe H, Ikeda S. Differences between solid and duct-ectatic types of pancreatic ductal carcinomas. *Cancer* 1992; 69: 1327–1333
- 16 Kann P, Bittinger F, Engelbach M et al. Endosonography of insulin-secreting and clinically non-functioning neuroendocrine tumors of the pancreas: criteria for benignancy and malignancy. *Eur J Med Res* 2001; 6: 385–390
- 17 Marcos HB, Libutti SK, Alexander HR et al. Neuroendocrine tumors of the pancreas in von Hippel-Lindau disease: spectrum of appearances at CT and MR imaging with histopathologic comparison. *Radiology* 2002; 225: 751–758
- 18 Notohara K, Hamazaki S, Tsukayama C et al. Solid-pseudopapillary tumor of the pancreas: immunohistochemical localization of neuroendocrine markers and CD10. *Am J Surg Pathol* 2000; 24: 1361–1371
- 19 Wiesenauer CA, Schmidt CM, Cummings OW et al. Preoperative predictors of malignancy in pancreatic intraductal papillary mucinous neoplasms. *Arch Surg* 2003; 138: 610–7; discussion 617–618
- 20 Holly EA, Chaliha I, Bracci PM et al. Signs and symptoms of pancreatic cancer: a population-based case-control study in the San Francisco Bay area. *Clin Gastroenterol Hepatol* 2004; 2: 510–517
- 21 Sand J, Nordback I. The differentiation between pancreatic neoplastic cysts and pancreatic pseudocyst. *Scand J Surg* 2005; 94: 161–164
- 22 Caillot JL, Rongieras F, Voiglio E et al. A new case of congenital cyst of the pancreas. *Hepatogastroenterology* 2000; 47: 916–918
- 23 Curry CA, Eng J, Horton KM et al. CT of primary cystic pancreatic neoplasms: can CT be used for patient triage and treatment? *AJR Am J Roentgenol* 2000; 175: 99–103
- 24 Fukukura Y, Fujiyoshi F, Hamada H et al. Intraductal papillary mucinous tumors of the pancreas. Comparison of helical CT and MR imaging. *Acta Radiol* 2003; 44: 464–471
- 25 Kubota K, Noie T, Sano K et al. Impact of intraoperative ultrasonography on surgery for cystic lesions of the pancreas. *World J Surg* 1997; 21: 72–76; discussion 77
- 26 Sperti C, Pasquali C, Decet G et al. F-18-fluorodeoxyglucose positron emission tomography in differentiating malignant from benign pancreatic cysts: a prospective study. *J Gastrointest Surg* 2005; 9: 22–28; discussion 28–29
- 27 Torresan F, Casadei R, Solmi L et al. The role of ultrasound in the differential diagnosis of serous and mucinous cystic tumours of the pancreas. *Eur J Gastroenterol Hepatol* 1997; 9: 169–172
- 28 Chatelain D, Hammel P, O'Toole D et al. Macrocystic form of serous pancreatic cystadenoma. *Am J Gastroenterol* 2002; 97: 2566–2571
- 29 Cheng TY, Su CH, Shyr YM et al. Management of pancreatic lesions in von Hippel-Lindau disease. *World J Surg* 1997; 21: 307–312
- 30 Sahani D, Prasad S, Saini S et al. Cystic pancreatic neoplasms evaluation by CT and magnetic resonance cholangiopancreatography. *Gastrointest Endosc Clin N Am* 2002; 12: 657–672
- 31 Sugiyama M, Atomi Y, Hachiya J. Intraductal papillary tumors of the pancreas: evaluation with magnetic resonance cholangiopancreatography. *Am J Gastroenterol* 1998; 93: 156–159
- 32 Madura JA, Wiebke EA, Howard TJ et al. Mucin-hypersecreting intraductal neoplasms of the pancreas: a precursor to cystic pancreatic malignancies. *Surgery* 1997; 122: 786–92; discussion 792–793
- 33 Azar C, Van de Stadt J, Rickaert F et al. Intraductal papillary mucinous tumours of the pancreas. Clinical and therapeutic issues in 32 patients. *Gut* 1996; 39: 457–464
- 34 Brugge WR. Evaluation of pancreatic cystic lesions with EUS. *Gastrointest Endosc* 2004; 59: 698–707
- 35 Ahmad NA, Kochman ML, Lewis JD et al. Can EUS alone differentiate between malignant and benign cystic lesions of the pancreas? *Am J Gastroenterol* 2001; 96: 3295–3300
- 36 Brandwein SL, Farrell JJ, Centeno BA et al. Detection and tumor staging of malignancy in cystic, intraductal, and solid tumors of the pancreas by EUS. *Gastrointest Endosc* 2001; 53: 722–727
- 37 Terris B, Ponsot P, Paye F et al. Intraductal papillary mucinous tumors of the pancreas confined to secondary ducts show less aggressive pathologic features as compared with those involving the main pancreatic duct. *Am J Surg Pathol* 2000; 24: 1372–1377
- 38 Lee LS, Saltzman JR, Bounds B et al. EUS-guided fine needle aspiration of pancreatic cysts: a retrospective analysis of complications and their predictors. *Clin Gastroenterol Hepatol* 2005; 3: 231–236
- 39 Volmar KE, Vollmer RT, Jowell PS et al. Pancreatic FNA in 1000 cases: a comparison of imaging modalities. *Gastrointest Endosc* 2005; 61: 854–861
- 40 O'Toole D, Palazzo L, Hammel P et al. Macrocystic pancreatic cystadenoma: The role of EUS and cyst fluid analysis in distinguishing mucinous and serous lesions. *Gastrointest Endosc* 2004; 59: 823–829
- 41 Centeno BA, Warshaw AL, Mayo-Smith W et al. Cytologic diagnosis of pancreatic cystic lesions. A prospective study of 28 percutaneous aspirations. *Acta Cytol* 1997; 41: 972–980
- 42 Recine M, Kaw M, Evans DB et al. Fine-needle aspiration cytology of mucinous tumors of the pancreas. *Cancer* 2004; 102: 92–99
- 43 Sperti C, Pasquali C, Guolo P et al. Serum tumor markers and cyst fluid analysis are useful for the diagnosis of pancreatic cystic tumors. *Cancer* 1996; 78: 237–243
- 44 Yamaguchi K, Enjoji M. Cystic neoplasms of the pancreas. *Gastroenterology* 1987; 92: 1934–1943
- 45 Walsh RM, Henderson JM, Vogt DP et al. Prospective preoperative determination of mucinous pancreatic cystic neoplasms. *Surgery* 2002; 132: 628–33; discussion 633–634
- 46 Frossard JL, Amouyal P, Amouyal G et al. Performance of endosonography-guided fine needle aspiration and biopsy in the diagnosis of pancreatic cystic lesions. *Am J Gastroenterol* 2003; 98: 1516–1524
- 47 Brugge WR, Lewandrowski K, Lee-Lewandrowski E et al. Diagnosis of pancreatic cystic neoplasms: a report of the cooperative pancreatic cyst study. *Gastroenterology* 2004; 126: 1330–1336
- 48 Hammel P, Voitot H, Vilgrain V et al. Diagnostic value of CA 72–4 and carcinoembryonic antigen determination in the fluid of pancreatic cystic lesions. *Eur J Gastroenterol Hepatol* 1998; 10: 345–348
- 49 van der Waaij LA, van Dullemen HM, Porte RJ. Cyst fluid analysis in the differential diagnosis of pancreatic cystic lesions: a pooled analysis. *Gastrointest Endosc* 2005; 62: 383–389
- 50 Zhou GX, Huang JF, Li ZS et al. Detection of K-ras point mutation and telomerase activity during endoscopic retrograde cholangiopancreatography in diagnosis of pancreatic cancer. *World J Gastroenterol* 2004; 10: 1337–1340
- 51 Telford JJ, Carr-Locke DL. The role of ERCP and pancreatoscopy in cystic and intraductal tumors. *Gastrointest Endosc Clin N Am* 2002; 12: 747–757
- 52 Dachman AH, Namieno T, Ichimura T et al. Mucin-producing pancreatic tumors: comparison of MR cholangiopancreatography with endoscopic retrograde cholangiopancreatography. *Radiology* 1998; 208: 231–237
- 53 Koito K, Namieno T, Ichimura T et al. Mucin-producing pancreatic tumors: comparison of MR cholangiopancreatography with endoscopic retrograde cholangiopancreatography. *Radiology* 1998; 208: 231–237
- 54 Sugiyama M, Atomi Y, Saito M. Intraductal papillary tumors of the pancreas: evaluation with endoscopic ultrasonography. *Gastrointest Endosc* 1998; 48: 164–171
- 55 Kobayashi G, Fujita N, Noda Y et al. Mode of progression of intraductal papillary-mucinous tumor of the pancreas: analysis of patients with follow-up by EUS. *J Gastroenterol* 2005; 40: 744–751
- 56 Irie H, Yoshimitsu K, Aibe H et al. Natural history of pancreatic intraductal papillary mucinous tumor of branch duct type: follow-up study by magnetic resonance cholangiopancreatography. *J Comput Assist Tomogr* 2004; 28: 117–122
- 57 Gan SI, Thompson CC, Lauwers GY et al. Ethanol lavage of pancreatic cystic lesions: initial pilot study. *Gastrointest Endosc* 2005; 61: 746–752
- 58 Kimura W, Makuuchi M. Operative indications for cystic lesions of the pancreas with malignant potential – our experience. *Hepatogastroenterology* 1999; 46: 483–491
- 59 Fernandez del Castillo CF, Targarona J, Thayer SP et al. Incidental pancreatic cysts: clinicopathologic characteristics and comparison with symptomatic patients. *Arch Surg* 2003; 138: 427–434
- 60 Le Borgne J, de Calan L, Partensky C. Cystadenomas and cystadenocarcinomas of the pancreas: a multiinstitutional retrospective study of 398 cases. *French Surgical Association. Ann Surg* 1999; 230: 152–161
- 61 Gigot JF, Deprez P, Sempoux C et al. Surgical management of intraductal papillary mucinous tumors of the pancreas: the role of routine frozen section of the surgical margin, intraoperative endoscopic staged biopsies of the Wirsung duct, and pancreaticogastric anastomosis. *Arch Surg* 2001; 136: 1256–1262

- ⁶² Chari ST, Yadav D, Smyrk TC et al. Study of recurrence after surgical resection of intraductal papillary mucinous neoplasm of the pancreas. *Gastroenterology* 2002; 123: 1500–1507
- ⁶³ Spinelli KS, Fromwiller TE, Daniel RA et al. Cystic pancreatic neoplasms: observe or operate. *Ann Surg* 2004; 239: 651–7; discussion 657–659
- ⁶⁴ Sohn TA, Yeo CJ, Cameron JL et al. Intraductal papillary mucinous neoplasms of the pancreas: an updated experience. *Ann Surg* 2004; 239: 788–97; discussion 797–799
- ⁶⁵ Lai EC, Lau WY. Intraductal papillary mucinous neoplasms of the pancreas. *Surgeon* 2005; 3: 317–324
- ⁶⁶ Falconi M, Salvia R, Bassi C et al. Clinicopathological features and treatment of intraductal papillary mucinous tumour of the pancreas. *Br J Surg* 2001; 88: 376–381
- ⁶⁷ Suzuki Y, Atomi Y, Sugiyama M et al. Cystic neoplasm of the pancreas: a Japanese multiinstitutional study of intraductal papillary mucinous tumor and mucinous cystic tumor. *Pancreas* 2004; 28: 241–246
- ⁶⁸ Kobari M, Egawa S, Shibuya K et al. Intraductal papillary mucinous tumors of the pancreas comprise 2 clinical subtypes: differences in clinical characteristics and surgical management. *Arch Surg* 1999; 134: 1131–1136