An Update on Laboratory Diagnostics in Haemophilia A and B

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Abstract

Haemophilia A (HA) and B (HB) are X-linked hereditary bleeding disorders caused by lack of activity of coagulation factors VIII (FVIII) or IX (FIX), respectively. Besides conventional products, modern replacement therapies include FVIII or FIX concentrates with an extended half-life (EHL-FVIII/FIX). Two main strategies for measuring plasma FVIII or FIX activity are applied: the one-stage clotting assay (OSCA) and the chromogenic substrate assay (CSA), both calibrated against plasma (FVIII/FIX) standards. Due to the structural modifications of EHL-FVIII/FIX, reagent-dependent assay discrepancies have been described when measuring the activity of these molecules. Assay discrepancies have also been observed in FVIII/FIX gene therapy approaches. On the other hand, nonfactor replacement by the bispecific antibody emicizumab, a FVIIIa-mimicking molecule, artificially shortens activated partial thromboplastin time–based clotting times, making standard OSCAs inapplicable for analysis of samples from patients treated with this drug. In this review, we aim to give an overview on both, the currently applied and future therapies in HA and HB with or without inhibitors and corresponding test systems suitable for accompanying diagnostics.

Zusammenfassung

Bei der Hämophilie A (HA) und B (HB) handelt es sich um X-chromosomal vererbte Blutungsstörungen, die durch einen Mangel an Aktivität der Gerinnungsfaktoren VIII (FVIII) bzw. IX (FIX) verursacht werden. Im Rahmen entsprechender Substitutionstherapien kommen heute auch FVIII- oder FIX-Konzentrate mit einer verlängerten...
**Introduction**

Haemophilia A (HA) and B (HB) are X-linked hereditary bleeding disorders caused by lack of activity of coagulation factors VIII (FVIII) or IX (FIX).\(^1\) To date, treatment of HA and HB is dominated by replacement therapies using plasma-derived or recombinant FVIII/FIX concentrates, including those with an extended half-life (EHL-FVIII/FIX).\(^2,3\) Especially in HA, a serious side effect of factor replacement remains the development of inhibitors. Approximately 30% of treated patients are affected, and require treatment with FVIII bypassing agents, such as recombinant activated FVII (rFVIIa) or plasma-derived activated prothrombin complex concentrate (aPCC).\(^4\) Besides these established bypassing drugs, recombinant porcine FVIII has become available for the treatment of HA inhibitor patients.\(^5\)

In addition to conventional intravenous factor replacement or bypassing schemes, emicizumab, a subcutaneously administered, bispecific monoclonal antibody that mimics the cofactor functions of activated FVIII (FVIIIa), has recently been approved for the treatment of HA patients with or without inhibitors.\(^6,7\) Furthermore, 20 years after the first attempts, gene therapy is becoming a real treatment option in both HA and HB.\(^8\)

For diagnosis and therapy monitoring, two main strategies for measuring plasma FVIII or FIX activity are applied in laboratories worldwide: the activated partial thromboplastin time (aPTT)-based one-stage clotting assay (OSCA) and the chromogenic substrate assay (CSA); both usually calibrated against plasma (FVIII/FIX) standard preparations.\(^9\) Nevertheless, owing to different underlying assay principles, gene mutation-dependent assay discrepancies have been described, requiring the application of both assay types for initial diagnosis and staging of disease severity.\(^10\) Regarding post-infusion monitoring, recombinant FVIII/FIX concentrates also show discrepant results, on the one hand between OSCA and CSA, and, on the other hand, between different types of OSCA.\(^11\) Due to deviant characteristics of expressed molecules, assay discrepancies have also been observed in FVIII/FIX gene therapy. Moreover, emicizumab excessively shortens aPTT-clotting times, making this assay or aPTT-based standard OSC assays inapplicable for the analysis of samples obtained from patients treated with this FVIIIa-mimetic drug.\(^12\)

The aforementioned issues illustrate that monitoring of modern HA/HB treatment options, including EHL concentrates, gene therapy and nonfactor replacement (e.g., emicizumab), require correspondingly adapted and/or qualified assays to ensure accurate and meaningful results. In the present review, we aim to give an overview on the molecules and mechanisms behind these therapies and to describe suitable test systems.

**Basic Principles of Functional Assays in Haemophilia Testing**

**FVIII/FIX Activity Assays**

Two major strategies for measuring plasma FVIII or FIX activity are applied, the aPTT-based OSCA and the CSA.\(^9\) The basic principles of these different types of assays are illustrated in Fig. 1. In the OSCA, the patient sample is diluted either using FVIII- or FIX-deficient plasma, followed by an aPTT measurement. Under such conditions, the aPTT solely depends on the FVIII or FIX activity in the original sample. Although the aPTT is a two-step process (addition of the aPTT reagent and incubation, followed by addition of CaCl\(_2\) as the starting reagent), aPTT-based determination of factor activity is designated as an OSCA. Hereby, formation of the tenase-complex (FXa + FVIIia) after recalcification and subsequent formation of prothrombinase, thrombin and fibrin proceeds without interruption (Fig. 1A). In contrast, the CSA is designed and denoted as a two-stage assay. During the first stage, the patient sample is strongly diluted and a specific reagent added (Fig. 1B: FVIII CSA; Fig. 1C: FIX CSA) assembling the tenase complex. After incubation, during the second stage of the CSA, the amount of generated FXa, which reflects the FVIII or FIX activity in the original sample,
is measured by addition of a corresponding chromogenic peptide substrate. Both the OSCA and the CSA are usually calibrated by the use of plasma standards calibrated against the World Health Organization (WHO) International Standard (IS) for factor VIII (and von Willebrand factor [vWF]) in plasma (available from the National Institute for Biological Standards and Control [NIBSC], code: 07/316) or the WHO IS for blood coagulation factors II, VII, IX, X in plasma (NIBSC code: 09/172) by the manufacturer.\textsuperscript{13,14}

Besides the general differences between OSCA and CSA, there are also significant methodological variances within these classes of assays. Regarding OSCAs, a multitude of differently composed aPTT reagents based on different contact phase activators (including ellagic acid, polyphenols, different types of silica, and kaolin) are available from different manufacturers and are used in laboratories worldwide. Within the class of CSAs, differences between assays mainly comprise the use of either human or bovine factors and differing factor compositions and concentrations.\textsuperscript{15}

Detection of FVIII/FIX Inhibitors ([Nijmegen]-Bethesda Assay)

The detection and quantification of antibodies that inhibit (neutralise) the activity of FVIII or FIX is usually done using the Bethesda protocol.\textsuperscript{16,17} In case of FVIII inhibitor testing, the Nijmegen-modification (Nijmegen-Bethesda Assay [NBA]) is regularly applied.\textsuperscript{18} The basic principle of the NBA is illustrated in Fig. 2. In brief, (heat inactivated) patient sample and, if necessary, dilutions thereof are mixed (1 + 1) with (buffered) pooled plasma ([B]PP). After incubation, the remaining factor activities are determined. In parallel, a control reaction consisting of BPP and factor-decient plasma or buffer is tested. The relative activity (RA) is calculated as shown in Fig. 2A. By definition, an inhibitor titre of one Bethesda unit (BU) per mL inhibits the factor activity by 50%, a titre of 2 BU/mL by 75% (Fig. 2B). To exclude any determinations based on borderline or low (poorly reproducible) factor activities, the range of valid RAs is limited from 75% (defining the detection limit of 0.42 BU/mL) down to 25%. Accordingly, samples with titres of greater than 2 BU/mL need further testing with increasing dilutions (Fig. 2B, C).

Heat inactivation of patient plasma samples (e.g., 56 °C for 30 minutes) is considered as an important pre-analytical step to eliminate remaining factor activities and to liberate anti-FVIII antibodies bound to FVIII or cross-reacting material.\textsuperscript{19,20} In principle, the kind of assay used for the measurement of remaining factor activities (OSC- or CS-assay) is of minor importance. However, the use of CSA formats may
generally lower the potential interference of non-target inhibitory substances such as lupus anticoagulants and may therefore increase assay specificity.\textsuperscript{21,22}

**Commonly Used OSC and CS Assay Reagents**

A recent survey initiated by the section ‘haemophilia’ of the German (D), Austrian (A), and Swiss (CH) Society of Thrombosis and Haemostasis Research (GTH) among haemophilia centres in the DACH countries revealed that the majority of centres primarily apply the OSCA format for both plasma FVIII and FIX activity measurements. Nevertheless, more than 80\% of centres also have a FVIII CSA available, while the application of a FIX CSA is less common. Manufacturers of the used coagulation analysers are Siemens Healthineers ([Siemens], Erlangen, Germany), IL Werfen (Barcelona, Spain), and Stago (Asnières-sur-Seine, France). Accordingly, the most often applied aPTT reagents comprise the following: Actin, Actin FS, Actin FSL, and Pathromtin SL (Siemens); APTT-SP, SynthAfax, and SynthAsil (IL Werfen); STA-PTT Automate (A), (STA-)Cephascreen, and (STA-)C.K. Prest (Stago). The most common FVIII CSA include the following: FVIII Chromogenic Assay (Siemens), Electrachrome FVIII (IL Werfen), TriniCHROM FVIII:C (Stago), COMATIC FACTOR VIII (Chromogenix/IL), BIOPHEN FVIII:C (Hyphen-Biomed, Neuville-sur-Oise, France), and TECHNOCHROM FVIII:C (Technoclone, Vienna, Austria).

Regarding chromogenic determination of FIX activity, only two CE/IVD-marked FIX CS assays are currently available on the European market: the BIOPHEN FIX (Hyphen-Biomed) and the Rox Factor IX assays (Rossix, Mölndal, Sweden). In this article, with respect to potential specific assay discrepancies, we focus on the aforementioned reagents and kits.

**Assay Discrepancies in the Diagnosis of HA and HB**

Discrepancies between the two major test principles (OSCA and CSA) have been observed in patients with mild and
moderate forms of HA. Several gene mutations have been shown to be associated with discrepant results. In a recently published study on nonsevere HA, however, even discrepancies between different OSC assays have been observed, calling for a larger cohort study to systematically address such inter-reagent variations.

Also with respect to HB it has been reported that FIX CSA appear to correlate better with the clinical picture at least in some cases of nonsevere HB. The authors concluded that both assays are required for a correct diagnosis and classification.

Accordingly, the recent World Federation of Haemophilia (WFH) guidelines for the management of haemophilia recommends the use of more than one type of factor assay for the detection and classification of all forms of HA or HB.

Drugs for and Monitoring of Factor Replacement Therapies in HA and HB

Conventional Plasma-Derived and Recombinant FVIII and FIX Concentrates

The treatment of HA and HB has developed from transfusion of whole blood, plasma, and cryoprecipitate to more targeted replacement therapies under use of plasma-derived (pd) or recombinant (r) human FVIII or FIX concentrates.

When compared with CSA, lower OSCA-based FVIII activities have been observed for highly purified plasma-derived as well as recombinant full-length (rFL)- or B-domain-deleted (BDD)-FVIII concentrates. It is noteworthy that according to specifications of the European Pharmacopoeia, potency labeling of FVIII concentrates in Europe is usually based on results from CSAs against the WHO IS for factor VIII concentrate (NIBSC code: 07/350). Thus, depending on the applied FVIII concentrate, significantly lower FVIII activity levels may be observed when using OSCAs rather than CSAs for the analysis of post-infusion samples. Such discrepancies, which are mainly attributable to assay-specific differences in phospholipid composition, usually do not exceed 20% for highly purified pd- or rFL-FVIII concentrates and, therefore, are considered to be clinically irrelevant.

Higher differences have been observed for certain recombinant BDD (rBDD)-FVIII concentrates which lack the FVIII B-domain. Moroctocog alfa (ReFacto® [AF]) has up to 50% lower activity when measured by OSCAs, with strong dependency on the aPTT reagent. To address this particular issue, a ReFacto AF Laboratory Standard for testing of plasma samples from patients receiving ReFacto AF has been made available.

In contrast to FVIII concentrates, potency labeling of FIX concentrates released to the European market is usually done using an OSCA in combination with the WHO IS for FIX concentrate (NIBSC code: 07/182). The IS for FIX as well as FVIII concentrate are plasma-derived preparations. It was demonstrated that rFIX concentrates yielded up to 30% lower activity levels in an OSCA calibrated against the IS for FIX concentrate when compared with a correspondingly calibrated CSA. Furthermore, also reagent-related discrepancies between applied OSCAs were found, while such effects were not observed when testing pd-FIX concentrates.

The aforementioned findings demonstrate that, due to structural and/or matrix-dependent differences between plasma-based assay calibrators and FVIII or FIX drug molecules, the introduction of recombinant factor concentrates led to assay discrepancies of varying degrees during both potency labeling and analyses of post-infusion samples. With the advent of modified recombinant EHL-FVIII/FIX concentrates, such discrepancies, as described below, have become more frequent.

Extended Half-Life (EHL) FVIII and FIX Concentrates

At present, a total of five EHL-FVIII concentrates (Damoctocog alfa pegol [Jivi®], Efmoroctocog alfa [Elocta®], Lonoctocog alfa [Afsyla®], Rurioctocog alfa pegol [Adynovia®], and Turoctocog alfa pegol [Esperoct®]) and three EHL-FIX concentrates (Albutrepenonacog alfa [Idelvion®], Eftrorncocog alfa [Alprolix®], and Nonacog Beta Pegol [Refixia®]) are available in Europe. Compared with conventional plasma-derived or recombinant factor concentrates, these EHL products have a 1.2- to 1.5-fold (FVIII, corresponding to the half-life of vWF) or 5-fold (FIX) prolonged half-life in the circulation. The modifications, introduced to achieve these characteristics (fusion with Fc fragment, albumin fusion or PEGylation) have various effects on the activation and/or activity pattern of these FVIII/FIX drug molecules. Consequently, divergent assessments of factor activity compared with plasma-derived FVIII or FIX used for assay calibration are observed. Potential assay discrepancies between commonly used test systems are described below for each factor concentrate. A synopsis is presented in Table 1.

Efmoroctocog Alfa (Elocta)

Efmoroctocog alfa (rFVIIIFc, Elocta) is a fusion protein combining a B-domain-deleted molecule and the Fc fragment of human immunoglobulin G1 (IgG1), to extend the circulatory half-life.

In an initial international field study based on FVIII deficiency plasma spiked with rFVIIIFc at different target concentrations, results obtained with various OSCAs showed good recovery and were comparable to those of correspondingly prepared rFL-FVIII preparations (Advate) analysed in parallel. During a more recent UK National External Quality Assessment Scheme for Blood Coagulation (NEQAS BC), based on spiked as well as post-infusion samples, the initial findings could be confirmed, again for a broad spectrum of applied OSCAs/aPTT reagents. In contrast to the OSCA results, higher results for rFVIIIFc are obtained when samples are analysed by CSAs. It was assumed that these discrepancies stem from differences between the WHO IS for FVIII concentrate or plasma. However, it is generally concluded that both the OSCA and the CSA may be used to safely monitor rFVIIIFc-based factor FVIII replacement therapies. However, overestimation of Elocta by CSAs should be considered.

Damoctocog Alfa Pegol (Jivi)

Damoctocog alfa pegol (Jivi) is a rBDD FVIII molecule with extended half-life due to site-specific PEGylation.

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Table 1 Summary of EHL/modified FVIII/FIX concentrates and applicable test systems

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<thead>
<tr>
<th>EHL-FVIII concentrates</th>
<th>EHL-FIX concentrates</th>
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<tr>
<td>Adynovi</td>
<td>afstyla</td>
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<tr>
<td>Eflocta</td>
<td>Esperoct</td>
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<tr>
<td>Jivi</td>
<td>Obizur</td>
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<tr>
<td>Alprolix</td>
<td>Idelvion</td>
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<td>Refixia</td>
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**OSC assays (aPTT reagents)**

<table>
<thead>
<tr>
<th>Activator: ellagic acid/polyphenols</th>
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<tbody>
<tr>
<td>Actin (Siemens)</td>
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<tr>
<td>Actin FS (Siemens)</td>
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<tr>
<td>Actin FSL (Siemens)</td>
</tr>
<tr>
<td>SynthAFax (IL Werfen)</td>
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<tr>
<td>Cephascreen (Stago)</td>
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</table>

| Activator: silica                   |
| Pathromtin SL (Siemens)             |
| APTT SP (IL Werfen)                 |
| SynthASil (IL Werfen)               |
| STA PPT A (Stago)                   |
| Activator: kaolin                   |
| C.K. Prest (Stago)                  |

| FVIII CS assays                     |
| Factor VIII chromogen (Siemens)     |
| COAMATIC FVIII (Chromogenix/IL Werfen)|
| Biophen FVIII (Hyphen-BioMed)       |
| Electrachrome FVIII (IL Werfen)     |
| Technochrom Factor VIII:C (Technoclone) |

| FIX CS assays                       |
| BIOPHEN Factor IX (Hyphen)          |
| ROX Factor IX (Rossix)              |

Abbreviations: aPTT, activated partial thromboplastin time; EHL, extended half-life.

- Acceptable recoveries described in the literature, may be applied for analysis of post-infusion samples. (✓)
- Acceptable recoveries found in field study but contradictory information available. (✓)
- Moderate and/or system-dependent discrepancies described. (✓)
- Not recommended for analysis of post-infusion samples due to described significant discrepancies. (✓)
- No individual or only insufficient data available. (✓)
- *2 manufacturers’ recommendation to apply a conversion factor of 2 (see text for potential restrictions). (✓)
- Overestimation observed. (✓)
- Underestimation observed. (✓)

*C2 manufacturers’ recommendation to apply a conversion factor of 2 (see text for potential restrictions).
During an international field study, a variety of FVIII OSCAs and CSAs were investigated using FVIII-deficient plasma spiked with various Jivi target concentrations. Overall, good recovery and agreement of results was stated for the applied CSAs (Siemens FVIII Chromogenic Assay, Electrachrome FVIII, and COAMATIC FACTOR VIII) within this study design. However, this was also true for most of the investigated OSCAs, except for SynthAFax and Actin FS, which showed pronounced overestimation (25–40%) of both FVIII preparations by CSAs was observed at lower PEG-rFVIII or Advate input concentrations. Overall, however, there appeared to be good comparability of measurement results between PEG-rFVIII and the conventional FVIII product Advate using a wide range of measurement systems.

In contrast to the results described earlier, another field study showed a more significant (concentration-dependent) overestimation of Adynovi when using both OSCAs and CSAs. Interestingly, this effect was also seen using product-specific calibrators. It is noted by the authors that, unlike the initial study described earlier, no congenital FVIII-deficient plasma was used for sample preparations. Furthermore, also a narrower repertory of reagents was compared within that study.

Nevertheless, since the currently available literature contains inconsistent information regarding the reliability of test systems for the determination of Adynovi plasma activity, individual evaluation of local assays may be considered.

**Rurioctocog Alfa Pegol (Adynovi)**

Rurioctocog alfa pegol (PEG-rFVIII, BAX 855, Adynovi) is a rFL-FVIII molecule conjugated with a polyethylene glycol (PEG) reagent (approximately 60% attached to the B-domain) for half-life extension.

In an initial international field study based on congenital FVIII-deficient plasma spiked with PEG-rFVIII at different input concentrations, obtained results showed concentration-dependent overestimation of PEG-rFVIII. However, comparable results were obtained with analogous rFL-FVIII preparations (Octocog alfa [Advate®]) tested in parallel. Especially for OSCAs that utilise aPTT reagents based on ellagic acid or polyphenol, overestimation (25–40%) was observed at lower PEG-rFVIII or Advate input concentrations. When silica- or kaolin-based aPTT reagents were used, target activity values were found. In contrast, overestimation (25–30%) of both FVIII preparations by CSAs was observed at higher PEG-rFVIII input concentrations. Overall, however, there appeared to be good comparability of measurement results between PEG-rFVIII and the conventional FVIII product Advate using a wide range of measurement systems.
Regarding analysis by different OSCAs, inconsistent results have been described, especially for the aPTT reagents SynthASiL and SynthAFax (IL Werfen). Significant underestimation was found when APTT-SP (IL Werfen), TriniCLOT (Stago), or STA PTT-Automate (Stago) was used.\(^{60,62}\) Accordingly, these aPTT reagents should not be applied for the analysis of Esperoct by OSCAs (►Table 1). As being true for Elocta, overestimation of Esperoct by CSAs should be considered (►Table 1).

**Albuprepotenacog Alfa (Idelvion)**

Albuprepotenacog Alfa (rIX-FP, Idelvion) is a recombinant fusion protein combining a FIX molecule with albumin for extended half-life.\(^{63}\)

In several studies, Pathromtin SL, which is also used for potency labeling of Idelvion, has been shown to be well suited as aPTT reagent in FIX OSCAs for monitoring of post-infusion samples.\(^{64,65}\) Although inconsistent results have been observed, most of the other applied aPTT reagents showed acceptable results, with the exception of Actin FS and CK Prest, that showed significant underestimation of Idelvion, while, in contrast, significant overestimation was found for SynthAFax as well as for the available FIX CSAs.\(^{66-68}\)

Interestingly, it has been recently proposed that the latter is due to impaired activation of rFIX-FP during Pathromtin SL (and colloidal silica)-based OSC measurements.\(^{69}\)

On the basis of the data published to date, an OSCA for the measurement of Idelvion appears to be preferable; however, the above-described aPTT reagents should not be applied (►Table 1).

**Eftrenonacog Alfa (Alprolix)**

Eftrenonacog alfa (Alprolix) is a recombinant fusion protein with extended half-life that combines a FIX molecule with the Fc fragment of human IgG1.\(^{70}\)

While potency labeling of Alprolix is done using an OSCA based on Actin, overestimation of Alprolix especially at lower concentrations was found when using OSCAs that apply ellagic acid–based aPTT reagents. While this was especially and reproducibly true for SynthAFax, a high interlaboratory variability of results was found for other aPTT reagents tested.\(^{71,72}\) In contrast to the ellagic acid–based aPTT reagents, silica-based reagents showed more acceptable results, while the kaolin-based CK Prest demonstrated significant underestimation. When using a CSA for sample analysis, acceptable recoveries were found.\(^{72}\) However, a recent single-centre study revealed analyser-dependent differences of assay results, as well as underestimation of Alprolix by CSAs at lower input concentrations.\(^{73}\) Furthermore, another field study also demonstrated interlaboratory differences during analysis of Alprolix, which, however, were not different from that of pFIX and rFIX processed in parallel.\(^{74}\)

In summary, one may conclude that, due to over- or underestimation of Alprolix, by SynthAFax or CK Prest, respectively, these two aPTT reagents should not be used in OSCAs applied for the determination of Alprolix activity. Furthermore, local evaluation of (other) ellagic acid–based aPTT reagents should be performed (►Table 1).

**Nonacog Beta Pegol (Refixia)**

Nonacog Beta Pegol (Refixia) is a recombinant FIX molecule with site-specific (glyco)PEGylation for half-life extension.\(^{75}\)

Laboratory studies showed that the application of FIX CSAs yields acceptable results when measuring Refixia in post-infusion samples.\(^{76}\) In contrast, a high variability of results was found when applying a series of different OSCAs. When using polyphenol- or ellagic acid–based aPTT reagents, acceptable results (STA-Cephascreen, SynthAFax, or DG Synth), but also significant underestimation of Refixia (e.g., using Actin FS and Actin FSL), were found. Underestimation was also found when using kaolin-based CK Prest. Regarding the silica-based aPTT reagents, however, SynthASiL (colloidal silica) led to an underestimation of more than 50%, while, as also true for most of the other silica-based reagents, Pathromtin SL (silicon dioxide particles) led to severe overestimation of Refixia.\(^{77}\)

As a cause for the latter effect, it could be demonstrated that the presence of silica leads to an activation of Refixia by FXIa but also kallikrein already during the incubation phase of the aPTT reagent in the absence of Ca\(^{2+}\) ions.\(^{78}\)

Thus, as outlined above, and also summarised in ►Table 1, CSAs and only selected OSCAs (aPTT reagents) appear to be suitable for reliable determination of Refixia activity in post-infusion samples.\(^{79,80}\)

**Laboratory Diagnostics during Treatment of HA with Emicizumab (Hemlibra)**

Emicizumab (Hemlibra®) is a subcutaneously administered, humanised, monoclonal, bi-specific IgG4 antibody that binds FIX(a) and FX(a), thereby mimicking the cofactor function of FVIIIa.\(^{81}\) In contrast to FVIIIa, emicizumab circulates at high (target) concentrations (30–80 µg/mL [200–530 nM] as measured in plasma) and binds only with moderate affinity (KD = 1–2 µM) and non-discriminatory to the human (pro)enzyme variants of FIX(a) or FX(a). Simultaneous binding of emicizumab to FIXa and FX, both presenting properly (pro)enzyme variants of FIX(a) or FX(a), thereby mimicking the cofactor function of FVIIIa.\(^{81}\)

In contrast to FVIII, emicizumab does not require activation and is not subject to regulation. The former characteristic leads to significant shortening of aPTT-based clotting times even at subtherapeutic plasma concentrations of emicizumab. The reason is that emicizumab requires no thrombin-mediated feedback activation but is ‘active’ as it is. Accordingly, in the presence of emicizumab, aPTT-based OSCAs configured and calibrated for quantitative detection of plasmatic coagulation factor activity exhibit clotting times outside the dynamic range of these test systems.\(^{84,85}\)
In contrast, emicizumab at therapeutic concentrations is dynamically detected by FVIII CSAs based on human FIXα and FX (cf. Fig. 18). However, it is important to note that any determined ‘FVIII activities’ must not be translated to FVIII-equivalent effects of emicizumab in vivo, but are merely derived from assay calibration against FVIII. A different picture emerges when using FVIII CSAs based on bovine factors. Such assays are not able to detect emicizumab, as it only binds to human FIX(a) and FX(a), and are therefore specific tools to measure (endogenous or exogenous) FVIII in the presence of emicizumab. However, assays to monitor the combined effect of FVIII and emicizumab are currently not available.

**Determination of Plasma Levels of Emicizumab**

To enable the determination of emicizumab in plasma, a modified FVIII OSCA has been developed. Using a higher prediluted plasma sample and appropriate calibrators and controls, it determines plasma concentrations (activity) of emicizumab between 10 and 100 μg/mL, adequately covering the therapeutic range of the drug.

Using emicizumab calibrators, quantitative determination of emicizumab also appears possible using human factor-based FVIII CSAs. It must be noted that any additionally present FVIII may distort measurement results especially of this assay type. Should it be necessary to determine the plasma level of emicizumab in the presence of FVIII, heat inactivation of the plasma sample may be considered.

**Laboratory Diagnostics in HA or HB with Inhibitors**

Neutralising anti-FVIII (drug) antibodies, so-called inhibitors, occur in up to 30% of treated patients with severe HA. In contrast, the formation of FIX inhibitors in severe HB is much less common.

In case of HA or HB with inhibitors, bypassing agents such as recombinant activated FVII (rFVIIa) or aPCCs may be applied. In HA with inhibitors, emicizumab has become a treatment option. In addition, recombinant porcine FVIII (rpFVIII, Susoctocog alpha [Obizur®]) has been approved for the treatment of bleeding episodes in adults with acquired HA (AHA).

To eliminate inhibitors formed in treated HA or HB patients, immune tolerance induction (ITI) therapy is currently the therapy of choice. Hereby, FVIII or FIX concentrates are administered at high frequencies and doses. While up to 70% of HA patients with inhibitors are in remission after ITI therapy, this strategy is less successful in patients suffering from HB with inhibitors.

Functional detection and quantification of FVIII or FIX inhibitors is usually done using the (N)BA, this raises the question whether or not established standard conditions for heat inactivation of patient samples to eliminate any remaining factor activities (Fig. 2) also apply for EHL-FVIII or FIX molecules. Indeed, corresponding analysis confirmed that the activity of all currently approved EHL-FVIII concentrates can be efficiently inactivated when incubated at 56 °C for 30 minutes.

**Monitoring of and Inhibitor Testing in the Treatment of AHA with Susoctocog Alfa (Obizur)**

Susoctocog alfa (Obizur) is a rpFVIII concentrate approved for the treatment of acute bleedings in AHA. In contrast to human (EHL-)FVIII concentrates, however, potency labeling of Obizur uses a FVIII OSCA rather than a CSA. In an international field study, based on congenital FVIII-deficient plasma spiked with Obizur at different input concentrations, OSCAs with a variety of aPTT reagents showed acceptable results. In contrast, applied CSAs underestimated the activity...
of Obizur by approximately 50%.\textsuperscript{55} Accordingly, a FVIII OSCA should be used for both, monitoring of Obizur plasma levels during therapy as well as determination of (residual) FVIII activities in Obizur-specific NBAs (\textit{\textsuperscript{\textdegree}Table 1}).

Indeed, while Obizur has been developed with the intention to bypass neutralising antibodies against human FVIII, cross-reactivity of such inhibitors with the underlying rFVIII molecule has been described.\textsuperscript{101} Thus, it may become necessary to perform specific testing for inhibitors against Obizur. For doing so, a corresponding laboratory standard is available from the manufacturer (Takeda). When diluted to 1 U/mL in (imidazole-buffered) FVIII-deficient plasma, this mixture, instead of FVIII in buffered pooled plasma (\textit{\textsuperscript{\textdegree}Fig. 2A}), serves as substrate during the NBA.

**Outlook to Future Treatment Options**

In addition to currently available treatment options for HA and HB, a variety of novel molecules and innovative therapy strategies is currently investigated in a series of clinical trials. Among others, these comprise FVIII/FIX gene therapy, factor concentrates with further improved properties and/or the possibility for subcutaneous application as well as strategies for rebalancing pro- and anticoagulant mechanisms.\textsuperscript{102–105} As also true for the currently established therapies, also these novel treatment regimens do and will prompt questions about the general need for treatment monitoring but also about their impact on assay results as well as the applicability of established test systems for patient sample analysis.\textsuperscript{11,106}

Currently, five (three HA and two HB) gene therapy approaches are within phase 3 clinical trials, making a licensed product possible within the near future.\textsuperscript{8}

In gene constructs used for HA, the factor VIII cassette is B-domain deleted, consistently resulting in OSCA results which were approximately 1.5 times higher than those obtained with chromogenic assays.\textsuperscript{107} Interestingly, this contrasts with the factor level discrepancies found in patients treated with BDD FVIII concentrates, where OSCAs generally yields lower results than CSAs.\textsuperscript{36,65}

In gene therapy for HB, a full-size FIX gene with higher specific activity than the native FIX gene (Padua variant) is most often used. FIX OSCAs yield approximately 1.5 times higher results than chromogenic assays, while aPTT-reagent-dependent differences have been observed.\textsuperscript{108}

Further research is needed to determine the relative accuracy of the different assay principles in patients with gene therapy in relation to clinical response and also in relation to the results of patients treated with different factor concentrates. Recently, it has been recommended to apply both assay principles to quantify FVIII and FIX activity in gene therapy trials and that the possibly more accurate lower values measured by the chromogenic test be relied upon.\textsuperscript{10}

**Summary and Conclusion**

High inter-laboratory variation in the measurement specifically of EHL-factors in post-infusion samples has become apparent. To some extent, this variation is considered to be within the range of ‘traditional’ (but previously less robustly examined) laboratory result variation, as observed for the measurement of conventional recombinant factor molecules. However, it was our aim to highlight that there are inappropriate test systems for certain factors, or inappropriate factors for certain test systems. Such combinations must be avoided to maintain patient safety. The treating physician and the laboratory need to harmonise their toolboxes to ensure that factor activities are assessed within acceptable ranges for all factor concentrates prescribed in the respective centre. As a minimum requirement, significant overestimation of factor activity must be excluded to ensure appropriate laboratory-based controls for prophylactic and therapeutic factor administration. While product-specific calibrators may be helpful to correct for potential assay discrepancies, this approach appears to be impractical in clinical routine due to the variety of available concentrates. However, OSCAs will still meet most requirements. Emicizumab poses considerable additional challenges, especially for small- and medium-size coagulation laboratories, because the use of CSAs becomes mandatory when examining samples from HA patients treated with this antibody. In the end, coagulation laboratories processing samples for HA and HB patients under factor replacement or alternative therapy will have to establish two different assays for FVIII and FIX, respectively: one OSCA and one CSA—at least. With new therapeutic options lying ahead, this will become more pressing during the next years.

**Conflict of Interest**

The authors declare that they have no conflict of interest.

**References**

14 Wilmot HV, Rakowski K, Gray E. The traceability of commercial plasma calibrators to the plasma international standards for factor VIII and factor IX. Int J Lab Hematol 2020;42(06):810–818
22 Oldenburg J, Pavlova A. Discrepancy between one-stage and chromogenic factor VIII activity assay results can lead to misdiagnosis of haemophilia A phenotype. Hemostaseologie 2010;30(04):207–211
36 Poupard C, Ternisien C, Desconclois C, Lasne D, Aillaud MF, Caron C. Discrepancies between one stage assay and chromogenic substrate assay in patients treated with recombinant or plasma-derived FVII and usefulness of a specific assay in ReFacto A®-treated patients. Haemophilia 2016;22(02):e101–e103
39 Wilmot HV, Gray E. Potency estimates for recombinant factor IX in the one-stage clotting assay are influenced by more than just the choice of activated partial thromboplastin time reagent. Haemophilia 2018;24(05):e363–e368
46 Church N, Leong L, Katterle Y, et al. Factor VIII activity of BAY 94-9027 is accurately measured with most commonly used assays: results from an international laboratory study. Haemophilia 2018;24(05):823–832
69 Rosén S, Bryngelhå P. FIX potency of rIXFusion protein is underestimated by one-stage methods using silica-based APTT reagents. Haemophilia 2020;26(02):340–345
81 Sampezi I, Isgawa T, Soeda T, et al. Identification and multidimensional optimization of an asymmetric bispecific IgG antibody


86 Amir J, Seghatchian J. Usefulness of chromogenic assays for potency assignment and recovery of plasma-derived rFVIII and FIX concentrates or their recombinant long acting therapeutic equivalents with potential application in treated pediatric hemophilic patients. Transfus Apheresis Sci 2018;57(03):363–369.


