

The *In Vitro* and *In Vivo* Effects of *Hypoxis hemerocallidea* on Indinavir Pharmacokinetics: Modulation of Efflux

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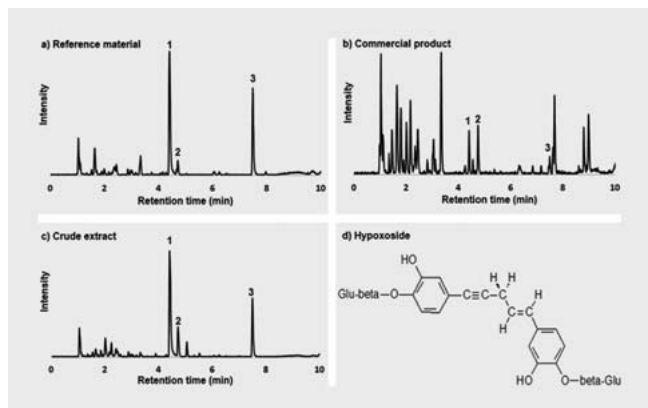
ABSTRACT

Hypoxis hemerocallidea (African potato) is a popular medicinal plant that has been used traditionally for the treatment of various disorders. Some HIV/AIDS patients use this traditional medicine together with their antiretroviral therapy. This study aimed to determine the impact of selected *H. hemerocallidea* materials (i.e., a commercial product, an aqueous extract, and biomass reference plant material) on the bidirectional permeability of indinavir across Caco-2 cell monolayers as well as the bioavailability of indinavir during an acute, single administration study in Sprague-Dawley rats. All of the selected *H. hemerocallidea* test materials demonstrated inhibition effects on indinavir efflux across Caco-2 cell monolayers, albeit to different extents. An increase in the bioavailability of indinavir was obtained *in vivo* when administered concomitantly with the *H. hemerocallidea* materials, albeit not statistically significantly. The change in bioavailability directly correlated with the *in vitro* permeability results. It can therefore be concluded that the change in permeability and bioavailability of indinavir was caused by efflux inhibition and this effect was dependent on the type of *H. hemerocallidea* material investigated, which was found to be in the following order: commercial product > aqueous extract > reference plant material. The clinical significance of the combined effect of efflux and metabolism inhibition by *H. hemerocallidea* should be determined in another *in vivo* model that expresses the cytochrome P450 3A4 enzyme.

Introduction

Contradicting utilisation rates of traditional medicines are reported in the scientific literature for different countries around the globe. For example, it is frequently stated that 80% of the population of developing countries, especially sub-Saharan Africa, use traditional medicines [1–4], which is based on data obtained in 1983 [5]. A more recent survey done in six middle-income countries (China, Ghana, India, Mexico, Russia, and South Africa) as part of a World Health Organization (WHO) Study on Global AGEing and adult health (SAGE) found that the use of traditional

medicine seems to be substantially lower (< 3%) [5]. Although traditional medicine use in Africa has decreased according to recent surveys, it varies widely among countries and regions on this continent [6]. There still seems to be a relatively large demand for medicinal plant usage by rural people, as shown by a study conducted in the Amatola region of the Eastern Cape Province in South Africa [7]. In addition, a survey in the Southern Africa region (including Botswana, Lesotho, South Africa, and Swaziland) found that 33.5% of participants indicated that people living with HIV/AIDS use complementary treatments, which include traditional medicines, to help manage their symptoms [8]. Another relatively



► **Fig. 1** Chromatograms (TIC) of *H. hemerocallidea* materials: **a** reference plant material, **b** commercial product, **c** aqueous crude extract, and **d** chemical structure of hypoxoside [24]. Peak 1 = hypoxoside, peak 2 = dehydroxyhypoxoside, peak 3 = geraniol glycoside.

recent survey in the eThekweni Metropolitan area of the KwaZulu Natal Province in South Africa showed that more than 65.0% of the participants reported use of traditional medicines before the diagnosis of HIV and 77.6% reported traditional medicine use after their HIV diagnosis, while 4.98% of the patients used traditional medicines concurrently with their antiretroviral medicines during the study period [9].

Use of herbal products in conjunction with conventional medicines may lead to herb-drug pharmacokinetic or pharmacodynamic interactions [10–12]. Pharmacokinetic interactions may occur due to changes in the absorption, distribution, metabolism, and/or elimination (ADME) of a drug as a result of coadministration with an herbal product [13,14]. Mechanisms of action by which herb-drug pharmacokinetic interactions usually take place include modulation of intestinal active efflux transporters, such as P-glycoprotein (P-gp), as well as modulation of metabolising enzymes, specifically the cytochrome P450 (CYP450) super family [15]. These pharmacokinetic interactions may lead to increased or decreased bioavailability of the coadministered drug. Consequences of these bioavailability changes may either be adverse effects and even drug toxicity due to increased drug plasma levels or lack of pharmacological responses and treatment failure due to decreased drug plasma levels [16].

Hypoxis hemerocallidea Fisch., C.A.Mey. & Avé-Lall. (Hypoxidaceae) is a popular traditional medicinal plant that is characterised by star-shaped flowers, which are bright yellow in colour, and by strap-like leaves [17]. This plant has a potato shaped tuberous rootstock (i.e., the corm), which is referred to as the “African potato”. These corms are washed and chopped and the decoction is administered orally after boiling [1]. *H. hemerocallidea* extract, which contains phytosterols as well as hypoxoside and its active analogue rooperol, has been used for the treatment of certain types of cancers, heart failures, nervous disorders, immune-related illnesses, and urinary tract infections [18], and has also been used for the treatment of benign prostate hyperplasia as an anti-inflammatory agent, antioxidant, anticonvulsant, and as an anti-

► **Table 1** Quantity of hypoxoside in each of the selected *H. hemerocallidea* materials.

Test material	Hypoxoside (mg/g)
Reference dried plant material	13.3
Commercial product	0.6
Aqueous extract	151.7

diabetic agent [19]. *In vitro* studies have suggested the possibility that *H. hemerocallidea* can cause pharmacokinetic interactions with drugs by means of inhibiting CYP450 enzymes as well as by modulation of efflux transporters [20]. Pharmacokinetic interactions were also observed between African potato decoctions and efavirenz in human volunteers. However, efavirenz is only a substrate for CYP450 enzymes and is not a substrate for efflux transporters such as P-gp [21]. Due to the interplay between P-gp efflux and CYP450-related metabolism, these two processes should ideally be considered together in terms of pharmacokinetic interactions [22,23]. This study was specifically conducted to identify pharmacokinetic interactions between *H. hemerocallidea* materials and an antiretroviral drug (i.e., indinavir) based on efflux modulation, since indinavir is a substrate for the efflux transporter P-gp.

Results and Discussion

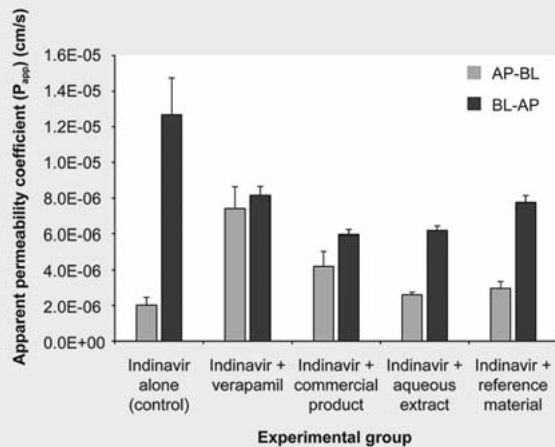
The LC/MS chromatograms (chemical fingerprinting) of the *H. hemerocallidea* materials are shown in ► **Fig. 1**, while the quantities of hypoxoside in each of the selected *H. hemerocallidea* test materials are shown in ► **Table 1**.

Chromatographic analysis revealed that all three of the selected *H. hemerocallidea* materials contained the marker molecule hypoxoside, albeit in different quantities. The aqueous extract of the corms contained a higher quantity of hypoxoside compared to that of the reference plant material and commercial product. The relatively low hypoxoside content of the commercial product can possibly be explained by the formulation composition, which caused a dilution effect due to other excipients in the dosage form as well as possible degradation of this phytochemical during manufacture of the product and/or storage of the raw materials.

The apparent permeability coefficient (P_{app}) values for indinavir in both directions across the Caco-2 cell monolayers in the presence of each selected *H. hemerocallidea* material as well as the control groups are shown in ► **Fig. 2**.

► **Table 2** presents the efflux ratio (ER) values calculated from the bidirectional transport of indinavir in combination with the *H. hemerocallidea* materials compared to the control groups.

Verapamil, a known P-gp inhibitor, demonstrated an increased uptake of indinavir in the apical-to-basolateral (AP-BL) direction compared to that of the negative control group (i.e., indinavir alone), which can be explained by an inhibition of efflux. Inhibition of efflux was confirmed by a decreased transport in the basolateral-to-apical (BL-AP) direction of indinavir as well as the ER value when compared to that of the negative control group. The Caco-



► **Fig. 2** Bidirectional P_{app} values calculated from indinavir transport across Caco-2 cell monolayers in combination with the selected *H. hemerocallidea* materials (500 $\mu\text{g}/\text{mL}$) as well as the negative control group (200 μM , indinavir alone) and positive control group (indinavir with verapamil (100 μM)). AP-BL = apical-to-basolateral direction, BL-AP = basolateral-to-apical direction, $n = 3$, error bars indicate standard deviation.

Caco-2 cell line therefore proved to be an acceptable model for testing efflux modulation of indinavir. Similarly, the transport of indinavir was increased in the AP-BL direction by all three selected *H. hemerocallidea* materials investigated in this study compared to that of the control group, while indinavir transport was decreased in the BL-AP direction compared to that of the control group. This clearly showed that active efflux transport of indinavir was inhibited by the selected *H. hemerocallidea* materials, albeit not to the same extent of efflux inhibition observed for verapamil.

The effect on the efflux of indinavir in the Caco-2 model depended on the type of *H. hemerocallidea* material, which contained different concentrations of hypoxoside. However, the efflux inhibition did not correlate directly with the hypoxoside concentration and therefore other phytoconstituents also contributed to this effect. From the *in vitro* transport results obtained in this study, it can be expected that coadministration of indinavir (as well as other antiretroviral drugs that are substrates for P-gp) with *H. hemerocallidea* extracts and plant materials will most probably lead to enhanced blood plasma levels of indinavir.

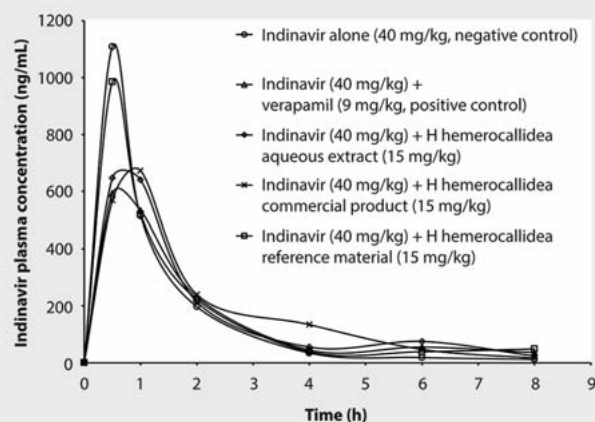
Indinavir is a known substrate for CYP3A4 [25], and *H. hemerocallidea* extracts have been shown to inhibit CYP3A4 enzyme activity within *in vitro* models [26], however, this study only focussed on the effect of the modulation of efflux on indinavir permeation and bioavailability by different *H. hemerocallidea* materials.

Validation of the LC/MS/MS method provided an accuracy between 94.0 and 106.9%, precision (coefficient of variance or CV) below 7.2%, and linearity (correlation coefficient or R^2) of 0.999 for indinavir in the quality control samples. The plasma concentration time curves for indinavir alone and in the presence of the selected *H. hemerocallidea* materials administered after a single administration (acute study) are shown in ► **Fig. 3**, while the mean

► **Table 2** ER values calculated from the bidirectional transport of indinavir across Caco-2 cell monolayers in the absence and presence of the selected *H. hemerocallidea* materials.

Experimental group	ER \pm SD*
Indinavir (200 μM) alone (negative control)	6.27 \pm 4.75
Indinavir (200 μM) with verapamil (100 μM , positive control)	1.10 \pm 0.40
Indinavir (200 μM) with <i>H. hemerocallidea</i> commercial product (500 $\mu\text{g}/\text{mL}$)	1.42 \pm 0.33
Indinavir (200 μM) with <i>H. hemerocallidea</i> aqueous extract (500 $\mu\text{g}/\text{mL}$)	2.37 \pm 1.70
Indinavir (200 μM) with <i>H. hemerocallidea</i> reference material (500 $\mu\text{g}/\text{mL}$)	2.61 \pm 1.02

*SD = standard deviation



► **Fig. 3** Plasma concentration time curves for indinavir alone (40 mg/kg, negative control) and in the presence of selected *H. hemerocallidea* materials (15 mg/kg) as well as verapamil (9 mg/kg, positive control); $n = 5$, error bars are omitted for reasons of clarity.

plasma concentrations at each time point with standard deviations are given in **Table 1S**, Supporting Information.

The bioavailability parameters for indinavir alone and in the presence of the selected *H. hemerocallidea* test materials after a single administration (acute study) are listed in ► **Table 3**. These bioavailability parameters include the area under the curve extrapolated to infinity ($AUC_{0-\infty}$) as well as maximum plasma concentration (C_{max}) and relative bioavailability (F_{rel}) values.

From ► **Table 3** it is clear that coadministration of the selected *H. hemerocallidea* test materials increased the bioavailability ($AUC_{0-\infty}$) of indinavir in rats compared to the negative control group (indinavir alone), albeit not statistically significantly ($p \geq 0.05$). In accordance with the *in vitro* transport results, where the *H. hemerocallidea* commercial product increased the AP-BL transport, the highest of all three of the selected materials, it also exhibited the highest enhancement of indinavir bioavailability

► **Table 3** Biopharmaceutical parameters for indinavir administered to rats in the absence and presence of the selected *H. hemerocallidea* materials.

Experimental group	AUC _{0-∞} (ng*min/mL) [#]	C _{max} (ng/mL) [#]	F _{rel}
Indinavir (40 mg/kg) alone (negative control)	1371.0 ± 389.8	1108.6 ± 484.3	1.00
Indinavir (40 mg/kg) with verapamil (9 mg/kg, positive control)	1501.3 ± 462.2	644.1 ± 488.8	1.10
Indinavir (40 mg/kg) with <i>H. hemerocallidea</i> aqueous extract (15 mg/kg)	1458.3 ± 512.6	813.6 ± 563.5	1.06
Indinavir (40 mg/kg) with <i>H. hemerocallidea</i> commercial product (15 mg/kg)	1564.4 ± 638.2	928.0 ± 493.0	1.14
Indinavir (40 mg/kg) with <i>H. hemerocallidea</i> reference plant material (15 mg/kg)	1456.6 ± 316.3	1022.0 ± 412.0	1.06

AUC_{0-∞} = area under the curve, C_{max} = maximum plasma concentration, F_{rel} = relative bioavailability. [#]Average ± standard deviation; n = 5

(AUC_{0-∞}) in the rats during the acute study. Overall, the bioavailability results from the acute study correlated well with the *in vitro* transport results in terms of efflux inhibition as expressed by the efflux ratio results (i.e., ER = 1.42 for the commercial product compared to ER = 2.37 and ER = 2.61 for the aqueous extract and reference material, respectively).

The pharmacokinetic effects of *H. hemerocallidea* materials on indinavir bioavailability in this study can therefore be attributed to the inhibition of efflux, and the extent of inhibition was dependent on the type of *H. hemerocallidea* material. As mentioned before, modulation of metabolism can also play a role in interactions between *H. hemerocallidea* and indinavir, but this was not investigated in this study. The rat serves as a model for certain cytochrome P450 enzymes (e.g., CYP1A2 and CYP2E1), but not for CYP3A4 [27], which is the most prevalent enzyme for indinavir metabolism in humans.

It can be deduced from this study that concurrent use of *H. hemerocallidea* products or extracts by people living with HIV/AIDS that are on antiviral treatment (i.e., indinavir) can cause pharmacokinetic interactions that can lead to increased blood levels of indinavir. However, in order to determine if this effect is clinically significant, a follow-up study is needed in an animal model that expresses the CYP3A4 enzyme (e.g., pig model) in order to determine the combined effect of efflux inhibition and metabolism inhibition on the bioavailability of indinavir.

Materials and Methods

Plant materials, chemical compounds, cell line, and growth media

The dried *H. hemerocallidea* bulb material (i.e., biomass reference material) was purchased from ChromaDex (sample HHC 001 stored at the Department of Pharmaceutics, North-West University), which was provided with a certificate of analysis as authenticated with high-performance thin-layer chromatography. The solid oral commercial product (Hypoxis VegeCaps, batch nr. HYO001, sample HHC 002 stored at the Department of Pharmaceutics, North-West University) containing 300 mg *H. hemerocallidea* plant tuber material (total mass of capsule content was 403.74 mg) was purchased from a local health shop. The aqueous extract was prepared from *H. hemerocallidea* corms (voucher HH 064 retained at the Department of Pharmaceutical Sciences,

Tshwane University of Technology) by weighing approximately 5 g of dried *H. hemerocallidea* corm material accurately in a 50-mL Erlenmeyer flask. A volume of 10 mL of deionized water was added and the mixture was sonicated at 45 °C for 30 min. After filtering the mixture through a filter paper (No. 4, Whatman), the filtrate was kept aside and the residue was returned to the flask. This process was repeated twice, where afterwards the filtrates were combined and the residue discarded. The water filtrate was frozen and subsequently freeze-dried overnight, and the extract was stored in a desiccator. All three of the *H. hemerocallidea* materials were chemically characterised by means of liquid chromatography linked to mass spectrometry (LC/MS) as described below in the next section.

Crixivan capsules (MSD, batch nr. w076460) containing 400 mg indinavir per capsule (total mass of capsule content was 658 mg) was purchased from a local pharmacy (sample HHC 003 stored at the Department of Pharmaceutics, North-West University). Verapamil was donated by Sandoz (South Africa) with a certificate of analysis stating a purity of 99.3% (sample HHC 004 stored at the Department of Pharmaceutics, North-West University).

The Caco-2 cell line was obtained from the European Collection of Cell Cultures (ECACC). The high-glucose DMEM, penicillin, and streptomycin were supplied by Separations. Whitehead Scientific supplied the nonessential amino acids, L-glutamine, and Trypsin-Versene mixture, while foetal bovine serum and amphotericin B were obtained from The Scientific Group.

Chemical characterisation of the *Hypoxis hemerocallidea* materials by means of LC/MS

Chemical fingerprinting as well as the determination of the quantity of hypoxoside in each of the three selected *H. hemerocallidea* materials was done by means of LC/MS. The chromatographic analysis was performed on a Waters Acquity UHPLC system with a photo diode array (PDA) detector (Waters). UHPLC separation was attained on an Acquity BEH C₁₈ column (150 mm × 2.1 mm, 1.7 µm particle size; Waters), which was maintained at 40 °C during analysis. The mobile phase consisted of 0.1% v/v formic acid in water (solvent A) and acetonitrile (solvent B) with a flow rate of 0.3 mL/min. A gradient elution was employed as follows: the analysis started with a 85% A:15% B mixture, which was changed to a mixture of 65% A:35% B in 7 min and then to 50% A:50% B in 1 min, which was maintained for 0.5 min after which it was changed back to the initial mixture in 0.5 min. The total running

time was 11 min. The standard and samples were injected into the mobile phase with an injection volume of 1.0 μL (full-loop injection). Mass spectrometry (MS) was operated in the negative ion electrospray mode. The desolvation gas was nitrogen (N_2), while the desolvation temperature was set to 350 $^\circ\text{C}$ at a flow rate of 500 L/h and the source temperature was set to 100 $^\circ\text{C}$. The capillary and cone voltages were set to 2500 V and 45 V, respectively. The analytical data were collected between 100 and 1000 m/z .

Bidirectional *in vitro* permeability across Caco-2 cell monolayers

The Caco-2 cells were cultured in high-glucose DMEM supplemented with 10% foetal bovine serum, 1% nonessential amino acids (NEAA), 1% penicillin/streptomycin, 1% 2 mM L-glutamine, and 1% amphotericin B (250 $\mu\text{g}/\text{mL}$). Culturing of the cells occurred at a temperature of 37 $^\circ\text{C}$ with 5% carbon dioxide and 95% humidified air in a Galaxy 170R incubator (Eppendorf). The growth medium was exchanged every second day under sterile conditions in a laminar flow hood. The cells were examined by means of a light microscope (Nikon Eclipse TS100/TS100F, Nikon Instruments) prior to exchange of the growth medium. The percentage confluency was estimated and the absence of any contamination was ensured. Subculturing by means of trypsinisation took place once the cells reached a confluency of 50–60%.

Caco-2 cells (passage 50) were seeded onto Transwell 6-well membrane filters (Corning Costar Corporation) with a surface area of 4.67 cm^2 and a pore diameter of 0.4 μm . The cell suspension was obtained through trypsinisation by adding a Trypsin-versene mixture and then incubated for 5 min. After cell detachment, 6 mL of pre-warmed growth medium were added to the flask. The cell suspension was extensively agitated with a pipette to ensure complete cell detachment and deagglomeration in order to form a suspension consisting of single cells. This single cell suspension was then transferred to a 50-mL tube. A Pasteur pipette was used to agitate the cell suspension to make sure that a homogenous cell distribution was present. A haemocytometer was used to count the cells in the suspension after Trypan blue was added. The average number of cells per square was calculated and this number was then multiplied with the dilution factor (5×10^4) in order to establish the total number of cells per mL in the cell suspension. The cell suspension was diluted to a concentration of 20 000 cells per mL, and a volume of 2.5 mL was pipetted into each apical chamber of the membrane filter plate wells. The growth medium was exchanged every second day and the cells were cultured for 21–24 days until intact epithelial monolayers were obtained.

The transepithelial electrical resistance (TEER) of each Caco-2 cell monolayer was measured with a Millcell ERS II meter (Millipore) prior to the commencement of the transport studies. Measurement of TEER is one method that is used to establish tight junction integrity of Caco-2 cell monolayers and a value of $\geq 150 \Omega$ is considered sufficient [28]. In this study, a TEER reading of higher than 250 Ω (equivalent to 1167.5 Ω/cm^2) was required before commencement of the transport studies [29]. The TEER was measured again at the end of the study to ensure cell monolayer integrity was maintained during the transport study.

To prepare a 200- μM indinavir solution (negative control), the correct equivalent mass of a Crixivan capsule content (i.e., 2.02 mg) was weighed and added to 10 mL DMEM and stirred until dissolved. For the test solutions, a sufficient amount of each of the selected *H. hemerocallidea* materials was added to the indinavir solution to reach a concentration of 500 $\mu\text{g}/\text{mL}$ for each of the *H. hemerocallidea* materials [30]. A 100- μM verapamil solution was prepared by dissolving 0.45 mg verapamil into 5 mL in the indinavir solution. For the transport in the AP-BL direction, the growth medium was removed from the basolateral chamber and replaced with a volume of 2.5 mL preheated transport medium (DMEM) buffered with HEPES. The Transwell plate was then kept in the incubator for 30 min before the growth medium was removed from the apical chambers and replaced with 2.5 mL of the test solutions. Samples (200 μL) were withdrawn from the basolateral chambers at time intervals of 20, 40, 60, 80, 100, and 120 min, and were replaced with an equal volume of preheated transport buffer. Samples were analysed by a previously described HPLC analysis method [29] to determine the concentration of indinavir that appeared in the acceptor chamber over time.

For the transport in the BL-AP direction, the same method was used, except that the growth medium was removed from the basolateral chambers and replaced with 2.5 mL of each test solution. Samples (200 μL) were withdrawn from the apical chambers at the time intervals of 20, 40, 60, 80, 100, and 120 min and immediately replaced with an equal volume of preheated DMEM.

Indinavir alone (200 μM) served as the negative control group, while indinavir in combination with 100 μM verapamil, a known P-gp inhibitor, served as the positive control group. The test solutions consisting of the selected *H. hemerocallidea* materials (500 $\mu\text{g}/\text{mL}$) in combination with indinavir (200 μM) formed the experimental groups. All transport experiments were conducted in triplicate.

The P_{app} values for the transport of indinavir was calculated according to the following equation [29]:

$$P_{\text{app}} = \frac{dQ}{dt} \left(\frac{1}{A \cdot C_0 \cdot 60} \right) \quad (1)$$

Where P_{app} is the apparent permeability coefficient (cm/s), dQ/dt is the permeability rate (amount permeated per minute), A is the diffusion area of the membrane (cm^2), and C_0 is the initial concentration of the model drug.

The ER value indicates any asymmetry in the directional transport of indinavir in combination with the different *H. hemerocallidea* materials. The ER value was calculated according to the following equation:

$$\text{ER} = \frac{P_{\text{app}} (B - A)}{P_{\text{app}} (A - B)} \quad (2)$$

Where $P_{\text{app}} (B-A)$ is the permeability coefficient for the permeation in the basolateral-to-apical direction and $P_{\text{app}} (A-B)$ is the permeability coefficient for the permeation in the apical-to-basolateral direction.

In vivo bioavailability studies

The *in vivo* pharmacokinetic study in Sprague Dawley rats [obtained from South African Vaccine Producers (SAVP)] was approved by the Faculty of Health Sciences Animal Ethics Committee of the University of Cape Town, South Africa (reference number: 015/041, date of approval: 17/03/2016). All animal experiments were implemented and conducted according to national and international accepted principles and standards for humane handling of animals. A total of 25 male Sprague Dawley rats weighing 250–300 g was randomly selected and divided into 5 different groups as summarised in ► Fig. 4.

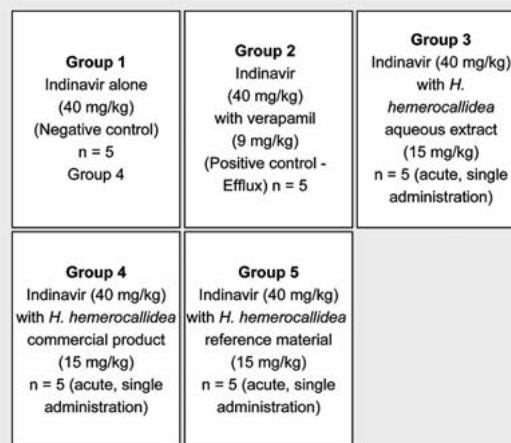
The test solutions consisted of the selected *H. hemerocallidea* materials at a concentration of 15 mg/kg coadministered with indinavir at a concentration of 40 mg/kg (based on equivalent Crixivan capsule content), while indinavir (40 mg/kg, based on equivalent Crixivan capsule contents) was administered alone as the negative control group. The positive control for efflux inhibition consisted of indinavir (40 mg/kg) with verapamil (9 mg/kg) [31]. The test solutions were administered by means of oral gavage at a volume of 500 µL per animal.

Blood samples (200 µL) were collected from the tail veins of the animals at predetermined time intervals of 0, 0.5, 1, 2, 4, 8, and 24 h after administration of each experimental and control test solution. Sample tubes were sprayed with heparin beforehand, which served as an anticoagulant. The blood samples were then centrifuged for 8 min at 14 000 rpm, after which the plasma was recovered from each blood sample. The plasma samples were kept at –80 °C until the indinavir analysis occurred.

A previously described LC/MS/MS method for the analysis of indinavir was used in order to analyse the plasma samples for indinavir concentration with slight modifications as outlined below [32].

Briefly, the liquid chromatography method entailed using a Kinetex F5 (4.6 × 100 mm, 2.6 µm) analytical column on an Agilent 1100 series HPLC. The mobile phase consisted of a mixture of A and B at 50:50 v/v, where mobile phase A was 0.1% formic acid in water and B was 0.1% formic acid in acetonitrile. The mobile phase was delivered at a constant flow rate of 500 µL/min. The column was kept in a column compartment at 40 °C. An autosampler injected 10 µL onto the HPLC column. The injection needle was rinsed with mobile phase before each injection for 30 s using the flush port wash station. The samples were cooled to 4 °C while awaiting injection.

Detection of indinavir and the internal standard (indinavir-*d*₆) was performed on an AB Sciex API 3200 mass spectrometer [electrospray ionisation (ESI) in the positive ion mode] and the settings on the apparatus are summarised in a previously published method [31]. Furthermore, a stock solution of indinavir was prepared in DMSO at a concentration of 1 mg/mL. Sprague-Dawley blank plasma samples were spiked with the stock solution to obtain standard 1 (STD 1) at a concentration of 2000 ng/mL. Dilution of this solution with blank plasma resulted in STD 2 (500 ng/mL), STD 3 (125 ng/mL), STD 4 (32 ng/mL), STD 5 (8 ng/mL), and STD 6 (2 ng/mL). Quality control samples were also prepared in the same pool of rat plasma at 1600 ng/mL, 400 ng/mL, 50 ng/mL, 10 ng/mL, and 2 ng/mL. The calibration standards and quality control samples were briefly vortexed and aliquoted into labelled poly-



► Fig. 4 Layout of the *in vivo* pharmacokinetic study design in Sprague Dawley rats.

propylene tubes, which were stored at –80 °C until analysis with LC/MS/MS. Samples above the upper limit of quantification were diluted 4 times with blank plasma and reanalysed in a repeat batch [32]. The method was validated in terms of linearity, accuracy, and precision.

The extraction was done by adding 200 µL of ice-cold 0.1% formic acid in methanol containing 20 ng/mL internal standard (indinavir-*d*₆) to a 20-µL sample of plasma, which was vortex mixed for 60 s. This was followed by ultrasonication for 10 min and centrifugation at 10 000 rpm for 10 min. Then, 180 µL of the supernatant were transferred into clean culture tubes and evaporated to dryness under nitrogen gas at 40 °C. The residue was reconstituted with 100 µL of 0.1% formic acid in water, and vortex mixed for 60 s. The extracts were transferred into a 96-well plate for injection. A volume of 10 µL of the reconstituted extract solutions was injected onto the column.

WinNonlin software (Pharsight Corporation) was used in order to construct the indinavir bioavailability profiles. Relevant pharmacokinetic parameters (i.e., C_{max} and $AUC_{0-\infty}$) were obtained. The F_{rel} of indinavir was calculated by using the following equation:

$$F_{rel} = \frac{[AUC_{0-\infty}]_A}{[AUC_{0-\infty}]_B} \quad (3)$$

Where $[AUC]_A$ represents the area under the curve for indinavir in the presence of the experimental material (i.e., each selected *H. hemerocallidea* material) and $[AUC]_B$ represents the area under the curve for indinavir alone.

Statistical analysis

Data analyses were performed with STATISTICA Ver 12. ANOVA's with Tukey's honest significant post hoc tests were performed, and statistically significant differences were accepted when $p < 0.05$. All results were verified with nonparametric Kruskal-Wallis and Dunn's post hoc tests.

Supporting information

Indinavir mean plasma concentrations at each time point after oral administration to rats in the absence and presence of the selected *H. hemerocallidea* materials are available as Supporting Information.

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Conflict of Interest

The authors declare no conflict of interest.

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