

# The *N*-Methyl-(2*S*, 4*R*)-*trans*-4-hydroxy-L-proline-Enriched Methanol Fraction from *Sideroxylon obtusifolium* Shows an Anticonvulsant Activity Associated with its Anti-inflammatory/Antioxidant Actions



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## Key words

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## ABSTRACT

Epilepsy is a neurological disorder characterized by recurrent seizures, resulting from excessive neuronal discharges. *Sideroxylon obtusifolium* is used in Brazil for its anti-inflammatory/antioxidant properties, known to be involved with epilepsy. The anticonvulsant effects of the methanol fraction from *S. obtusifolium* leaves, rich in *N*-methyl-(2*S*,4*R*)-*trans*-4-hydroxy-L-proline, were investigated on pilocarpine- and pentylenetetrazole-induced convulsion models. Mice were pretreated with *N*-methyl-(2*S*,4*R*)-*trans*-4-hydroxy-L-proline (50, 100, 200 mg/kg, p.o.) and, 1 h later, by pilocarpine (400 mg/kg, i.p.) or pentylenetetrazole (80 mg/kg, i.p.). The animals were observed for latency to the first convulsion and latency to death. Immediately after death, brain areas from the pilocarpine groups were harvested for biochemical measurements. The latency to the first convulsion and latency to death increased after *N*-methyl-(2*S*,4*R*)-*trans*-4-hydroxy-L-proline treatment compared with the pilocarpine- or pentylenetetrazole-only groups. In both convulsion models, sodium valproate (reference drug) was used as a positive control. Additionally, the decreases in striatal dopamine and 3,4-dihydroxyphenylacetic acid contents observed in the pilocarpine-only group were partially prevented in the *N*-methyl-(2*S*,4*R*)-*trans*-4-hydroxy-L-proline-treated groups. While brain gamma-aminobutyric acid and glutamate contents decreased and increased, respectively, after pilocarpine only, these changes were also prevented by *N*-methyl-(2*S*,4*R*)-*trans*-4-hydroxy-L-proline. Similarly, *N*-methyl-(2*S*,4*R*)-*trans*-4-hydroxy-L-proline reduced the brain oxidative stress by decreasing the levels of nitrite and lipid peroxidation and increasing the glutathione content of the pilocarpine-only group. The increases in hippocampal expressions for interleukin 6, interferon-gamma, and glial fibrillary acidic protein, after pilocarpine only, were decreased to normal levels by *N*-methyl-(2*S*,4*R*)-*trans*-4-hydroxy-L-proline. In conclusion, the study showed significant anticonvulsant effects for *N*-methyl-(2*S*,4*R*)-*trans*-4-hydroxy-L-proline, probably related to its anti-inflammatory/antioxidant properties. *N*-Methyl-(2*S*,4*R*)-*trans*-4-hydroxy-L-proline effects were potentiated by VPA (sodium valproate), thus it may also interact with the GABAergic system, as we had recently shown.

## ABBREVIATIONS

DA	dopamine
DG	dentate gyrus
DOPAC	3,4-dihydroxyphenylacetic acid
DTNB	5,5'-dithio-bis-(2-nitrobenzoic acid)
GABA	gamma-aminobutyric acid
GFAP	glial fibrillary acidic protein
GLU	glutamate
GSH	glutathione
IFN- $\gamma$	interferon-gamma
IL-6	interleukin 6
LPS	lipopolysaccharide
MDA	malondialdehyde
MRI	magnetic resonance image
NMP	<i>N</i> -methyl-(2 <i>S</i> , 4 <i>R</i> )- <i>trans</i> -4-hydroxy-L-proline
PFC	prefrontal cortex
Pilo	pilocarpine
PTZ	pentylentetrazole
ROS	reactive oxygen species
TBARS	thiobarbituric acid reactive substances
TC	temporal cortex
TLE	temporal lobe epilepsy
VPA	sodium valproate

## Introduction

According to the National Institute of Neurological Disorders and Stroke (USA), epilepsies are a spectrum of brain disorders, ranging from severe, life-threatening, and disabling to more benign forms. In epilepsy, the normal pattern of neuronal activity becomes disturbed and the individual may show convulsions, muscle spasms, and loss of consciousness. Epilepsy affects 65 000 000 people worldwide and can be a major burden in seizure-related disabilities, mortality, and comorbidities [1]. TLE is the most common and prevalent refractory form of adult focal epilepsy [2]. It involves recurrent seizures, arising in the mesial structures of the hippocampus, amygdala, and entorhinal cortex and, thus, it is better named mesial temporal lobe epilepsy [3, 4].

The main characteristics of TLE are epileptic foci in the limbic system, an initial precipitating injury, the latent period, and the presence of hippocampal sclerosis. All these features are reproduced in rodent models of epilepsy by the systemic injection of high doses of Pilo [5]. Therefore, the Pilo model of epilepsy is a valuable tool for studying the pathogenesis of TLE and evaluating potential antiepileptogenic drugs [6]. Furthermore, the understanding of the disease pathogenesis relies largely on the use of animal models.

*Sideroxylon obtusifolium* (Humb. ex Roem. & Schult.) T.D. Penn. belongs to the Sapotaceae family and is a medicinal plant used in Brazil due to its analgesic and anti-inflammatory activities, as already demonstrated by us [7, 8] and others [9]. Furthermore, experimental and clinical evidence indicates that there is a close relationship between epileptogenesis and brain inflammation. Epileptic seizures increase inflammatory mediators in the brain and

the likelihood of recurrent seizures [10]. Additionally, brain neuronal hyperexcitability and oxidative injury produced by free radicals may play a role in the initiation and progression of epilepsy [11].

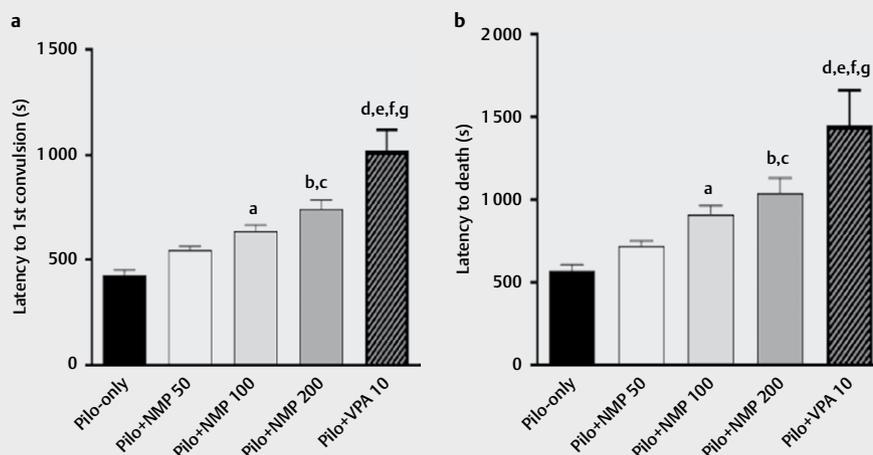
Thus, considering the involvement of oxidative stress and neuroinflammation in epileptic seizures, the objectives of the present work were to investigate the possible anticonvulsant properties of the L-proline derivative NMP in models of Pilo- and PTZ-induced convulsions in mice. This compound is present in the decoction and mainly in the methanol fraction from the leaves of *S. obtusifolium*. In addition, there is also evidence for the involvement of dopaminergic neurotransmission [12, 13] and increased astrocyte activation [14, 15] in these experimental models of convulsions. Thus, we also focused on behavioral, biochemical, and immunohistochemical analyses to detect the possible benefits of NMP, mainly on the Pilo-induced convulsive processes and epilepsy.

## Results

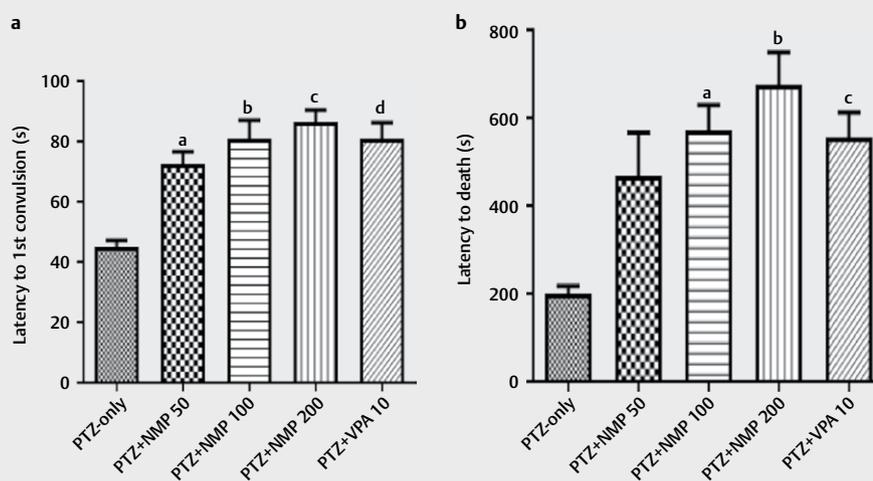
In Pilo-induced seizures, a model of TLE, two behavioral parameters were analyzed: the latency to the first convulsion and the latency to death. While no significant change was noticed with the lowest NMP dose (50 mg/kg, p.o.), we showed 1.5 and 1.7 times increases in the latency to the first convulsion after treatment with the doses of 100 and 200 mg/kg, respectively, relative to the untreated control (Pilo-only group). In addition, 2.4 times increases were seen in the VPA group (reference group) compared with the control group [ $F(4,72) = 24.09$ ,  $p < 0.0001$ ]. Similarly, while no significant change was observed with the dose of 50 mg/kg, significant increases (1.6 and 1.8 times, respectively) were observed in the latency to death after treatments with the two higher NMP doses (100 and 200 mg/kg, respectively) compared to the control group. Also, increases of 2.5 times were noticed in the VPA group compared with the control [ $F(4,60) = 17.0$ ,  $p < 0.0001$ ] (► **Fig. 1**). Another experiment using the same model but a lower Pilo dose (350 mg/kg, i.p.) was performed after a 7-day daily treatment with NMP, and the results on the latency to the first convulsion and the latency to death were similar to those above (data not shown).

In the PTZ-induced seizure model, increases in the latency to the first convulsion ranging from 1.6 to 1.9 times were observed after acute treatments with NMP at doses of 50, 100, and 200 mg/kg, p.o., while the VPA-treated group showed an increase of 1.8 times compared with the control group (PTZ only) [ $F(4,76) = 15.0$ ,  $p < 0.0001$ ]. Increases in the latency to death, ranging from 2.3 to 3.4 times, were observed after treatments with NMP (50, 100, 200 mg/kg, p.o.) when compared with PTZ only [ $F(4,63) = 10.57$ ,  $p < 0.0001$ ] (► **Fig. 2**). The VPA-treated group showed a 2.8 times increase. In another experiment, we demonstrated that the effect of a lower dose of NMP (25 mg/kg) was potentiated by VPA, suggesting that the GABAergic system might be a pharmacological target for NMP (data not shown).

The dopaminergic system is a neuromodulatory system in the brain and has a significant effect on neuronal excitability. It is known to change in the epileptic brain [16]. The Pilo-only group showed a 63% decrease in striatal DA contents and this decrease was significantly attenuated (41, 19, 31%) in the striata after treatments



► **Fig. 1** Effects of the methanol fraction, rich in NMP, from *S. obtusifolium* on the latency to the 1<sup>st</sup> convulsion and latency to death in the model of Pilo-induced convulsions in mice. Groups tested (6 to 20 animals): Pilo only, Pilo + NMP treated, and Pilo + VPA treated. **a** Latency to the 1<sup>st</sup> convulsion (a) vs. Pilo only,  $p=0.0003$ ; (b) vs. Pilo only,  $p<0.0001$ ; (c) vs. Pilo + NMP 50,  $p=0.0016$ ; (d) vs. Pilo only,  $p<0.0001$ ; (e) vs. Pilo + NMP 50,  $p<0.0001$ ; (f) vs. Pilo + NMP 100,  $p<0.0001$ ; (g) vs. Pilo + NMP 200,  $p=0.0006$ . **b** Latency to death (a) vs. Pilo only,  $p=0.0045$ ; (b) vs. Pilo only,  $p<0.0001$ ; (c) vs. Pilo + NMP 50,  $p<0.0069$ ; (d) vs. Pilo only,  $p<0.0001$ ; (e) vs. Pilo + NMP 50,  $p<0.0001$ ; (f) vs. Pilo + NMP 100,  $p=0.0005$ ; (g) vs. Pilo + NMP 200,  $p=0.0116$  (one-way ANOVA and Tukey multiple comparisons test).

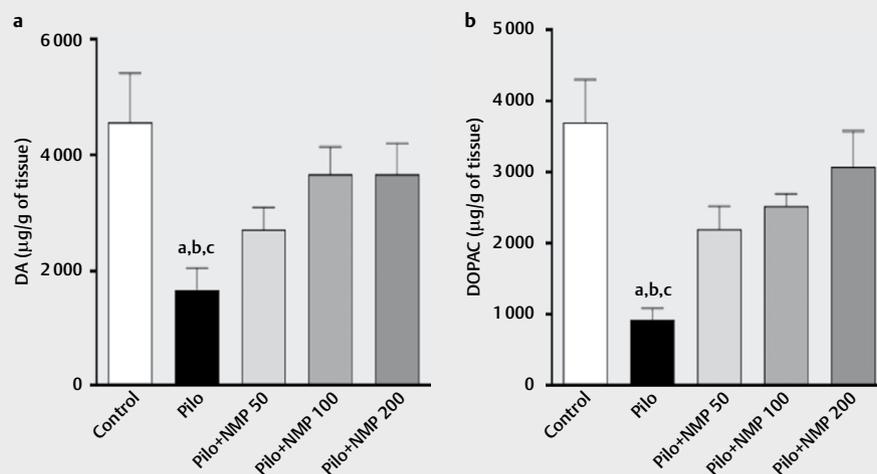


► **Fig. 2** Effects of the methanol fraction, rich in NMP, from *S. obtusifolium* on the latency to the 1<sup>st</sup> convulsion and latency to death in the model of PTZ-induced convulsions in mice. Groups tested (6 to 22 animals): PTZ only (control), PTZ + NMP treated, and PTZ + VPA treated. **a** Latency to the 1<sup>st</sup> convulsion (a) vs. PTZ only,  $p=0.0023$ ; (b) vs. PTZ only,  $p<0.0001$ ; (c) vs. PTZ only,  $p=0.0001$ ; (d) vs. PTZ only,  $p=0.0002$ . **b** Latency to death (a) vs. PTZ only,  $p<0.0001$ ; (b) vs. PTZ only,  $p<0.0001$ ; (c) vs. PTZ only,  $p=0.0150$  (one-way ANOVA and Tukey multiple comparisons test).

with NMP at doses of 50, 100, and 200 mg/kg, respectively [ $F(4,38)=4.85$ ,  $p=0.0029$ ]. A similar result was demonstrated for DOPAC contents, with a 75% decrease in the Pilo-only group, but only 41, 32, and 23% decreases after NMP treatments with doses of 50, 100, and 200 mg/kg, *p.o.*, respectively [ $F(4,43)=8.278$ ,  $p<0.0001$ ] (► **Fig. 3**).

GABA and GLU are the main inhibitory and excitatory brain amino acids, respectively, thus, as such, very important for seizure development. We showed high GABA level decreases in the brain

areas tested (PFC, hippocampus, and striatum) in the Pilo-only group in relation to normal controls. Thus, while 62 to 65% decreases were observed in the PFC [ $F(3,15)=4.773$ ,  $p=0.0157$ ] and striatum [ $F(3,15)=7.399$ ,  $p=0.0029$ ], an even higher decrease (84%) was demonstrated in the hippocampus [ $F(3,17)=5.35$ ,  $p=0.0087$ ]. These changes were reversed after NMP treatments with doses of 100 and 200 mg/kg. On the contrary, in the Pilo-only group, significant increases were observed in GLU contents (5.7, 4.7, and 6.6 times increases, respectively) in the PFC [ $F(3,18)=15.29$ ,  $p<0.0001$ ],



► **Fig. 3** Effects of the methanol fraction, rich in NMP, from *S. obtusifolium* on the striatal contents of DA and its metabolite DOPAC in mice subjected to Pilo-induced convulsions. Groups tested (3 to 13 animals): control (normal), Pilo only, and Pilo + NMP treated. **a** DA (a) vs. control,  $p < 0.05$ ; (b) vs. Pilo only,  $p < 0.05$ ; (c) vs. Pilo only,  $p < 0.05$ . **b** DOPAC (a) vs. control,  $p < 0.001$ ; (b) vs. Pilo only,  $p < 0.05$ ; (c) vs. Pilo only,  $p < 0.01$  (one-way ANOVA and Tukey multiple comparisons test).

hippocampus [ $F(3,17) = 23.68$ ,  $p < 0.0001$ ], and striatum [ $F(3,18) = 21.63$ ,  $p < 0.0001$ ] compared to the normal controls. These changes were partially reversed after NMP treatments in all three areas (► **Fig. 4**).

Nitrite measurements are used to indicate oxidative stress resulting from excessive release of free radicals. We determined brain nitrite content as an index of oxidative stress in PFC [ $F(3,20) = 6.123$ ,  $p = 0.0040$ ], hippocampus [ $F(3,22) = 6.588$ ,  $p = 0.0024$ ], and striatum [ $F(3,20) = 68.87$ ,  $p < 0.0001$ ]. Our data showed increases ranging from 1.7 to 1.9 times in these areas in the Pilo-only group compared with the normal controls. The values were almost or completely normalized after NMP treatments (100 and 200 mg/kg) in all groups (► **Fig. 5**).

Lipid peroxidation is the oxidative degradation of lipids proceeding as a free radical chain reaction mechanism. We showed increases from 1.4 to 1.5 times in the lipid peroxidation index in the Pilo-only group compared with normal controls in all brain areas tested: PFC [ $F(3,53) = 13.78$ ,  $p < 0.0001$ ], hippocampus [ $F(3,39) = 6.918$ ,  $p = 0.0008$ ], and striatum [ $F(3,32) = 6.965$ ,  $p = 0.0010$ ]. The values returned to those of normal controls after treatment with NMP at the doses of 100 and 200 mg/kg (► **Fig. 6**).

GSH is considered a very important brain antioxidant for mammals and is essential to the cellular detoxification of ROS in brain cells [16]. We showed significant decreases in brain GSH content, ranging from 38 to 47% in the Pilo-only group relative to the normal controls: PFC [ $F(3,49) = 5.508$ ,  $p = 0.0024$ ], hippocampus [ $F(3,51) = 10.54$ ,  $p < 0.0001$ ], and striatum [ $F(3,48) = 6.017$ ,  $p = 0.0015$ ]. The highest decrease was observed in the PFC area. The values from all NMP-treated groups were not significantly different from those of normal controls (► **Fig. 7**).

Evidence has indicated that epileptic seizures can induce cytokine production, thus influencing the epileptic process [17]. Increases of 4 times in the IL-6 values [ $F(3,24) = 14.67$ ,  $p < 0.0001$ ]

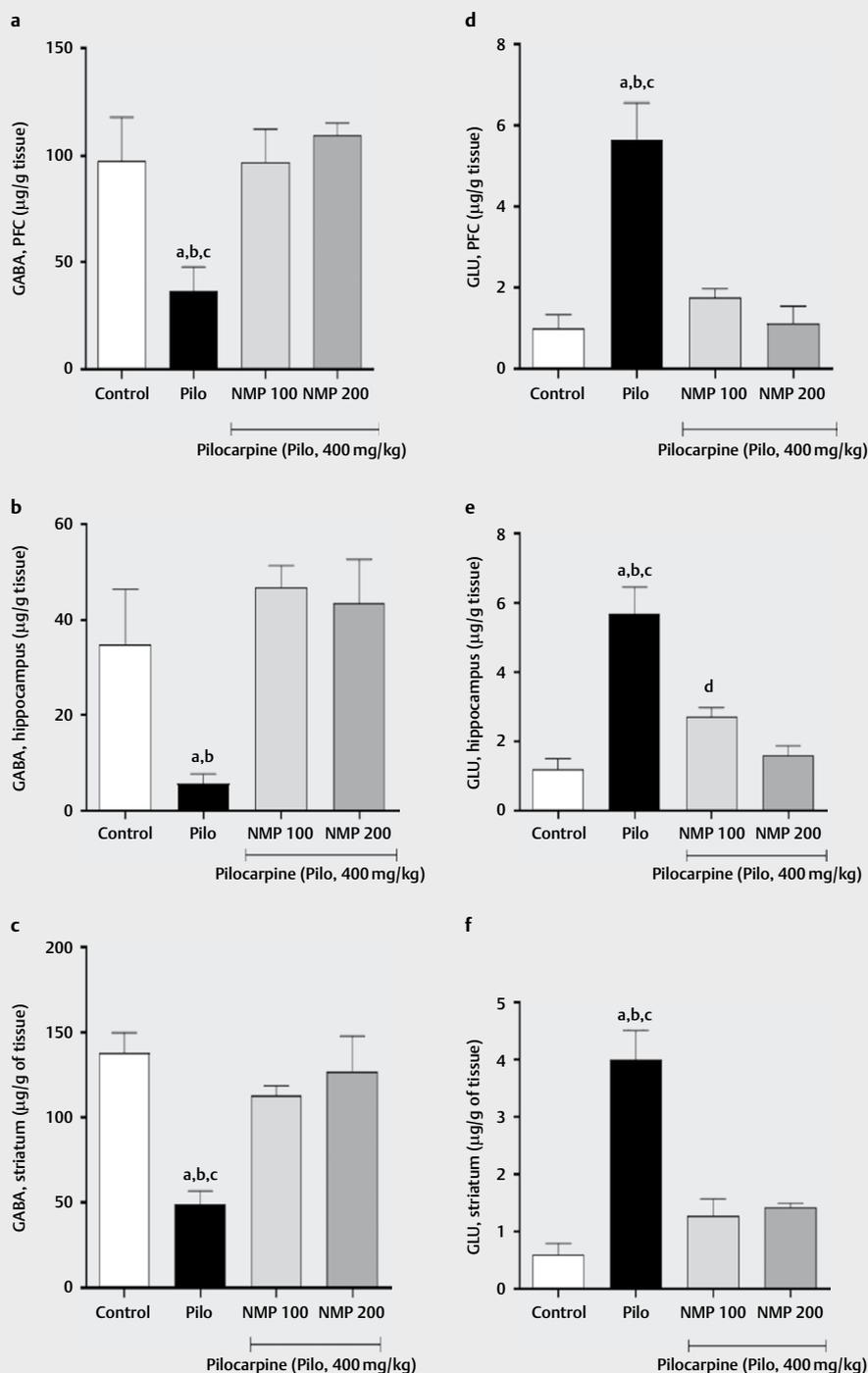
and 2 times in the IFN- $\gamma$  values [ $F(3,27) = 7.337$ ,  $p = 0.0009$ ] were observed in the hippocampus of the Pilo-only group compared with normal controls, values which were completely normalized after NMP treatments at doses of 100 and 200 mg/kg (► **Fig. 8**).

Glia cell activation is known to occur following seizures [15]. GFAP immunostaining assays were performed in the hippocampus and TC areas and showed increases ranging from 5 times (CA3 area) to 8 times (CA1 area) in the Pilo-only group compared with normal controls: TC [ $F(3,12) = 34.3$ ,  $p < 0.0001$ ], CA3 [ $F(3,12) = 746$ ,  $p < 0.0001$ ], and CA1 [ $F(3,12) = 190.5$ ,  $p < 0.0001$ ]. The values went down towards those of the controls in the CA3 area after NMP treatment (100 and 200 mg/kg). Increases in the immunoreactivity were even higher in the DG [ $F(3,12) = 34.98$ ,  $p < 0.0001$ ], which presented almost an 11 times increase in the Pilo-only group in relation to normal controls. These increases were attenuated after treatment with NMP 100 mg/kg (around a 3 times increase) and 200 mg/kg (around a 4 times increase) (► **Fig. 9**).

## Discussion

Epilepsy is a neurological disease whose mechanisms involved with seizure generation are not completely understood. Therefore, the search for new antiepileptic drugs for treating around 30% of patients refractory to conventional therapies remains greatly in need. Up to now, the most important action mechanisms for seizure generation focus on the hyperactivity of excitatory amino acid systems, insufficient GABA<sub>A</sub> receptor-mediated neurotransmission, and disturbances in the properties of neuronal membranes [18].

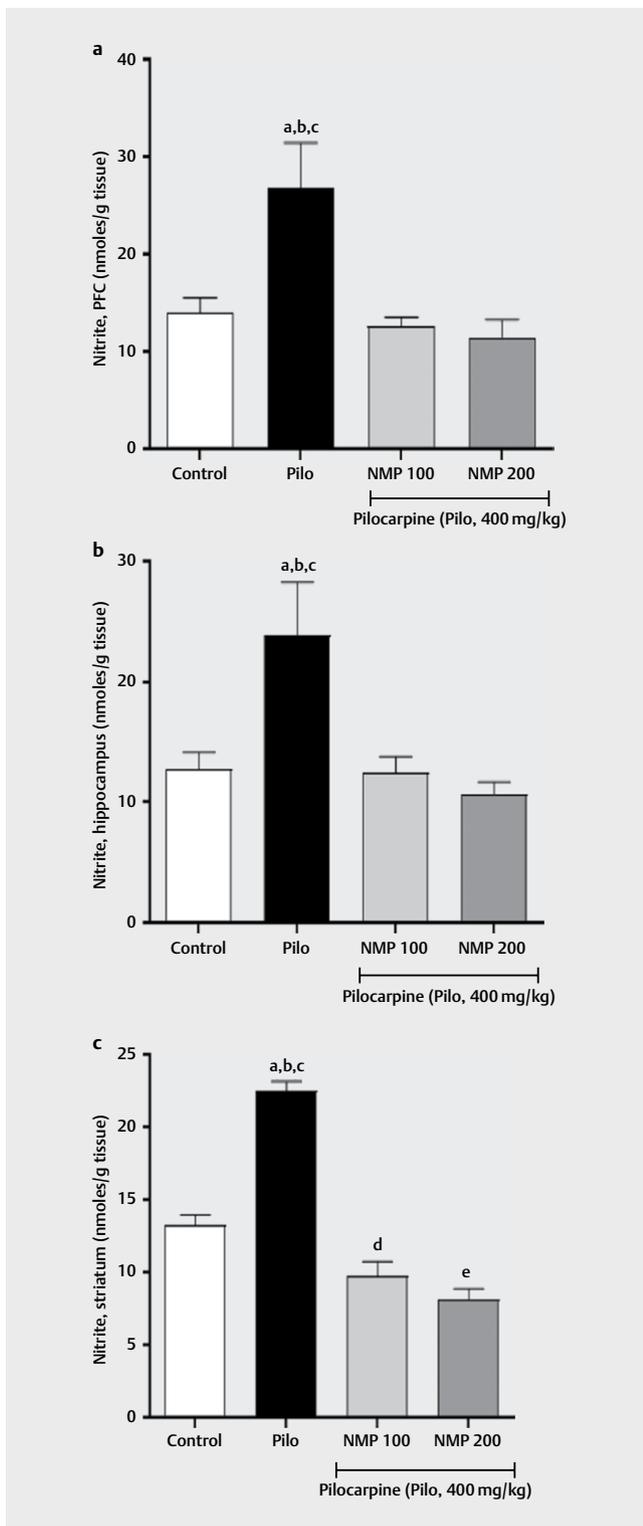
In the present work, we aimed at investigating the anticonvulsant activity of the methanol fraction rich in NMP, an L-proline derivative, and major bioactive constituents present in *S. obtusifolium* leaves. Considering the beneficial properties of *S. obtusifolium* and the increasing evidence for the inflammatory processes involved with con-



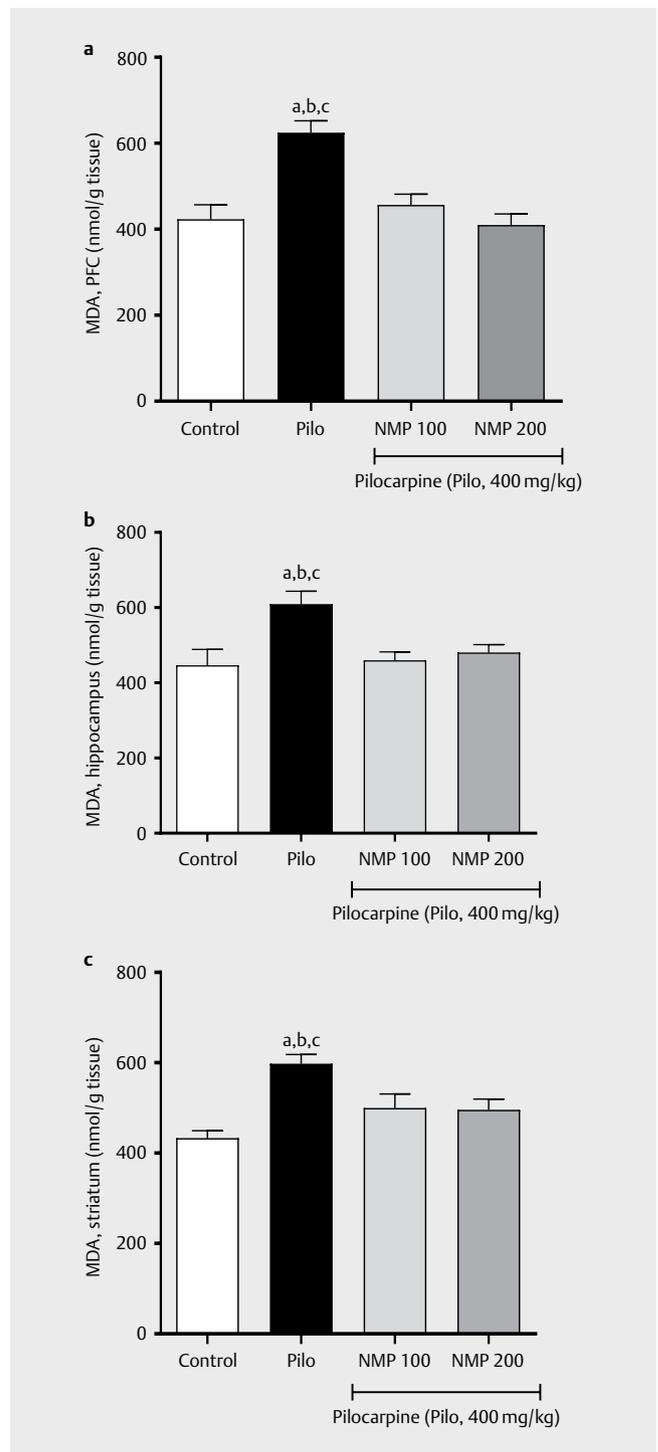
► **Fig. 4** Effects of the methanol fraction, rich in NMP, from *S. obtusifolium* on brain GABA and GLU in mice subjected to Pilo-induced convulsions. Groups tested (4 to 6 animals): control (normal), Pilo only, and Pilo + NMP treated. **a** GABA, PFC (**a**) vs. control,  $p < 0.05$ ; (**b**) vs. Pilo only,  $p < 0.05$ ; (**c**) vs. Pilo only,  $p < 0.05$ . **b** GABA, hippocampus (**a**) vs. Pilo only,  $p < 0.01$ ; (**b**) vs. Pilo only,  $p < 0.05$ . **c** GABA, striatum (**a**) vs. control,  $p < 0.01$ ; (**b**) vs. Pilo only,  $p < 0.05$ ; (**c**) vs. Pilo only,  $p < 0.01$ . **d** GLU, PFC (**a**) vs. control,  $p < 0.001$ ; (**b**) vs. Pilo only,  $p < 0.001$ ; (**c**) vs. Pilo only,  $p < 0.001$ . **e** GLU, hippocampus (**a**) vs. control,  $p < 0.001$ ; (**b**) vs. control,  $p < 0.05$ ; (**c**) vs. Pilo only,  $p < 0.001$ ; (**d**) vs. Pilo only,  $p < 0.001$ . **f** GLU, striatum (**a**) vs. control,  $p < 0.001$ ; (**b**) vs. Pilo only,  $p < 0.001$ ; (**c**) vs. Pilo only,  $p < 0.001$  (one-way ANOVA and Tukey multiple comparisons test).

vulsions and epilepsy [19], we showed that the L-proline derivative (NMP) significantly increased latency to the first convulsion and latency to death in a Pilo-induced convulsion model in mice.

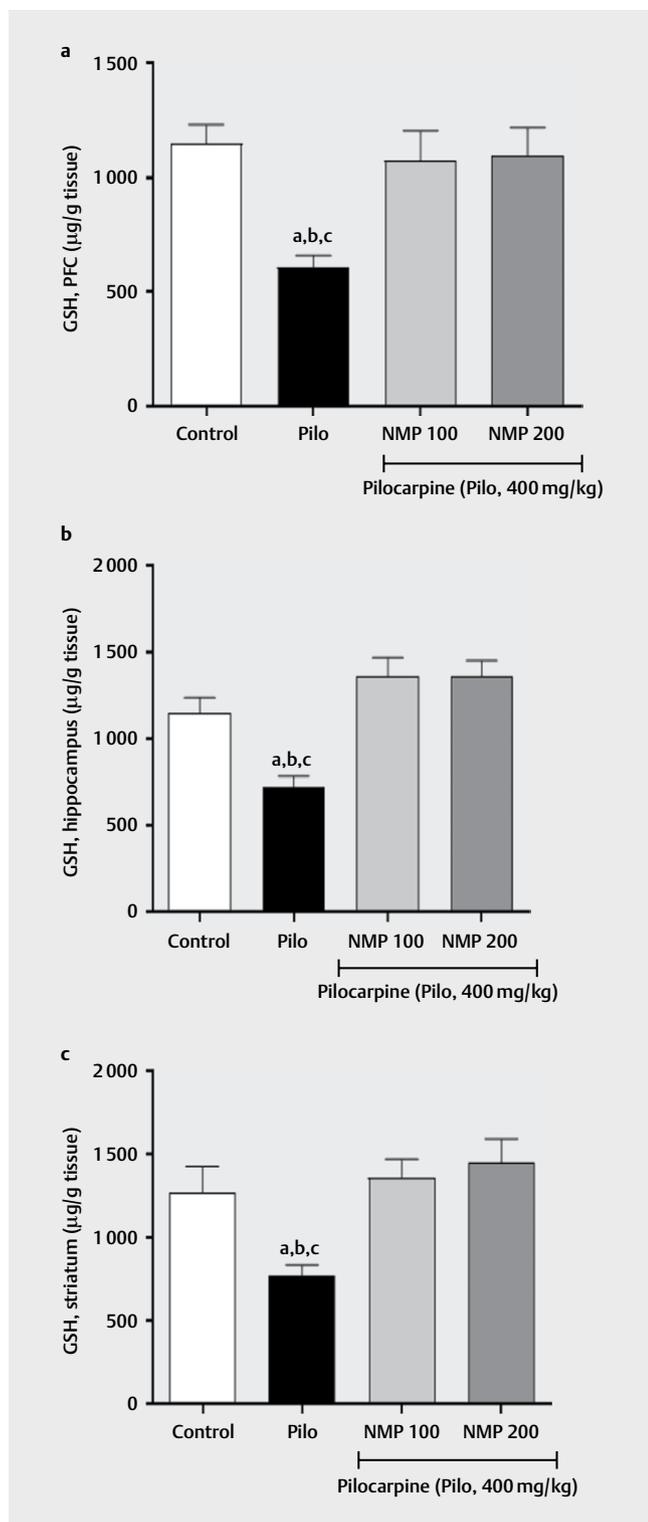
Furthermore, the dopaminergic system has been shown to have a seizure-modulating effect that depends upon the brain dopamine receptor subtypes [12]. Evidence indicates that drugs stimulating



► **Fig. 5** Effects of the methanol fraction, rich in NMP, from *S. obtusifolium* on brain nitrite content in mice subjected to Pilo-induced convulsions. Groups tested (5 to 7 animals): control (normal), Pilo only, and Pilo + NMP treated. **a** PFC (**a**) vs. control,  $p < 0.05$ ; (**b**) vs. Pilo + NMP 100,  $p < 0.05$ ; (**c**) vs. Pilo + NMP 200,  $p < 0.01$ . **b** Hippocampus (**a**) vs. control,  $p < 0.05$ ; (**b**) vs. Pilo + NMP 100,  $p < 0.001$ ; (**c**) vs. Pilo + NMP 200,  $p < 0.01$ . **c** Striatum (**a**) vs. control,  $p < 0.001$ ; (**b**) vs. Pilo + NMP 100,  $p < 0.05$ ; (**c**) vs. Pilo + NMP 200,  $p < 0.001$ ; (**d**) vs. control,  $p < 0.05$ ; (**e**) vs. control,  $p < 0.001$  (one-way ANOVA and Tukey multiple comparisons test).



► **Fig. 6** Effects of the methanol fraction, rich in NMP, from *S. obtusifolium* on brain lipid peroxidation (MDA, nmol/g tissue) in mice subjected to Pilo-induced convulsions. Groups tested (5 to 16 animals): control (normal), Pilo only, and Pilo + NMP treated. **a** PFC (**a**) vs. control,  $p < 0.001$ ; (**b**) vs. Pilo + NMP 100,  $p < 0.001$ ; (**c**) vs. Pilo + NMP 200,  $p < 0.001$ . **b** Hippocampus (**a**) vs. control,  $p < 0.01$ ; (**b**) vs. Pilo + NMP 100,  $p < 0.01$ ; (**c**) vs. Pilo + NMP 200,  $p < 0.01$ . **c** Striatum (**a**) vs. control,  $p < 0.01$ ; (**b**) vs. Pilo + NMP 100,  $p < 0.05$ ; (**c**) vs. Pilo + NMP 200,  $p < 0.05$  (one-way ANOVA and Tukey multiple comparisons test).



► **Fig. 7** Effects of the methanol fraction, rich in NMP, from *S. obtusifolium* on GSH content ( $\mu\text{g/g}$  tissue) in mice subjected to Pilo-induced convulsions. Groups tested (12–14 animals): control (normal), Pilo only, and Pilo + NMP treated. **a** PFC (**a**) vs. control,  $p < 0.01$ ; (**b**) vs. Pilo + NMP 100,  $p < 0.05$ ; (**c**) vs. Pilo + NMP 200,  $p < 0.01$ . **b** Hippocampus (**a**) vs. control,  $p < 0.01$ ; (**b**) vs. Pilo + NMP 100,  $p < 0.001$ ; (**c**) vs. Pilo + NMP 200,  $p < 0.001$ . **c** Striatum (**a**) vs. control,  $p < 0.05$ ; (**b**) vs. Pilo + NMP 100,  $p < 0.01$ ; (**c**) vs. Pilo + NMP 200,  $p < 0.01$  (one-way ANOVA and Tukey multiple comparisons test).

the dopaminergic system, such as apomorphine and amphetamines, as well as antiparkinsonian drugs, such as pergolide and bromocriptine, present antiepileptic and anticonvulsant effects [20, 21]. Dopaminergic neurons also seem to modulate synaptic plasticity, a phenomenon affected by seizure activity [22]. Corroborating with these findings, we showed that the striatum from brains of animals subjected to Pilo-induced convulsions presented decreased DA and DOPAC contents, effects which were partially reversed in the NMP-treated groups.

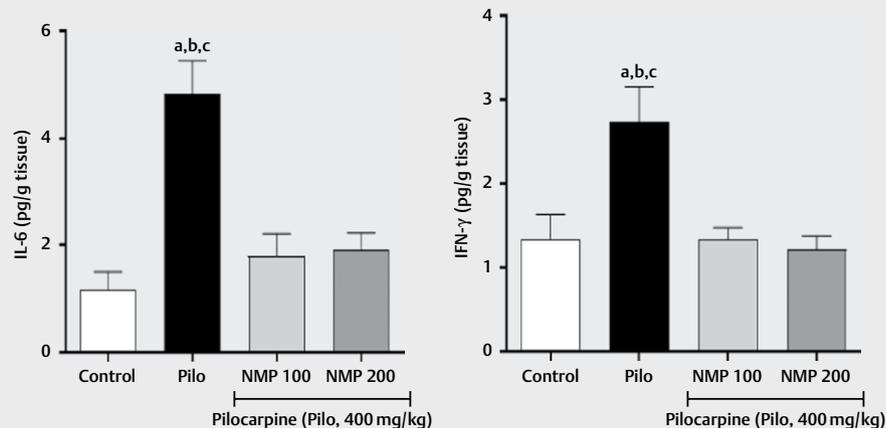
Besides the neuromodulatory role of the dopaminergic system, an intense change in neuronal activity in excitatory (glutamatergic) and inhibitory (GABAergic) neurotransmissions is also known to occur in epilepsy [16]. Earlier, GLU and GABA were shown to be centrally involved in the kindling process that constitutes a model of complex partial seizures [23]. In the present work, the changes in the contents of brain GABA and GLU, observed in mice after the pilocarpine-induced convulsions, were reversed by NMP treatments.

The oxidative stress and excessive ROS production are involved with the initiation and progression of epileptic seizures [24]. Oxidative stress in the hippocampus was previously observed [25] after Pilo-induced seizures in rats, as evidenced by changes in GSH, nitrite content, and lipid peroxidation. Later, a direct relationship between lipid peroxidation and nitrite content was seen in some brain areas after acute seizure activity [26]. Now, we showed that the Pilo-induced seizures in mice cause higher increases in nitrite and lipid peroxidation and also decreases GSH contents in the mice brain, which are indexes of oxidative stress. The treatment of such Pilo groups with NMP prevented all these changes.

The tripeptide GSH, among other physiological functions, is highly involved in brain protection against ROS [27]. Additionally, a compromised brain GSH system has been connected to the oxidative stress, occurring in neurological diseases, including epilepsy [28]. Furthermore, a widespread impairment of the GSH system in epileptic patients was also demonstrated [29]. We showed that Pilo-induced seizures significantly decreased GSH content and these values were increased and brought towards normality in all brain areas tested after the NMP treatments.

A large body of evidence indicates a role for inflammatory mediators in epileptogenesis [12, 30] and epileptic seizures. In addition, the increase in inflammatory mediators was shown to result in secondary brain damage and the likelihood of recurrent seizures [10]. We observed significant increases in brain IL-6 and IFN- $\gamma$  levels in the presence of Pilo-induced seizures in the mouse hippocampus. These changes were completely brought back to control levels after NMP treatments. Post-ictal serum increases in cytokine levels, including IL-6 and IFN- $\gamma$ , were demonstrated in epileptic patients [31], corroborating with our findings.

Previous studies [32] suggested that alterations in gene expression are involved in the genesis of epilepsy and also in the induction of GFAP expression in astrocytes. Earlier evidence [33] reported intense neuronal activity in the hippocampus, leading to a rapid and drastic increase in GFAP expression. Thus, astrogliosis is considered an important feature in epilepsy and may also be involved in the development and persistence of seizures that result in prominent hypertrophy of astrocytes [34]. Although reactive astrocytes are commonly found in epileptic foci, the knowledge of whether



► **Fig. 8** Effects of the methanol fraction, rich in NMP, from *S. obtusifolium* on IL-6 and IFN- $\gamma$  contents in mice subjected to Pilo-induced convulsions. Groups tested (7 to 8 animals): control (normal), Pilo only, and Pilo + NMP treated. IL-6 (a) vs. control,  $p < 0.001$ ; (b) vs. Pilo + NMP 100,  $p < 0.01$ ; (c) vs. Pilo + NMP 200,  $p < 0.01$ . IFN- $\gamma$  (a) vs. control,  $p < 0.01$ ; (b) vs. Pilo + NMP 100,  $p < 0.01$ ; (c) vs. Pilo + NMP 200,  $p < 0.01$ . (one-way ANOVA and Tukey multiple comparisons test).

astrogliosis is a cause or a consequence of epileptogenesis is still an unsolved question [35].

Even though some studies [15, 36, 48] have shown that astrocyte activation needs some time (hours to days) to occur, we showed that Pilo-treated mice present acute and significant increases in GFAP immunostainings in the hippocampus and TC shortly after seizures. These changes were highly attenuated by treatment with NMP. However, others [37] also detected acute astrocyte activation in the brain by MRI, providing insight into acute and reversible brain injury processes in neurologic patients. It is important to point out that these data corroborate our findings. Furthermore, dysfunctional astrocytes are crucial players in epilepsy and should be considered potential and promising targets in therapies for this neurologic disorder [38].

Altogether, the data of the present work strongly suggest a neuroprotective effect for this L-proline derivative (NMP), highly present in the methanol fraction from *S. obtusifolium*, in the model of Pilo-induced seizures in mice. Part of this is probably a consequence of the anti-inflammatory property of NMP, as already shown by us [7]. In an earlier study [39], L-proline was shown to suppress clonic-tonic and focal clonic seizures in ouabain-induced seizures in rats.

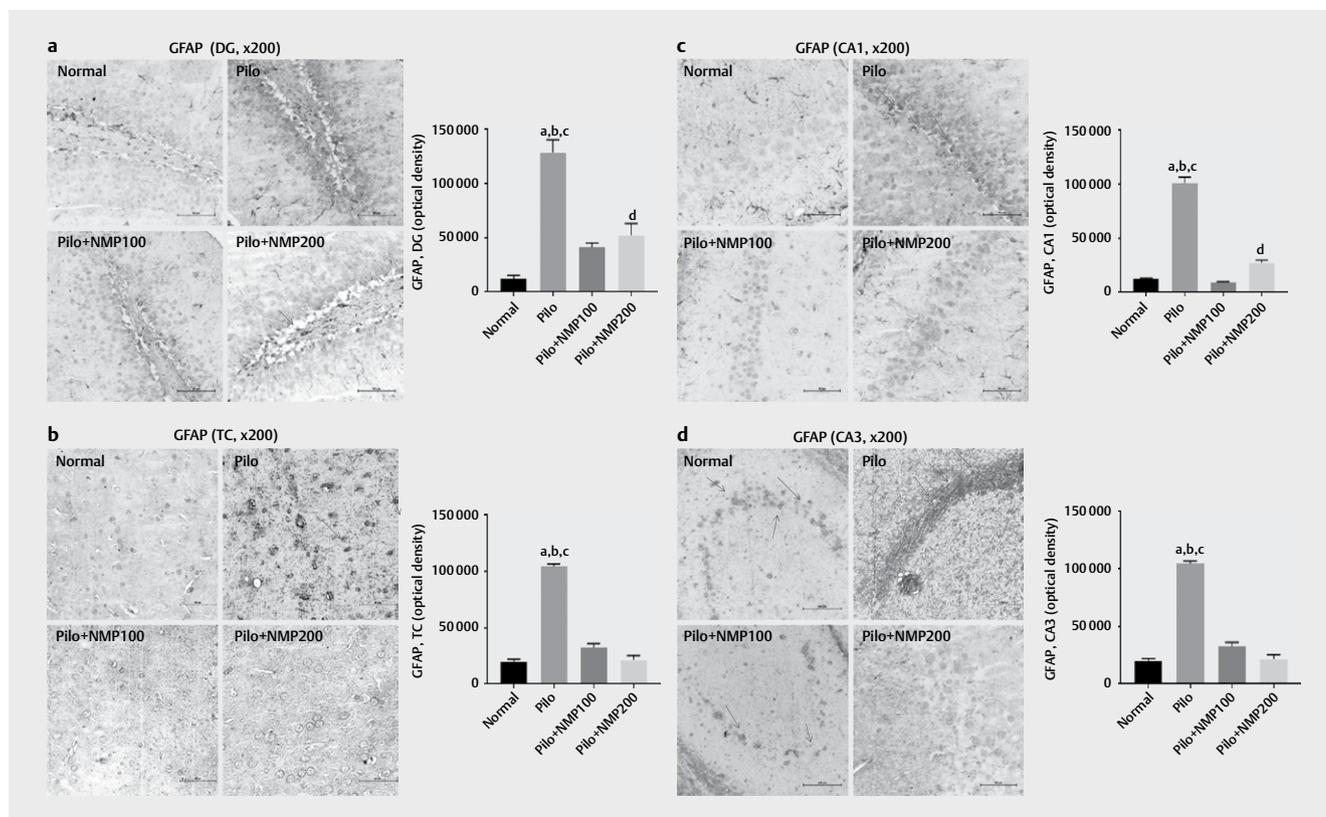
Furthermore, considering the important role of GABA in the mechanism and treatment of epilepsy, we found that NMP may also act through the GABAergic pathway, since it significantly increased both latencies to the first convulsion and death in the PTZ model. In addition, sodium VPA, used as a reference, when combined with a lower NMP dose (25 mg/kg) potentiated the NMP effect on the latency to death. Interestingly, similar findings were also observed with the standard compounds L-proline and *trans*-4-hydroxy-L-proline and also, as expected, with NMP itself (data are not shown). Most bioactive small molecules, such as NMP, are known to interact with proteins or other macromolecule targets, which results in

their final biological effects. Recently, we reported that one of the possible targets for the NMP molecule is the GABA transporter (GAT-1) [40]. These results point again to GABA neurotransmission involvement in the NMP action mechanism.

Furthermore, at least one-third of epileptic patients do not present any benefit from conventional pharmacological treatment [41, 42]. Recently, the amino acid D-leucine was shown to potentially protect mice, when administered before the onset of the kainic acid-induced seizures [43]. The authors found that the D-leucine, present in trace amounts in the brain, worked even better than L-leucine against both kainic acid and electroshock-induced seizures. Earlier, L-proline and derivatives, such as *trans*-4-hydroxy-L-proline, *cis*-4-hydroxy-D-proline, and 3,4-dehydro-DL-proline, were shown to be weak anticonvulsants when used alone, but were able to potentiate the anticonvulsant effect of vigabatrin similarly to that reported for glycine [44].

The intracellular L-proline accumulation in mammalian cells has been demonstrated to correlate with decreased ROS levels and increased protection against oxidative stress [45]. L-proline was also shown to improve the survival rate of vitrified mouse oocytes for protecting mitochondrial functions [46]. The coadministration of L-proline and LPS pointed out the ability of L-proline in preventing the harmful effects of LPS [47]. These authors indicated that LPS induces inflammation and oxidative stress in the rat cerebral cortex and cerebellum, and the coadministration with L-proline prevents these LPS effects.

In conclusion, we showed, for the first time, the neuroprotective effects of the proline derivative NMP isolated from *S. obtusifolium*. These NMP neuroprotective properties are surely associated with its anti-inflammatory and antioxidant effects. However, considering that NMP also presented anticonvulsive activity in the PTZ model, GABAergic neurotransmission is probably an NMP target also involved with the drug anticonvulsive action.



► **Fig. 9** Effects of the methanol fraction, rich in NMP from *S. obtusifolium*, on brain GFAP immunohistochemistry in mice subjected to Pilo-induced convulsions (3 animals per group). Brain areas tested: TC, hippocampal CA3 and CA1 subfields, DG. Groups tested: control (normal), Pilo only, and Pilo+NMP. **a DG:** (a) vs. control,  $p < 0.001$ ; (b) vs. Pilo+NMP100,  $p < 0.001$ ; (c) vs. Pilo+NMP200,  $p < 0.01$ ; (d) vs. control,  $p < 0.05$ . **b TC:** (a) vs. control,  $p < 0.0001$ ; (b) vs. Pilo+NMP100,  $p < 0.0001$ ; (c) vs. Pilo+NMP200,  $p < 0.0001$ . **c CA1:** (a) vs. control,  $p < 0.001$ ; (b) vs. Pilo+NMP100,  $p < 0.001$ ; (c) vs. Pilo+NMP200,  $p < 0.001$ ; (d) vs. control,  $p < 0.05$ . **d CA3:** (a) vs. control,  $p < 0.001$ ; (b) vs. Pilo+NMP100,  $p < 0.001$ ; (c) vs. Pilo+NMP200,  $p < 0.001$ . (one-way ANOVA and Tukey multiple comparison test).

## Materials and Methods

### Plant material and preparation of the methanol fraction containing the bioactive constituent N-methyl-(2S,4R)-trans-4-hydroxy-L-proline

*S. obtusifolium* leaves were collected and handled as previously described [8] and identified (voucher specimen #10,648) by Maria Arlene Pessoa da Silva, Ph.D., botanist at the Herbarium “Dárdano de Andrade Lima”, Regional University of Cariri (URCA), Crato, Brazil. The methanol fraction from the leaves decoction used in the present work contains the bioactive compound NMP and was obtained according to a procedure previously described [7]. The amount of N-methyl-(2S, 4R)-trans-4-hydroxy-L-proline present in the methanol fraction (NMP) tested was also studied by qHNMR, as shown in **Supporting Information**. In addition, spectra for both the methanol fraction and the standard compound (NMP) were also carried out (see **Supporting Information**).

### Animals

Male Swiss mice (25–33 g) were maintained at  $25 \pm 2^\circ\text{C}$ , under a 12/12-h light/dark cycle. Food and water were provided *ad libitum*. The study was submitted and approved by the Ethics Committee on Animal Research of the Faculty of Medicine of the Federal Uni-

versity of Ceará (CEUA/UFC), under the number 59/17, on February 26, 2018.

### Drugs and reagents

Pilo and PTZ were purchased from Sigma-Aldrich and showed a degree of purity of around 99%. Depakene (liquid containing 50 mg/mL sodium valproate) was from Abbott Laboratórios do Brasil Ltda. All other drugs and reagents were of analytical grade.

### Pilocarpine-induced seizures and behavioral assessment

Mice (6–20 animals per group) were pretreated by gavage with NMP (50, 100, and 200 mg/kg, *p.o.*, dissolved in distilled water, 0.1 mL/100 g) or VPA (10 mg/kg, used as the reference or positive control). After 1 h, seizures were induced in all groups by the systemic administration of Pilo (400 mg/kg, *i.p.*). The Pilo-only group was the control group. Then, the animals were placed in individual cages and observed for the following behavioral parameters: latency to the first seizure (elapsed time before the first seizure) and latency to death (time elapsed until death). The seizure was characterized primarily by the hind limb extension and/or uncoordinated jump. Immediately after death, the animals were decapitated and

the brain areas (PFC, hippocampus, and striatum) were dissected for biochemical tests.

### Pentylentetrazole-induced seizures

Although the present study focused on the Pilo-induced seizure model, we also performed the model of PTZ-induced seizures, since GABA neurotransmission may be a possible target for NMP. PTZ is considered to have an antagonistic action on GABA<sub>A</sub> receptors. For that, mice (6–22 animals per group) were pretreated by gavage with NMP (50, 100, and 200 mg/kg, p.o., dissolved in distilled water) or VPA (10 mg/kg) as a reference or positive control. After 1 h, seizures were induced in all groups by the systemic administration of PTZ (80 mg/kg, i.p., PTZ-only group). The following behavioral parameters observed were: latency to the first seizure (elapsed time before the first seizure) and latency to death (time elapsed until death). The seizure was characterized primarily by the hind limb extension and/or uncoordinated jump. The possible potentiation of the NMP effect by VPA was also determined.

### Neurochemical determinations of dopamine and 3,4-dihydroxyphenylacetic acid by HPLC

The contents of DA and its metabolite DOPAC were determined by HPLC (electrochemical detector, model L-ECD-6A; Shimadzu) in the striata from 3 to 10 animals per group according to previously described methods [14]. Briefly, homogenates were prepared in 10% HClO<sub>4</sub> and centrifuged at 4 °C (25000 × g, 15 min). The supernatants were filtered and 20 μL were injected into the HPLC column (flow of 0.6 mL/min). Monoamines were quantified by comparison with standards and the results are expressed as ng/g tissue.

### Brain gamma-aminobutyric acid and glutamate determinations by HPLC

Brain areas (PFC, hippocampus, and striatum) from 4–6 animals were used for the determination of amino acid concentrations. This assay was carried out by reversed-phase HPLC involving pre-column derivatization with orthophthalaldehyde according to a method previously described [48]. The results are expressed as μg/g tissue.

### Determination of brain nitrite contents

Griess reagent was added to a 96-well plate containing supernatants of brain homogenates (PFC, hippocampus, and striatum areas) from 5–7 animals. The absorbance was measured using a microplate reader at 560 nm. In addition, the same brain areas of the animals treated only with vehicle were dissected in the absence of the seizure inducer drug (basal level group). Previously, a standard curve for nitrite was generated using concentrations of 100, 50, 25, 12.5, 6.25, 3.12, and 1.56 nmol/mL [49]. The results are expressed as nmol/g tissue.

### Determination of brain lipid peroxidation by thiobarbituric acid reactive substances assay

Lipid peroxidation expresses the oxidative stress induced by ROS reactivity. This method is used for the determination of MDA in biological samples. The brain areas (PFC, hippocampus, and striatum) from 5 to 7 animals were used for the preparation of 10% homogenates in 1.15% KCl. Then, 250 μL were added to 1 mL 10% TCA, followed by addition of 1 mL 0.6% thiobarbituric acid. After agitation,

this mixture was maintained in a water bath (95–100 °C, 15 min), cooled on ice, and centrifuged (1500 × g/5 min). The TBARS content was determined on a plate reader at 540 nm, with results expressed in nmol MDA/g tissue. A standard curve with MDA was also performed [50].

### Determination of the brain concentration of reduced glutathione

The determination of the GSH concentration was performed in the brains of 12 animals. The assay is based on the reaction of Ellman reagent (DTNB) with the free thiol, giving a disulfide plus 2-nitro-5-thiobenzoic acid mixture. The sample preparation of brain areas (PFC, hippocampus, and striatum) was carried out as follows: 10% homogenates in phosphate buffer were added to an Eppendorf containing 50 μL distilled water and 10 μL trichloroacetic acid (50%). After centrifugation (1000 × g for 15 min at 4 °C), the supernatants (60 μL) were added to the cooled ELISA microplates. Immediately before the readings at 412 nm, 102 μL of the mixture of Tris HCL buffer, and 0.65 mL of 0.01 M DTNB in methanol were added to each well. The concentration of reduced GSH is expressed as μg/g tissue based on a standard curve of GSH.

### Immunoassays for hippocampal interferon-gamma and interleukin 6

The brain area (hippocampus) from 7 animals per group was homogenized in 8 volumes of PBS buffer containing a protease (EMD Biosciences) inhibitor and centrifuged (9000 × g, 5 min). The concentration of the cytokines in 100 μL samples was determined by an immune-enzymatic assay (ELISA; R&D Systems) according to the manufacturer's protocol. The results are expressed in pg/g tissue.

### Immunohistochemical assays for glial fibrillary acidic protein

Hippocampal sections (5 μm, 3 animals per group) were fixed in buffered formalin followed by 70% alcohol. This was followed by deparaffinization, hydration in xylol and ethanol, immersion in 0.1 M citrate buffer (pH 6), and microwave heating (18 min) for antigen recovery. After cooling, the sections were washed with PBS followed by the endogenous peroxidase blockade (15 min) with a 3% H<sub>2</sub>O<sub>2</sub> solution. The sections were incubated overnight (4 °C) with the primary antibodies (anti-GFAP; Sigma-Aldrich) and diluted in PBS according to the manufacturer's instructions. The next day, the sections were washed in PBS, incubated (30 min) with the secondary biotinylated rabbit antibody (anti-IgG) diluted in PBS (1:200 dilution), washed again in PBS, and incubated (30 min) with the conjugated streptavidin-peroxidase complex. After a final washing, the sections were stained with 3,3 diaminobenzidine-peroxide, dehydrated, and mounted in microscope slides for analyses. The data were semiquantified with the Image J software [51].

### Statistical analyses

The results are presented as means ± SEM and were analyzed by one-way ANOVA followed by the Tukey test for multiple comparisons. The immunohistochemical data were analyzed by Image J software. All results were considered significant at p < 0.05.

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## Conflict of Interest

The authors declare that they have no conflict of interest.

## References

- [1] Moshé SL, Perucca E, Ryvlin P, Tomson T. Epilepsy: New advances. *Lancet* 2015; 385: 884–898
- [2] Cifelli P, Grace AA. Pilocarpine-induced temporal lobe epilepsy in the rat is associated with increased dopamine neuron activity. *Int J Neuropsychopharmacol* 2012; 15: 957–964
- [3] Engel J Jr.. Mesial temporal lobe epilepsy: what have we learned? *Neuroscientist* 2001; 7: 340–352
- [4] Thom M, Bertram EH. Temporal lobe epilepsy. *Handb Clin Neurol* 2012; 107: 225–240
- [5] Curia G, Longo D, Biagini G, Jones RSG, Avoli M. The pilocarpine model of temporal lobe epilepsy. *J Neurosci Methods* 2008; 172: 143–157
- [6] Scorza FA, Arida RM, Naffah-Mazzacoratti Mda G, Scerni DA, Calderazzo L, Cavalheiro EA. The pilocarpine model of epilepsy: what have we learned? *An Acad Bras Cienc* 2009; 81: 345–365
- [7] de Aquino PE, Magalhães TR, Nicolau LA, Leal LK, de Aquino NC, Dos Santos SM, Neves KR, Silveira ER, Viana GS. The anti-inflammatory effects of N-methyl-(2S,4R)-trans-4-hydroxy-L-proline from *Syderoxylon obtusifolium* are related to its inhibition of TNF-alpha and inflammatory enzymes. *Phytomedicine* 2016; 24: 14–23
- [8] de Aquino PEA, de Souza TFG, Santos FA, Viana AFSC, Louchard BO, Leal LKAM, Rocha TM, Evangelista JSAM, de Aquino NC, de Alencar NMN, Silveira EDR, Viana GSB. The Wound Healing Property of N-Methyl-(2 S,4 R )-trans-4-Hydroxy-L-Proline from *Sideroxylon obtusifolium* is Related to its Anti-Inflammatory and Antioxidant Actions. *Evid-Based Complementary Altern Med* 2019; 29: 2515690X19865166
- [9] Araujo-Neto V, Bomfim RR, Oliveira VOB, Passos AMPR, Oliveira JPR, Lima CA, Mendes SS, Estevam CS, Thomazzi SM. Therapeutic benefits of *Sideroxylon obtusifolium* (Humb. ex Roem. & Schult.) T.D. Penn., Sapotaceae, in experimental models of pain and inflammation. *Rev Bras Farmacogn* 2010; 20: 933–938
- [10] Shimada T, Takemiya T, Sugiura H, Yamagata K. Role of inflammatory mediators in the pathogenesis of epilepsy. *Mediators Inflamm* 2014; 2014: 1–8
- [11] Geronzi U, Lotti F, Grosso S. Oxidative stress in epilepsy. *Expert Rev Neurother* 2018; 18: 427–434
- [12] Bozzi Y, Borrelli E. The role of dopamine signaling in epileptogenesis. *Front Cell Neurosci* 2013; 7: 157
- [13] Turski L, Cavalheiro EA, Bortolotto ZA, Ikonomidou-Turski C, Kleinrok Z, Turski WA. Dopamine-sensitive anticonvulsant site in the rat striatum. *J Neurosci* 1988; 8: 4027–4037
- [14] Machado-Filho JA, Correia AO, Montenegro AB, Nobre ME, Cerqueira GS, Neves KR, Naffah-Mazzacoratti Mda G, Cavalheiro EA, de Castro Brito GA, de Barros Viana GS. Caffeine neuroprotective effects on 6-OHDA-lesioned rats are mediated by several factors, including pro-inflammatory cytokines and histone deacetylase inhibitions. *Behav Brain Res* 2014; 264: 116–125
- [15] Shapiro LA, Wang L, Ribak CE. Rapid astrocyte and microglial activation following pilocarpine-induced seizures in rats. *Epilepsia* 2008; 49: 33–41
- [16] Rezaei M, Sadeghian A, Roohi N, Shojaei A, Mirnajafi-Zadeh J. Epilepsy and dopaminergic system. *Physiol Pharmacol* 2017; 21: 1–14
- [17] Li G, Bauer S, Nowak M, Norwood B, Tackenberg B, Rosenow F, Knake S, Oertel WH, Hamer HM. Cytokines and epilepsy. *Seizure* 2011; 20: 249–256
- [18] Lasoń W, Chlebicka M, Rejdak K. Research advances in basic mechanisms of seizures and antiepileptic drug action. *Pharmacol Reports* 2013; 65: 787–801
- [19] Vezzani A, Granata T. Brain inflammation in epilepsy: experimental and clinical evidence. *Epilepsia* 2005; 46: 1724–1743
- [20] Bozzi Y, Borrelli E. Dopamine in neurotoxicity and neuroprotection: What do D2 receptors have to do with it? *Trends Neurosci* 2006; 29: 167–174
- [21] Bozzi Y, Dunleavy M, Henshall DC. Cell signaling underlying epileptic behavior. *Front Behav Neurosci* 2011; 5: 45
- [22] Hansen N, Manahan-Vaughan D. Dopamine D1/D5 receptors mediate informational saliency that promotes persistent hippocampal long-term plasticity. *Cereb Cortex* 2014; 24: 845–858
- [23] Bradford HF. Glutamate, GABA, and epilepsy. *Prog Neurobiol* 1995; 47: 477–511
- [24] Shin EJ, Jeong JH, Chung YH, Kim WK, Ko KH, Bach JH, Hong JS, Yoneda Y, Kim HC. Role of oxidative stress in epileptic seizures. *Neurochem Int* 2011; 59: 122–137
- [25] Freitas RM, Vasconcelos SMM, Souza FCF, Viana GSB, Fonteles MMF. Oxidative stress in the hippocampus after pilocarpine-induced status epilepticus in Wistar rats. *FEBS J* 2005; 272: 1307–1312
- [26] Júnior HV, de França Fonteles MM, Mendes de Freitas R. Acute seizure activity promotes lipid peroxidation, increased nitrite levels, and adaptive pathways against oxidative stress in the frontal cortex and striatum. *Oxid Med Cell Longev* 2009; 2: 130–137
- [27] Dringen R. Metabolism and functions of glutathione in the brain. *Prog Neurobiol* 2000; 62: 649–671
- [28] Dringen R, Hirrlinger J. Glutathione pathways in the brain. *Biol Chem* 2003; 384: 505–516
- [29] Mueller SG, Trabesinger AH, Boesiger P, Wieser HG. Brain glutathione levels in patients with epilepsy measured by in vivo (1)H-MRS. *Neurology* 2001; 57: 1422–1427
- [30] Friedman A, Dingleline R. Molecular cascades that mediate the influence of inflammation on epilepsy. *Epilepsia* 2011; 52: 33–39
- [31] Sinha S, Patil SA, Jayalekshmy V, Satishchandra P. Do cytokines have any role in epilepsy? *Epilepsy Res* 2008; 82: 171–176
- [32] Steward O, Torre ER, Tomasulo R, Lothman E. Seizures, and the regulation of astroglial gene expression. *Epilepsy Res Suppl* 1992; 7: 197–209
- [33] Steward O, Torre ER, Tomasulo R, Lothman E. Neuronal activity up-regulates astroglial gene expression. *Proc Natl Acad Sci USA* 1991; 88: 6819–6823
- [34] Khurgel M, Ivy GO. Astrocytes in kindling: relevance to epileptogenesis. *Epilepsy Res* 1996; 26: 163–175
- [35] Robel S, Buckingham SC, Boni JL, Campbell SL, Danbolt NC, Riedemann T, Sutor B, Sontheimer H. Reactive astrogliosis causes the development of spontaneous seizures. *J Neurosci* 2015; 35: 3330–3345
- [36] Shapiro LA, Wang L, Ribak CE. Rapid astrocyte and microglial activation following pilocarpine-induced seizures in rats. *Epilepsia* 2008; 49: 33–41
- [37] Sibson NR, Lowe JP, Blamire AM, Martin MJ, Obrenovitch TP, Anthony DC. Acute astrocyte activation in the brain detected by MRI: new insights into T1 hypointensity. *J Cereb Blood Flow Metab* 2008; 28: 621–632

- [38] Coulter DA, Steinhäuser C. Role of astrocytes in epilepsy. *Cold Spring Harb Perspect Med* 2015; 5: a022434
- [39] Tsukada Y, Inoue N, Donaldson J, Barbeau A. Ouabain-induced seizures in rats: suppressive effects of various amino acids against ouabain-induced seizures in rats. *Can J Physiol Pharmacol* 1973; 51: 885–889
- [40] Aquino PEAA, Bezerra JR, Nascimento TS, Tavares J, Lustosa IR, Chaves-Filho AJM, Mottin M, Gaspar DM, Andrade GM, Neves KRT, Biagini G, Silveira ER, Viana GSB. A proline derivative-enriched fraction from *Sideroxylon obtusifolium* protects the hippocampus from intracerebroventricular pilocarpine-induced injury associated with status epilepticus in mice. *Int J Mol Sci* 2020; 21: 4188
- [41] Mitchell JW, Seri S, Cavanna AE. Pharmacotherapeutic and non-pharmacological options for refractory and difficult-to-treat seizures. *J Cent Nerv Syst Dis* 2012; 4: 105–115
- [42] Schmidt D. Drug treatment of epilepsy: Options and limitations. *Epilepsy Behav* 2009; 15: 56–65
- [43] Hartman AL, Santos P, O’Riordan KJ, Stafstrom CE, Marie Hardwick J. Potent anti-seizure effects of D-leucine. *Neurobiol Dis* 2015; 82: 46–53
- [44] Sarhan S, Seiler N. Proline and proline derivatives as anticonvulsants. *Gen Pharmacol Vasc Syst* 1989; 20: 53–60
- [45] Zhang L, Becker DF. Connecting proline metabolism and signaling pathways in plant senescence. *Front Plant Sci* 2015; 6: 552
- [46] Zhang L, Xue X, Yan J, Yan LY, Jin XH, Zhu XH, He ZZ, Liu J, Li R, Qiao J. L-proline: a highly effective cryoprotectant for mouse oocyte vitrification. *Sci Rep* 2016; 6: 26326
- [47] Andrade VS, Rojas DB, de Andrade RB, Kim TDH, Vizuete AF, Zanatta Â, Wagner M, Gonçalves CS, Wannmacher CMD. A possible anti-inflammatory effect of proline in the brain cortex and cerebellum of rats. *Mol Neurobiol* 2017; 55: 4068–4077
- [48] Arisi GM, Ruch M, Foresti ML, Mukherjee S, Ribak CE, Shapiro LA. Astrocyte alterations in the hippocampus following pilocarpine-induced seizures in aged rats. *Aging Dis* 2011; 2: 294–300
- [49] Green LC, Wagner DA, Glogowski J, Skipper PL, Wishnok JS, Tannenbaum SR. Analysis of nitrate, nitrite, and [<sup>15</sup>N] nitrate in biological fluids. *Anal Biochem* 1982; 126: 131–138
- [50] Draper HH, Hadley M. Malondialdehyde determination as index of lipid peroxidation. *Methods Enzymol* 1990; 186: 421–431
- [51] Hsu SM, Raine L, Fanger H. Use of avidin-biotin-peroxidase complex (ABC) in immunoperoxidase techniques: A comparison between ABC and unlabeled antibody (PAP) procedures. *J Histochem Cytochem* 1981; 29: 577–580R