

# Scalable Synthesis of L-*allo*-Enduracididine: The Unusual Amino Acid Present in Teixobactin

Namdeo Gangathade<sup>a,b</sup>

Kiranmai Nayani<sup>a</sup>

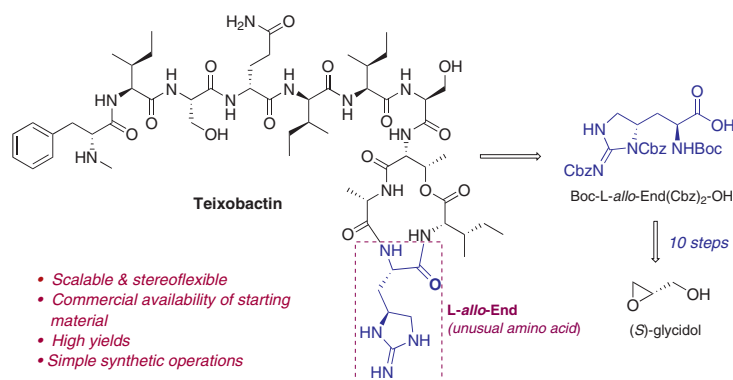
Hemalatha Bukya<sup>a,b</sup>

Prathama S. Mainkar<sup>a,b</sup>

Srivari Chandrasekhar<sup>\*a,b</sup>

<sup>a</sup> Department of Organic Synthesis and Process Chemistry, CSIR-Indian Institute of Chemical Technology (IICT), Hyderabad 50007, India  
srivari@iict.res.in

<sup>b</sup> Academy of Scientific and Innovative Research (AcSIR), Ghaziabad 201002, India



Received: 18.05.2021

Accepted after revision: 13.06.2021

Published online: 13.06.2021

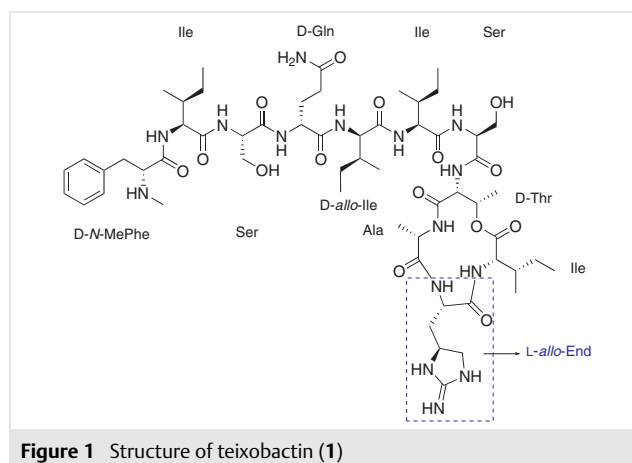
DOI: 10.1055/a-1528-0625; Art ID: st-2021-v0191-I



**Abstract** A scalable synthesis of L-*allo*-enduracididine is achieved from commercially available (S)-glycidol in ten linear steps involving well-established synthetic transformations. The synthetic route is flexible and can be used to synthesize all four diastereomers by changing the stereochemistry of glycidol and Sharpless asymmetric dihydroxylation reagent.

**Key words** antibiotics, teixobactin, depsipeptide, unusual amino acid, L-*allo*-enduracididine, Staudinger reaction, Sharpless asymmetric dihydroxylation

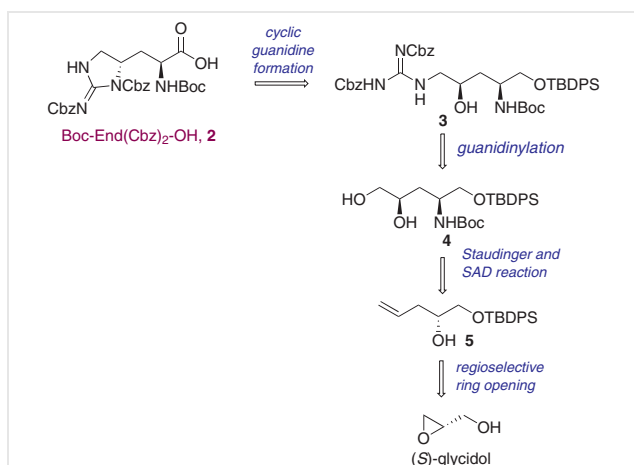
According to WHO, ESKAPE pathogens appear as a major public health concern in hospital acquired infections in critically ill or immunocompromised patients.<sup>[1]</sup> In early 2015, a novel cyclic depsipeptide teixobactin (**1**) was isolated from screening of an unculturable  $\beta$ -proteobacteria (*Eleftheria terrae*) by iChip technique.<sup>[2]</sup> Teixobactin exhibits excellent activities against Gram-positive pathogens, including methicillin-resistant *Staphylococcus aureus* (MRSA, MIC 0.25  $\mu\text{g/mL}$ ), vancomycin-resistant *Enterococcus* species (VRE, MIC 0.5  $\mu\text{g/mL}$ ) and *Mycobacterium tuberculosis* (*Mtb*, MIC 0.125  $\mu\text{g/mL}$ ).<sup>[2]</sup> Teixobactin works as a lipase II inhibitor, like vancomycin, by not allowing pentapeptide incorporation into glycopeptidic cell wall of bacteria, thus rendering it susceptible to rupture.<sup>[3]</sup> In addition, **1** is also found to inhibit lipase III, another important component of bacterial cell-wall synthesis. Teixobactin is an undecapeptide and encompasses an unusual amino acid, L-*allo*-enduracididine<sup>[4]</sup> (L-*allo*-End) and four D-amino acids (Figure 1). The structure of teixobactin contains a depsipeptide macrolide and peptide side chain.



The phenomenal biological activity of **1** prompted research groups to take up the total synthesis<sup>[5–9]</sup> of teixobactin and analogues<sup>[10]</sup> to elucidate its pharmacophore<sup>[11]</sup> towards discovering new antibiotics. So far, five total syntheses of **1** are reported, four solid-phase<sup>[5–7],[9]</sup> and one solution-phase.<sup>[8]</sup> The bottleneck in the synthesis of teixobactin is the availability of the unnatural amino acid, L-*allo*-enduracididine. A careful literature survey revealed easy access to L-*allo*-enduracididine will help in developing faster and affordable steps in synthesis of **1** and analogues on gram scale. The groups which achieved total<sup>[5–9]</sup> and partial<sup>[12]</sup> synthesis have relied on the synthesis of enduracididine either from (2S,3R)-4-hydroxy ornithine (which is obtained from L-aspartic acid)<sup>[13]</sup> developed by Rudolph et al. and Peoples et al. or from protected *trans*-hydroxyproline<sup>[14]</sup> developed by Yuan et al. Recently, Rao and co-workers reported L-*allo*-End precursor on gram scale via intramolecular guanidinylation followed by alcoholysis.<sup>[9]</sup>

Our own efforts to complete the total synthesis of teixobactin are hinged on the commercial nonavailability of enduracididine. We have already achieved teixobactin peptide side-chain synthesis in solution phase as well as in solid phase.<sup>[15]</sup> Thus, we desired to develop an alternate synthesis of *L-allo*-enduracididine which will be scalable and stereoflexible. Herein, we report the synthesis of this unusual amino acid from (*S*)-glycidol which is commercially available.

Accordingly, the retrosynthetic analysis envisioned the construction of suitably protected *L-allo*-enduracididine (Boc-End(Cbz)<sub>2</sub>-OH, **2**) through an intramolecular nucleophilic substitution of guanidine compound **3**, which in turn could be achieved from diol **4** through guanidinylation. The diol **4** could be obtained from homoallylic alcohol **5** by Staudinger reaction followed by Sharpless asymmetric dihydroxylation (SAD). The homoallylic alcohol **5** could be synthesized by regioselective ring opening of (*S*)-glycidol (Scheme 1).



**Scheme 1** Retrosynthesis of *L-allo*-enduracididine (Boc-End(Cbz)<sub>2</sub>-OH, **2**)

Based on the retrosynthetic analysis, (*S*)-glycidol was converted into **2** (Scheme 2). The primary hydroxyl group of commercially available (*S*)-glycidol was protected as *tert*-butyldiphenylsilyl ether (in 95% yield)<sup>[16]</sup> and regioselective ring opening of epoxide was carried out using a reported procedure which gave homoallylic alcohol **5** in 100 g scale.<sup>[16,17]</sup> The regioselective opening of epoxide was achieved with CuI catalyst and vinylmagnesium bromide to get alcohol **5** in 96% yield. Mesylation of alcohol **5** followed by azide displacement using NaN<sub>3</sub> gave azido pentenol **6** with inversion of configuration at C-2 and 90% yield over two steps. The azide **6** was reduced under Staudinger reaction conditions using TPP in THF–H<sub>2</sub>O (3:1) in the presence of (Boc)<sub>2</sub>O to provide *N*-Boc-protected amine **7** in 92% yield. The second chirality was introduced via Sharpless asymmetric dihydroxylation<sup>[18]</sup> using AD mix-β and methanesulfonamide in *t*-BuOH–H<sub>2</sub>O (1:1) at 0 °C for 20 h to realize the diol **4** in 92% yield as a separable diastereomeric mixture

(by silica gel column chromatography) in 7:3 ratio with the required diastereomer being the major isomer. Our plan was to convert this diol into amino alcohol to couple with *N,N'*-Di-Cbz-1*H*-pyrazole-1-carboxamide to introduce guanidine moiety. Initially, the diol **4** was monotosylated in situ with Ts<sub>2</sub>O/2,4,6-collidine/pyridine in CH<sub>2</sub>Cl<sub>2</sub> at ≤ –10 °C, treated with ammonium hydroxide in EtOH at 60 °C to give amino alcohol via epoxide<sup>[19]</sup> which on further treatment with *N,N'*-di-Cbz-1*H*-pyrazole-1-carboxamide<sup>[5,12]</sup> gave guanidine derivative **3** in 52% overall yield for four sequential transformations without purification of intermediates. To improve the yield of guanidine derivative **3** further, we thought of an alternative synthetic sequence. Selective mesylation of primary alcohol in compound **4** with MsCl/Et<sub>3</sub>N in CH<sub>2</sub>Cl<sub>2</sub> at ≤ –30 °C, followed by treatment with NaN<sub>3</sub> in DMF at 70 °C gave azido alcohol **8** in 87% yield. Then, the azide **8** was reduced under Staudinger reaction conditions (TPP, THF–H<sub>2</sub>O) to provide amino alcohol which on further treatment with *N,N'*-di-Cbz-1*H*-pyrazole-1-carboxamide<sup>[5,12]</sup> gave the guanidine derivative **3** in 85% yield (Scheme 2).<sup>[20]</sup>

The intramolecular cyclization of **3** via triflate<sup>[5,12]</sup> using triflic anhydride and *N,N*-diisopropylethylamine at –78 °C allowed us to construct the enduracididine skeleton **9** in 90% yield.<sup>[21]</sup> This upon deprotection of silyl group with TBAF in THF afforded alcohol **10** in 95% yield. Finally, the oxidation of the obtained primary alcohol **10** using DMP gave aldehyde which upon Pinnick–Lindgren oxidation using a combination of sodium chlorite and NaH<sub>2</sub>PO<sub>4</sub> in *t*-BuOH–H<sub>2</sub>O provided the target building block, *L-allo*-enduracididine (Boc-End(Cbz)<sub>2</sub>-OH, **2**) in 74% yield over two steps, which is being used to complete the total synthesis of teixobactin. A small portion of the carboxylic acid **2** was converted into the corresponding methyl ester **11** using K<sub>2</sub>CO<sub>3</sub>/MeI in 76% yield. The present approach allows the synthesis of *L-allo*-enduracididine in gram scale due to commercial availability of starting material and simple synthetic operations.

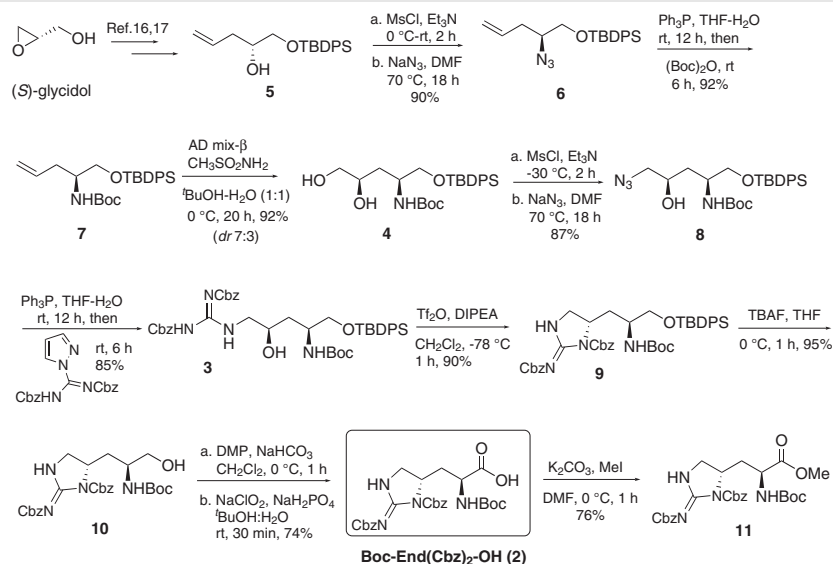
In conclusion, a stereoflexible and scalable synthesis of Boc-End(Cbz)<sub>2</sub>-OH, an unusual amino acid building block of potent depsipeptide antibiotic teixobactin, has been achieved in ten steps with an overall yield of 22.75%. By changing the stereochemistry of starting material, viz., glycidol and dihydroxylating agent, other diastereomers can be synthesized with equal ease.

## Conflict of Interest

The authors declare no conflict of interest.

## Funding Information

N. G. thanks the Council of Scientific and Industrial Research (CSIR) for the research fellowship. K. N. thanks the Council of Scientific and

Scheme 2 Synthesis of L-*allo*-enduracididine 2

Industrial Research, Indian Institute of Chemical Technology (CSIR-IICT) for fellowship and research facilities under the National Laboratories Scheme of the Council of Scientific and Industrial Research (CSIR, 11/3/Rectt.-2020). H. B. thanks the Indian Council of Medical Research (ICMR), Government of India for research fellowship. P. S. M. thanks the Indian Council of Medical Research (ICMR, AMR/IN/111/2017-ECD-II) for research grant. S. C. thanks the Science and Engineering Research Board (SERB, SB/S2/JCB-002/2015), Government of India for J C Bose fellowship.

## Acknowledgment

The authors thank the Council of Scientific and Industrial Research (CSIR), Ministry of Science and Technology, Government of India for research facilities.

## Supporting Information

Supporting information for this article is available online at <https://doi.org/10.1055/a-1528-0625>.

## References and Notes

- (1) (a) Willyard, C. *Nature* **2017**, *543*, 7643. (b) Arias, C. A.; Murray, B. E. *N. Engl. J. Med.* **2015**, *372*, 1168. (c) Zhen, X.; Lundborg, C. S.; Sun, X.; Hu, X.; Dong, H. *Antimicrob. Resist. Infect. Control* **2019**, *8*, 137. (d) Mulani, M. S.; Kamble, E. E.; Kumkar, S. N.; Tawre, M. S.; Pardesi, K. R. *Front. Microbiol.* **2019**, *10*, 539.
- (2) Ling, L. L.; Schneider, T.; Peoples, A. J.; Spoering, A. L.; Engels, I.; Conlon, B. P.; Mueller, A.; Schaeberle, T. F.; Hughes, D. E.; Epstein, S.; Jones, M.; Lazarides, L.; Steadman, V. A.; Cohen, D. R.; Felix, C. R.; Fetterman, K. A.; Millett, W. P.; Nitti, A. G.; Zullo, A. M.; Chen, C.; Lewis, K. *Nature* **2015**, *517*, 455.
- (3) Ng, V.; Chan, W. C. *Chem. Eur. J.* **2016**, *22*, 12606.
- (4) Atkinson, D. J.; Naysmith, B. J.; Furkert, D. P.; Brimble, M. A. *Beilstein J. Org. Chem.* **2016**, *12*, 2325.
- (5) Giltrap, A. M.; Dowman, L. J.; Nagalingam, G.; Ochoa, J. L.; Lington, R. G.; Britton, W. J.; Payne, R. J. *Org. Lett.* **2016**, *18*, 2788.
- (6) Jin, K.; Sam, L. H.; Po, K. H. L.; Lin, D.; Zadeh, E. H. G.; Chen, S.; Yuan, Y.; Li, X. *Nat. Commun.* **2016**, *7*, 12394.
- (7) Liu, L.; Wu, S.; Wang, Q.; Zhang, M.; Wang, B.; He, G.; Chen, G. *Org. Chem. Front.* **2018**, *5*, 1431.
- (8) Gao, B.; Chen, S.; Hou, Y. N.; Zhao, Y. J.; Ye, T.; Xu, Z. *Org. Biomol. Chem.* **2019**, *17*, 1141.
- (9) Zong, Y.; Fang, F.; Meyer, K. J.; Wang, L.; Ni, Z.; Gao, H.; Lewis, K.; Zhang, J.; Rao, Y. *Nat. Commun.* **2019**, *10*, 3268.
- (10) (a) Jad, Y. E.; Acosta, G. A.; Naicker, T.; Ramtahal, M.; El-Faham, A.; Govender, T.; De la Torre, B. G.; Albericio, F. *Org. Lett.* **2015**, *17*, 6182. (b) Parmar, A.; Iyer, A.; Vincent, C. S.; Van Lysebetten, D.; Prior, S. H.; Madder, A.; Taylor, E. J.; Singh, I. *Chem. Commun.* **2016**, *52*, 6060. (c) Yang, H.; Chen, K. H.; Nowick, J. S. *ACS Chem. Biol.* **2016**, *11*, 1823. (d) Monaim, S. A. H. A.; Jad, Y. E.; Acosta, G. A.; Naicker, T.; Ramchuran, E. J.; El-Faham, A.; Govender, T.; Kruger, H. G.; De la Torre, B. G.; Albericio, F. *RSC Adv.* **2016**, *6*, 73827. (e) Monaim, S. A. H. A.; Jad, Y. E.; Ramchuran, E. J.; El-Faham, A.; Govender, T.; Kruger, H. G.; De la Torre, B. G.; Albericio, F. *ACS Omega* **2016**, *1*, 1262. (f) Monaim, S. A. H. A.; Noki, S.; Ramchuran, E. J.; El-Faham, A.; Albericio, F.; De la Torre, B. G. *Molecules* **2017**, *22*, 1632. (g) Monaim, S. A. H. A.; Ramchuran, E. J.; El-Faham, A.; Albericio, F.; De la Torre, B. G. *J. Med. Chem.* **2017**, *60*, 7476. (h) Wu, C.; Pan, Z.; Yao, G.; Wang, W.; Fang, L.; Su, W. *RSC Adv.* **2017**, *7*, 1923. (i) Jin, K.; Po, K. H. L.; Wang, S.; Reuven, J. A.; Wai, C. N.; Lau, H. T.; Chan, T. H.; Chen, S.; Li, X. *Bioorg. Med. Chem.* **2017**, *25*, 4990. (j) Parmar, A.; Iyer, A.; Lloyd, D. G.; Vincent, C. S.; Prior, S. H.; Madder, A.; Taylor, E. J.; Singh, I. *Chem. Commun.* **2017**, *53*, 7788. (k) Parmar, A.; Iyer, A.; Prior, S. H.; Lloyd, D. G.; Goh, E. T. L.; Vincent, C. S.; Pallag, T. P.; Bachrati, C. Z.; Breukink, E.; Madder, A.; Lakshminarayanan, R.; Taylor, E. J.; Singh, I. *Chem. Sci.* **2017**, *8*, 8183. (l) Schumacher, C. E.; Harris, P. W. R.; Ding, X.-B.; Krause, B.; Wright, T. H.; Cook,

- G. M.; Furkert, D. P.; Brimble, M. A. *Org. Biomol. Chem.* **2017**, *15*, 8755. (m) Ng, V.; Kuehne, S. A.; Chan, W. C. *Chem. Eur. J.* **2018**, *24*, 9136. (n) Parmar, A.; Lakshminarayanan, R.; Iyer, A.; Mayandi, V.; Goh, E. T. L.; Lloyd, D. G.; Chalasani, M. L. S.; Verma, N. K.; Prior, S. H.; Beuerman, R. W.; Maddar, A.; Taylor, E. J.; Singh, I. *J. Med. Chem.* **2018**, *61*, 2009.
- (11) (a) Parmar, A.; Prior, S. H.; Iyer, A.; Vincent, C. S.; Van Lysebetten, D.; Breukink, E.; Maddar, A.; Taylor, E. J.; Singh, I. *Chem. Commun.* **2017**, 53, 2016. (b) Yang, H.; Du Bois, D. R.; Ziller, J. W.; Nowick, J. S. *Chem. Commun.* **2017**, 53, 2772. (c) Chen, K. H.; Le, S. P.; Han, X.; Frias, J. M.; Nowick, J. S. *Chem. Commun.* **2017**, 53, 11357. (d) Yang, H.; Wierzbicki, M.; Du Bois, D. R.; Nowick, J. S. *J. Am. Chem. Soc.* **2018**, *140*, 14028. (e) Monaim, S. A. H. A.; Jad, Y. E.; El-Faham, A.; De la Torre, B. G.; Albericio, F. *Bioorg. Med. Chem.* **2018**, *26*, 2788. (f) Yang, H.; Pishenko, A. V.; Li, X.; Nowick, J. S. *J. Org. Chem.* **2020**, *85*, 1331. (g) Gunjal, V. B.; Thakare, R.; Chopra, S.; Reddy, D. S. *J. Med. Chem.* **2020**, *63*, 12171.
- (12) Dhara, S.; Gunjal, V. B.; Handore, K. L.; Reddy, D. S. *Eur. J. Org. Chem.* **2016**, 4289.
- (13) (a) Rudolph, J.; Hannig, F.; Theis, H.; Wischnat, R. *Org. Lett.* **2001**, *3*, 3153. (b) Peoples, A. J.; Hughes, D.; Ling, L. L.; Millett, W.; Nitti, A.; Spoering, A.; Steadman, V. A.; Chiva, J. Y. C.; Lazarides, L.; Jones, M. K.; Poulence, K. G.; Lewis, K. WO2014089053 **2014**. (c) Peoples, A. J.; Hughes, D.; Ling, L. L.; Millett, W.; Nitti, A.; Spoering, A.; Steadman, V. A.; Chiva, J. Y. C.; Lazarides, L.; Jones, M. K.; Poulence, K. G.; Lewis, K. US20140194345, **2014**.
- (14) Craig, W.; Chen, J.; Richardson, D.; Thorpe, R.; Yuan, Y. *Org. Lett.* **2015**, *17*, 4620.
- (15) (a) Sangeetha, D.; Nayani, K.; Mainkar, P. S.; Chandrasekhar, S. *Synlett* **2019**, *30*, 2268. (b) Bukya, H.; Nayani, K.; Gangireddy, P.; Mainkar, P. S. *Eur. J. Org. Chem.* **2020**, 5358.
- (16) Carneiro, V. M. T.; Avila, C. M.; Balunas, M. J.; Gerwick, W. H.; Pilli, R. A. *J. Org. Chem.* **2014**, *79*, 630.
- (17) Bonini, C.; Chiummiento, L.; Lopardo, M. T.; Pullez, M.; Colobert, F.; Solladie, G. *Tetrahedron Lett.* **2003**, *44*, 2695.
- (18) (a) Sharpless, K. B.; Amberg, W.; Bennani, Y. L.; Crispino, G. A.; Hartung, J.; Jeong, K. S.; Kwong, H. L.; Morikawa, K.; Wang, Z. M.; Xu, D.; Zhang, X.-L. *J. Org. Chem.* **1992**, *57*, 2768. (b) Kolb, H. C.; Sharpless, K. B. *Chem. Rev.* **1994**, *94*, 2483.
- (19) (a) Austad, B. A.; Calkins, T. L.; Fang, F. G.; Horstmann, T. E.; Hu, Y.; Lewis, B. M.; Niu, X.; Noland, T. A.; Orr, J. D.; Schnaderbeck, M. J.; Zhang, H.; Asakawa, N.; Asai, N.; Chiba, H.; Hasebe, T.; Hoshino, Y.; Ishizuka, H.; Kajima, T.; Kayano, A.; Komatsu, Y.; Kubota, M.; Kuroda, H.; Miyazawa, M.; Tagami, K.; Watanabe, T. *Synlett* **2013**, *24*, 333. (b) Sun, D.-Y.; Han, G.-Y.; Gong, J.-X.; Nay, B.; Li, X.-W.; Guo, Y.-W. *Org. Lett.* **2017**, *19*, 714.
- (20) **Synthetic Procedure for Guanidine Derivative 3** Triphenylphosphine (2.0 g, 7.2 mmol) was added to a stirred solution of azide **8** (1.27 g, 2.5 mmol) in THF–H<sub>2</sub>O (15 mL, 3:1) at 0 °C. Then the reaction was allowed to warm to room temperature and stirred for 12 h. After this period, to the reaction Goodman's reagent (*N,N'*-di-Cbz-1*H*-pyrazole-1-carboxamide, 968 mg, 2.5 mmol) was added, and the mixture was stirred for another 6 h. The reaction was extracted with EtOAc (2 × 100 mL), the organic layer was washed with brine (75 mL), dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, and concentrated under vacuum. The residue was purified by column chromatography on silica gel using 85:15 hexanes–EtOAc (v/v) as eluent to give **3** (1.7 g, 85%) as a white solid. TLC: *R*<sub>f</sub> = 0.5 (hexanes–EtOAc, 7:3); mp 92 °C; [α]<sub>D</sub><sup>20</sup> +6.2 (c 1.1, CHCl<sub>3</sub>). IR (neat): ν<sub>max</sub> = 3370, 3334, 2946, 1640, 1057 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ = 11.65 (s, 1 H), 8.71–8.68 (br s, 1 H), 7.57–7.50 (m, 4 H), 7.40–7.16 (m, 16 H), 5.10 (s, 2 H), 5.04 (s, 2 H), 4.82 (d, *J* = 8.8 Hz, 1 H), 4.55 (s, 1 H), 3.86–3.75 (m, 1 H), 3.75–3.59 (m, 3 H), 3.51 (dd, *J* = 10.3, 3.7 Hz, 1 H), 3.23–3.08 (m, 1 H), 1.65–1.53 (m, 1 H), 1.37 (s, 9 H), 1.35–1.25 (m, 1 H), 0.98 (s, 9 H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>): δ = 163.6, 157.3, 156.1, 153.5, 136.7, 135.6, 135.5, 134.6, 132.9, 132.7, 129.9, 129.8, 128.6, 128.5, 128.4, 128.3, 128.1, 127.8, 127.8, 80.1, 68.0, 67.0, 66.3, 48.6, 46.3, 38.1, 28.3, 26.8, 19.2. HRMS (ESI): *m/z* calcd for [M + H]<sup>+</sup>: C<sub>43</sub>H<sub>55</sub>N<sub>4</sub>O<sub>8</sub>Si: 783.3779; found: 783.3783.
- (21) **Synthetic Procedure for Intramolecular Cyclization of Guanidine Derivative 3; (S)-Benzyl-2-[(benzyloxy)carbonylimino]-5-((S)-2-[(tert-butoxycarbonyl)amino]-3-[(tert-butyl diphenylsilyloxy)propyl]imidazolidine-1-carboxylate (9)**  
To a solution of **3** (3.2 g, 4 mmol) in anhydrous CH<sub>2</sub>Cl<sub>2</sub> (20 mL) was added DIPEA (3.6 mL, 20 mmol), followed by Tf<sub>2</sub>O (0.76 mL, 4.5 mmol) dropwise at –78 °C under nitrogen atmosphere. After stirring for 1 h, the reaction was quenched by the addition of ammonium chloride (100 mL), the two layers were separated, and the aqueous layer was extracted with CH<sub>2</sub>Cl<sub>2</sub> (100 mL). The combined organic layer was washed with brine (50 mL), dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, and concentrated under vacuum to give a colorless oil. The residue was purified by column chromatography on silica gel using 70:30 hexanes–EtOAc (v/v) as eluent to give **9** (2.81 g, 90%) as a white solid. TLC: *R*<sub>f</sub> = 0.5 (hexanes–EtOAc, 1:1); mp 84 °C; [α]<sub>D</sub><sup>20</sup> –7.5 (c 1.0, CHCl<sub>3</sub>). IR (neat): ν<sub>max</sub> = 3758, 3709, 3481, 3367, 2940, 1710, 1259, 1159 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ = 8.82–8.41 (br s, 1 H), 7.63–7.52 (m, 4 H), 7.47–7.27 (m, 16 H), 5.27 (s, 2 H), 5.16 (q, *J* = 12.4 Hz, 2 H), 4.63 (d, *J* = 8.9 Hz, 1 H), 4.34 (t, *J* = 8.4 Hz, 1 H), 3.79 (t, *J* = 8.4 Hz, 1 H), 3.72–3.60 (m, 2 H), 3.52 (dd, *J* = 9.9, 3.4 Hz, 1 H), 3.43 (d, *J* = 9.5 Hz, 1 H), 2.07 (t, *J* = 12.0 Hz, 1 H), 1.63 (t, *J* = 11.8 Hz, 1 H), 1.43 (s, 9 H), 1.01 (s, 9 H). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>): δ = 156.0, 135.6, 135.6, 135.3, 133.1, 132.9, 130.1, 130.0, 128.8, 128.4, 128.2, 128.0, 127.9, 79.8, 68.4, 67.5, 66.5, 54.0, 48.7, 36.1, 28.5, 26.9, 19.3. HRMS (ESI): *m/z* calcd for [M + H]<sup>+</sup>: C<sub>43</sub>H<sub>53</sub>N<sub>4</sub>O<sub>7</sub>Si: 765.3662; found: 765.3678.