The NFkB Signaling Pathway Is Involved in the Pathophysiological Process of Preeclampsia

Der NFkB-Signalweg ist am pathophysiologischen Prozess der Präeklampsie beteiligt

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ABSTRACT

The high prevalence of preeclampsia (PE) is a major cause of maternal and fetal mortality and affects the long-term prognosis of both mother and baby. Termination of pregnancy is currently the only effective treatment for PE, so there is an urgent need for research into its pathogenesis and the development of new therapeutic approaches. The NF κ B family of transcription factors has an essential role in inflammation and innate immunity. In this review, we summarize the role of NF κ B in normal and preeclampsia pregnancies, the role of NF κ B in existing treatment strategies, and potential NF κ B treatment strategies.

ZUSAMMENFASSUNG

Die hohe Prävalenz von Präeklampsie (PE) ist eine wesentliche Ursache mütterlicher und fetaler Mortalität und wirkt sich auch auf die Langzeitprognose von Mutter und Kind aus. Der Schwangerschaftsabbruch stellt zurzeit die einzig effektive Therapie gegen PE dar. Damit besteht ein dringender Bedarf nach weiterer Forschung zur Pathogenese von PE sowie zur Entwicklung neuer Therapieansätze. Die NFkB-Familie der Transkriptionsfaktoren spielt eine wichtige Rolle in Entzündungsprozessen und für die angeborene Immunität. In dieser Übersichtsstudie fassen wir die Rolle von NFkB bei normalen und Präeklampsie-Schwangerschaften sowie die Bedeutung von NFkB in bereits existierenden Therapien und potenzielle NFkB-Behandlungsstrategien zusammen.

Introduction

Preeclampsia (PE) is a hypertensive disorder of pregnancy (HDP), characterised by oedema, proteinuria, and hypertension-related symptoms, and can only be diagnosed after the 20 th week of pregnancy [1]. PE is a common syndrome in pregnancy, affecting approximately 3-5% of pregnancies worldwide. The incidence of PE has been increasing for nearly 30 years [2]. It can lead to fetal growth restriction (FGR), abortion, low birth weight and other fetal and neonatal complications. Besides, it can adversely affect the long-term prognosis of mother and baby and even lead to maternal and infant death, such as increased risk of chronic hypertension, diabetes mellitus, ischemic heart disease, cerebrovascular disease, kidney disease, thromboembolism, hypothyroidism, and even impaired memory in the later maternal period and [3] increased risk of obesity in offspring and others metabolic diseases with PE. When PE presents with severe symptoms of eclampsia or haemolysis, elevated liver enzymes and low platelets (HELLP syndrome), maternal mortality is even higher. Some researchers have found that the onset of PE is related to maternal immune imbalance and placental inflammation [4]. However, the pathogenesis of PE is not fully understood in humans because of this special period of pregnancy. The only treatment currently available is termination of the pregnancy. Low-dose aspirin in early pregnancy has been shown to prevent PE, but only if taken before 16 weeks' gestation [5, 6, 7]. Thus, it is crucial to screen during the first trimester in order to effectively prevent PE [8]. However, due to the lack of comprehensive understanding of the pathogenesis of PE, existing screening protocols are inaccurate. It is therefore important to have a thorough understanding of the pathogenesis of PE.

PE has a complex pathogenesis involving a number of different mechanisms. The main causes of PE are oxidative stress at the maternal–fetal interface, maternal inflammation, insufficient remodelling of the uterine spiral arteries and vascular endothelial injury [9, 10, 11]. All these mechanisms are mediated by NF κ B. During pregnancy, women experience a physiological inflammatory response that can promote remodelling of the uterine spiral arteries by regulating trophoblast infiltration. Overactivation of the inflammatory response leads to immune imbalance and vascular endothelial damage, which promotes the development of pregnancy complications associated with PE [2, 12]. Therefore, in this review, we systematically elaborate on the role of NF κ B in the development and treatment of PE.

The NFkB family consists of seven proteins, RelA (p65), RelB, c-Rel, NFB1 (p105/p50), and NFB2 (p100/p52), which are encoded by five distinct genes and affect the expression of over 400 genes [13, 14]. It plays an important role in normal and complex pregnancies by regulating pathways such as inflammation, oxidative stress, cell proliferation, differentiation, apoptosis, and angiogenesis [15]. In resting cells, NFkB is present in the cytoplasm and bound to its inhibitor (IkB), which dissociates by phosphorylation in an environment rich in reactive oxygen species and cytokines. NFkB is translocated from the cytoplasm to the nucleus where it recognises and binds to specific DNA sequences and acts as a transcription factor regulating the expression of inflammatory factors and MMPs (Matrix Metalloproteinases) [16, 17]. There are studies suggesting that the development of PE may be a familial risk,

which may be due to genetic and epigenetic changes [18, 19]. Compared to women with no family history of PE, NFkB expression in PE is increased by 23.35 percent [20].

NFκB Signal Pathway in Normal Gestation and PE

Semi-allogeneic fetuses exposed to maternal immune systems during pregnancy are usually not rejected and develop in the mother's womb until birth. The fetal–maternal interface is the site of regulation of this substantial immune tolerance and the placental chorionic villi are critical for this management. For a normal pregnancy, inflammation must be tightly controlled. Macrophages help to maintain pregnancy by promoting tolerance to semi-allogeneic fetuses and maintaining the homeostatic environment necessary for healthy fetal development [21, 22]. Macrophage differentiation can also be regulated by the NFKB pathway.

Normal gestation

In order to help control the window of implantation, NFkB is upregulated in the decidua prior to fertilisation. When the embryo implants, there is a severe inflammatory response, which may be triggered by the production of paternally derived alloantigens in the embryo [23]. The uterus releases pro-inflammatory cytokines during the implantation window to activate and attract numerous immune cells to the endometrium, causing local endometrial injury that aids effective implantation [24, 25]. Studies support the idea that endometrial biopsies performed during the natural cycle prior to IVF treatment can significantly increase the likelihood of implantation and clinical pregnancy [25]. Furthermore, higher NFkB expression in the first trimester is associated with increased MMP expression. MMPs are important extracellular matrix (ECM) remodeling proteinases that control uterine remodeling, a process that is essential for healthy pregnancies [26]. High MMP-9 levels promote uterine ECM degradation and relax intercellular connections, promoting extravillous trophoblast cells to invade and uterine spiral artery remodeling [27, 28].

NFkB expression is also increased in late pregnancy, which can enhance cervical maturation, induce membrane rupture, and cause uterine contractions [15, 29]. When labor begins, inflammatory cytokines in the chorion are increased and act on the myometrium via the NFkB pathway, boosting oxytocin receptor expression and starting labor [30]. At the same time, MMP activates at the start of labor by lysing the cytoskeletal substrate, triggering muscular contraction. By inhibiting the biological role of progesterone, a pregnancy-maintaining factor, and promoting the expression of oxytocin receptors and prostaglandin synthase, myometrial inflammation plays a significant role in labor [31, 32, 33].

During pregnancy maintenance, maternal inflammatory levels were largely suppressed and NFkB transcript levels decreased, which was associated with increased regulatory T cells. A subset of T cells called "Treg" cells play a role in regulating the immune system and preventing organ rejection [34, 35]. Th1 and Th17 effector cells differentiate into regulatory T cells at this stage and mediate the development of immune tolerance [29]. The expression of the Foxp3 gene and cytokines (e.g. IL-2 binding) are associated

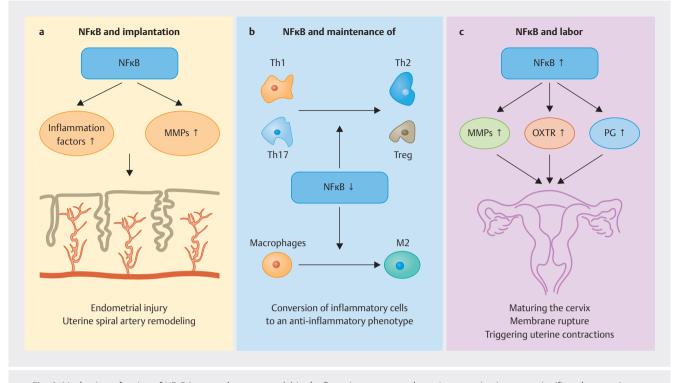


Fig. 1 Mechanism of action of NFκB in normal pregnancy. (a) In the first trimester, pre-eclampsia expression increases significantly, secreting a large number of inflammatory factors and MMPs, promoting endometrial damage and spiral artery remodelling, and enabling embryos to implant successfully. (b) During pregnancy maintenance, NFκB levels decrease, immune cells differentiate towards the anti-inflammatory phenotype, Th1 and Th17 differentiate into T2 and Treg cells, and macrophages differentiate towards the M2 phenotype, inducing maternal immune tolerance to the fetus. (c) NFκB levels rise again in the third trimester, promoting the expression of prostaglandins, MMPs and oxytocin receptors, inducing cervical maturation, leading to rupture of the membranes and uterine contractions.

with the differentiation of Treg [36]. Meanwhile, placental macrophages differentiate towards the M2 phenotype under NFkB mediation. M2 macrophages promote tissue repair, angiogenesis and homeostasis, whereas M1 macrophages are generally considered to be anti-inflammatory. The intense maternal inflammatory response is thus suppressed [37, 38] (► Fig. 1).

PE pregnancy

 $\mathsf{NF}\mathsf{\kappa}\mathsf{B}$ is generally activated at the beginning of pregnancy by the action of IKKa and IKKB proteins [39]. NFkB inhibitors are degraded by the combined action of IKK α and IKK β , together with IKKγ. IKKα is a key activator of NFκB in the canonical and noncanonical pathways. CK2 α , the catalytic subunit of the CK2 protein, is the key activator of the atypical NFkB activation pathway [40]. Activators (IKKα and CK2α) are decreased and inhibitors (I κ B β and I κ B α) are increased in PE placenta, which may mean that all three NFkB activation pathways are downregulated in PE [14]. Interestingly, the role of IKK α depends on its localization, i.e., activating NFkB in the cytoplasm and terminating NFkB-mediated gene expression in the nucleus [41]. Therefore, depletion in the cellular IKKα level may influence the nuclear transcriptional activity of NFkB, leading to elevated concentrations of factors whose genes are regulated by NF κ B (\triangleright Fig. 2a). One possibility is that the NF κ B inhibitors whose levels are elevated in this paper may favor NFkB activation by switching from an inhibitory role to a

chaperone-like function, thus supporting the transport of NFκB in an inactive form into the nucleus [14]. The levels of p53/RSK1 in HTR8/SVneo cells cultured with PE serum were elevated. The p53/ RSK1 complex is known to activate NFκB by its phosphorylation at Ser536, independent of the cytoplasmic degradation of the kappa B inhibitor. This suggests that factors existing in the serum of preeclamptic women influence the activation of NFκB by the p53/ RSK1 pathway in hypoxic conditions [42] (**>** Fig. 2b). However, experimental data were obtained from placentas obtained during late pregnancy. The activation pathway of NFκB in early and midpregnancy is still worth exploring.

Several studies have shown that, compared to normal pregnancy, NF_KB in the placenta and maternal circulation of PE women is significantly increased, which leads to an increase in maternal inflammation levels [43, 44, 45, 46]. Hofbauer cells (HBCs) are natural macrophages of the human placenta, predominantly with an anti-inflammatory M2 phenotype. However, under the influence of persistent inflammation during pregnancy, there is a transition to the M1 phenotype, which may be based on an increase in NF_KB expression [47]. Syncytiotrophoblast dysfunction is a key feature in PE, reports have observed disruption of syncytiotrophoblast apical microvilli, indicating a loss of apical polarity, which can be disrupted by pro-inflammation [48].

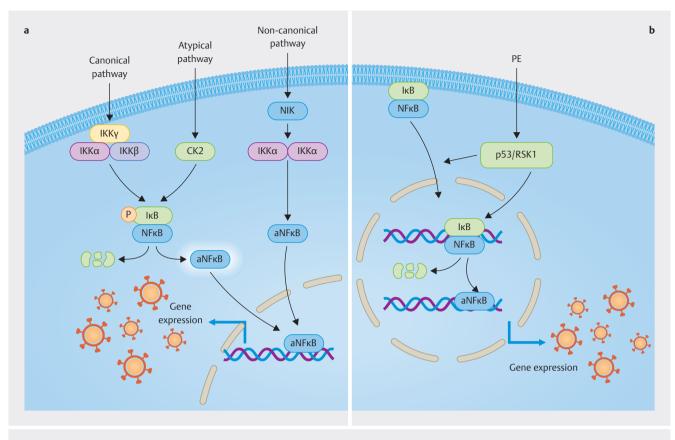


Fig. 2 NFκB activation pathway in cells. (a) Canonical, atypical, and non-canonical activation pathways for NFκB. IKKα, IKKβ, and IKKγ are activators of NFκB in the canonical and non-canonical pathways. CK2 is the activator of the atypical NFκB activation pathway. IκB is an inhibitor of the NFκB pathway. (b) Activation pathway of NFκB in HTR8/SVneo cells cultured with PE serum [40].

Pro-inflammatory or Anti-inflammatory Factors Affect NFκB Signaling Pathway

The NFKB signaling pathway is regulated by a variety of regulators in human. The risk factors for PE are similar to those for cardiovascular diseases, such as elderly pregnancy, obesity, metabolic disorders, kidney disease, autoimmune diseases [49], and a family history of preeclampsia. These factors can lead to placental inflammation. However, the exact aetiology of PE is unclear, but it has been hypothesized that while genetic and environmental factors both affect how the condition develops, genetic factors have a greater impact [50]. In addition, a number of special pathogens, such as periodontal pathogens and gut microbes, are also involved in the development of PE through the NFKB pathway [11, 51].

Periodontal pathogens

A large number of clinical studies have found a positive correlation between periodontitis and the incidence of PE [52, 53]. One of the "key pathogens" in the aetiology of periodontitis is Porphyromonas gingivalis, which is also the most common pathogenic bacterium in amniotic fluid and placental tissue [54, 55]. Transmission of specific periodontal bacteria to placental tissue via the bloodstream stimulates upregulation of Toll-like receptor 4 (TLR-4) expression and downregulation of Peroxisome proliferator-activated receptor γ (PPAR- γ) expression. This in turn increases NF κ B activity in placental tissue, ultimately promoting the development of PE [51, 56]. The possibility that these bacteria are part of the placental microbiome is controversial, as there is no agreement on whether the placental microbiome is present in healthy full-term pregnancies [57, 58]. However, periodontal disease during pregnancy is significantly associated with unfavorable short- and longterm offspring outcomes, which may be due to epigenetic changes [57]. Early diagnosis and treatment of periodontal disease in the first trimester or even before conception may be helpful in preventing and effectively minimising problems with conception and obstetrical issues [59, 60].

Gut microbe

Studies have shown that patients with PE have significant dysbiosis and a reduced diversity of gut bacteria. Among these, the intestinal flora associated with LPS production was significantly elevated in the intestinal flora of the PE group. Alterations in the gut microbiota may alter the profile of short-chain fatty acids released by bacteria, potentially leading to metabolic syndrome and hypertension [61, 62, 63, 64]. Early pregnant rats exposed to ultra-low doses of lipopolysaccharide (LPS) develop PE due to placental TLR4 activation [65]. LPS induces Prostaglandin-endoperoxide synthase 2 (PTGS2) and LncRNA BC030099 manipulates the inflammatory response via the NFκB pathway [33, 66].

Genetic factors

Placental omics analysis provides clues for exploring the occurrence and therapeutic targets of PE at the genetic level. Daniel Vaiman et al. identified 16 specific genes associated with placental disease, such as PKN3, PTTG1, etc., through a new method, which are involved in the development of PE through various pathways [67]. The NFKB pathway can be activated by the expression of inflammation-related genes such as NEMO, which is a key regulator of NFKB activation [68]. Agata Sakowicz et al. suggest that the presence of identical NEMO gene variants in the maternal and fetal genomes may increase the likelihood that PE offspring will develop PE [69]. This view may provide new insights into the genetic causes and pathogenesis of PE.

Over the past few years, microRNAs (miRNAs) have emerged as important regulators of gene expression in inflammatory and immune responses. Numerous miRNAs modify the activity of NFkB either by targeting upstream and downstream NFkB activating kinases or other NFkB signaling elements [13]. For example, MiR-517–3 p, which has high levels in the placenta of PE, targets the mRNA encoding TNFAIP3-interacting protein 1 (TNIP1), an inhibitor of the NFkB pathway, to activate NFkB pathway. Activation of NFkB increases production of the cytokine TNF superfamily member 15 (TNFSF15), leading to the upregulation of anti-angiogenic soluble vascular endothelial growth factor receptor 1 (sFlt-1). The Cajal bodies (CBs) are upstream regulators of miR-517-3 p, highly expressed in PE [13, 70]. By suppressing the production of HDAC2, miR-23a was able to stimulate the NFkB pathway [71]. In addition to inhibiting downstream phosphorylated AKT (p-AKT) and NFkB expression, unidirectional miR-219a can also inhibit downstream VEGF/NFkB signaling to inhibit trophoblast proliferation and invasion [72]. And other microRNAs, such as miR-21, miR-155, miR-31-5 p and miR-138, are involved in the development of PE through the NFkB pathway [73, 74, 75, 76, 77].

Despite lacking the ability to encode proteins, long non-coding RNAs regulate disease progression, possibly by affecting microRNA function [78, 79]. Placental overexpression of the lncRNA MIR503 host gene (MIR503 HG) is a microRNA-competitive endogenous RNA (ceRNA) whose cna inhibits the phosphorylation of IkB and the nuclear translocation of the NFkB signaling subunit p65 to regulate NFkB pathway [80].

Other factors

Many physical and social factors can also contribute to the development of PE, such as race or ethnicity, economic level, maternal disease, genetic polymorphisms and amniotic stretch [81, 82]. One interesting study found that polyhydramnios and multiple gestations were associated with an increased risk of preterm birth [83]. Justin et al. believe that stretching causes a downregulation of nuclear factor-E2-related factor 2 (Nrf2), accompanied by activation of the NFkB, leading to a pro-inflammatory state, which in turn leads to membrane thinning [84]. This may be explained by the subsequent infiltration of white blood cells that produce MMPs [85]. Studies have found that MTHFR (methylenetetrahydrofolate reductase) polymorphisms and diabetes can lead to impaired placental development [86], which may also be one of the reasons for the occurrence of PE, but the specific mechanism is still worth studying.

Protective factor

PPARγ, a transcription factor involved in glycolipid metabolism [87], is involved in placental cell metabolism, anti-inflammatory pathways, and oxidative stress [88]. PPARγ can regulate NFκB signal, inhibit its DNA binding activity, and promote its degradation [89, 90]. In multiple studies, PPARgamma appears to be down-regulated in the placenta of PE patients [91, 92]. Due to the decrease in PPAR-γ, NFκB is overactivated, resulting in the release of pro-inflammatory factors such as IL-6, IL-8, and TNF- α from the placenta [93], resulting in vascular dysfunction and maternal PE [56]. In addition, abnormal expression of the PPAR gene is also associated with increased cardiovascular risk in PE offspring. Decreased PPAR expression increases the sensitivity of blood vessels to Ang II. Adult children with complicated pregnancies are more sensitive to Ang II, resulting in endothelial dysfunction [94, 95].

Some interesting studies suggest that smoking may reduce the risk of PE [96, 97]. In animal studies, LPS-induced PE-like symptoms in pregnant rats are greatly reduced by nicotine activation of alpha7 nicotinic acetylcholine receptors (7nAChRs). In human blood samples have shown that activating 7nAChR reduces NFkB activation in monocytes from PE patients and balances the production of pro-inflammatory and anti-inflammatory cytokines [98, 99]. This idea has also been supported by in vitro experiments in which nicotine significantly reduced activation of NFkB and increased the survival of endothelial cells, which may reduce the likelihood of developing PE in smokers [100]. However, smoking is not regarded as a therapy option for PE due to the negative effects of nicotine on health.

NFκB Signaling Influences PE Development

Poor placental implantation and remodelling of the maternal spiral arteries are associated with PE. The inability of the placental spiral artery to remodel properly and the inappropriate infiltration of trophoblast cells into the maternal decidua are considered to be the early stages of PE formation. Impaired trophoblast activity hinders the remodelling of the uterine spiral artery, which may lead to placental oxidative stress and the release of inflammatory factors into the maternal circulation, causing systemic vascular endothelial damage and further PE-related pregnancy complications [22]. Therefore, many of the molecules that regulate trophoblast and endothelial function may influence the development of PE.

Trophoblast cell function

In PE, overexpression of Fas, ANKRD37 and triggering receptor expressed on myeloid cells-1 (TREM-1) contributes to the development of PE by preventing trophoblast cell invasion and migration, through the NF κ B pathway [101, 102, 103]. These substances cause NF κ B to be activated and interact with Inc-SLC4A1-1, resulting in an increase in CXCL8 mRNA expression. CXCL8 can promote TNF- α and IL1 β . The release of cytokines activates inflammation

related pathways, ultimately leading to increased cell apoptosis and reduced proliferation of trophoblasts, inhibiting their migration and vitality [104]. In addition, the activation of the NFkB pathway can also inhibit the invasion of trophoblast cells by increasing the production of PTEN [105]. By controlling NFkB pathway in PE, reducing the expression of PTGS2 and FPR2 can inhibit the pro-invasive and pro-inflammatory effects of LPS on trophoblast invasion [106, 107].

Matrix metalloproteinase

The activation of the PI3 K/AKT and ERK1/2 pathways in human trophoblast cells promotes NFkB translocation to the nucleus, activates MMP-9 promoter transcription after binding to DNA, and increases MMP-9 levels in trophoblast cells. Meanwhile, NFkB indirectly enhances MMP-2 activity by controlling enzymes associated with post-translational processing, such as increasing the expression of MT-MMP (model metalloproteinase) [17, 108]. Many studies have shown that MMP-2, MMP-9 are associated with uterine artery remodelling by promoting trophoblast cell proliferation, migration and invasion [109, 110]. There are significant differences in the levels of serum MMP-2 and MMP-9 at different stages of pregnancy. MMP-2 is most highly expressed in the first trimester and is involved in early placental formation. MMP-9 expression is lowest in the first trimester and highest in the second and third trimesters, which may be related to the promotion of placental and fetal development by MMP-9 [111]. After PE, serum MMP-2 levels are significantly increased and MMP-9 levels are significantly decreased [112]. Ongoing research shows that the expression of MMPs can be regulated by a variety of substances, such as JSH-23 and MIR503 HG, which can reduce MMP levels by inhibiting the NFkB signaling pathway [11, 66, 80]. Nuclear receptor coactivator 6. a transcriptional coactivator, can activate NFkB to induce MMP-9 transcription [113].

The levels of MMP-2 and MMP-9 are also associated with autophagy. Autophagy regulates trophoblast invasion by targeting NFkB activity [114]. NFkB signaling is generally thought to occur prior to autophagy activation [115], but studies have also shown that autophagy itself has the ability to degrade NFkB signaling components through various signaling pathways [116, 117, 118].

Endothelial cell function

eNOS can catalyze the synthesis of NO by vascular endothelial cells and maintain vasodilation by activating PKG [119, 120]. NFκB is a key factor in impairing vascular function and remodelling in human chronic inflammatory diseases by affecting endothelial progenitor cells (EPCs) and endothelial cell function [121]. Placental dysplasia can lead to placental dysfunction, resulting in the release of STBEV into the maternal bloodstream, which causes endothelial failure by activating NFκB, causing oxidative and nitrative stress and reducing eNOS expression and NO bioavailability [122]. Through interactions with IKK, GSTP1 can suppress NFκB signaling. This reduces iNOS expression and stimulates apoptosis, which controls NO-mediated ROS production [123]. By blocking the p38 MAPK/ NFκB pathway, cathepsin C downregulation improves HUVEC function. Endothelial cell dysfunction is protected by cathepsin C knockdown, which also offers a novel and promising strategy for the treatment of PE [9].

NFkB Pathway: Targets for Drug Therapy in PE

Common drugs act on NFkB

Many drugs can target NF κ B pathway to improve PE-like symptoms. Aspirin is the drug most commonly used clinically to prevent PE. Aspirin inhibits activation of the NF κ B pathway by preventing NF κ B nuclear translocation and binding to DNA motif elements or by binding to the IKK α protein as a competitive inhibitor of ATP [44, 124, 125]. Aspirin prevents redox-sensitive NF κ B/miR-155/ eNOS axis thereby reversing TNF α -mediated downregulation of eNOS and endothelial failure [126].

Magnesium sulphate is one of the most commonly used drugs in obstetrics and is often used to lower blood pressure, prevent eclampsia and protect fetal nerves. Magnesium sulphate also has an inhibitory effect on the NF κ B pathway [124, 127]. In animal studies, MgSO₄ was found to protect cranial nerves in PE rats by inhibiting the NF κ B/ICAM-1 signaling cascade [128]. In addition, magnesium salts can also reduce the placental hyperinflammatory response in an NF κ B-dependent manner [129].

Other anti-inflammatory, antineoplastic and vitamin supplementation therapies have also been tried to treat PE by targeting NFkB. Such as sulfasalazine and vitamin D supplementation, both have been shown to inhibit the NFkB pathway and reduce PE-like symptoms in animal models [130, 131]. Through the PPARy–NFkB axis, rosiglitazone can reduce trophoblast-associated inflammation [95]. Metformin (MET) has been shown to improve LPS-induced PE symptoms and placental damage. This process may inhibit TLR4/ NFkB/PFKFB3 signaling in the trophoblast to treat PE by reversing glucose metabolic reprogramming and NLRP3 inflammasomeinduced pyroptosis [132, 133]. Therefore, metformin and other NFkB signaling inhibitors may be potential treatments for PE.

Potential treatment options

In addition, targeting substances upstream of NFkB to block its mechanism of action has been shown in a large number of cell and animal studies to prevent or treat PE or improve the poor prognosis of PE. Such as USP14, Mammalian ste20-like kinase 1 (MST1) and proteinase-activated receptors-1 (PAR-1). USP14 expression levels are significantly upregulated in placental tissues of PE patients, which can activate NFkB. Through the development of drugs targeting USP14 may be helpful in the prevention of PE [134]. MST1, part of the tumor necrosis factor (TNF- α) receptor 1 signaling complex, reduces the effect of TNF- α on NF κ B signaling. Leonurine (LNR) is one of the active components of motherwort [135], which can exert anti-inflammatory and anti-apoptotic effects by upregulating MST1 and inhibiting NFkB signaling [136, 137]. MMP-1 can encourage the release of IL-8 from vascular smooth muscle, and it is markedly enhanced in the PE circulation. Recruitment of neutrophils might be aided by IL-8 [138]. Neutrophil expression of PAR-1 is specific for pregnancy. PAR-1 activates ROCK and then activates NFkB, causing vascular endothelial inflammatory injury [139]. PE may be prevented by blocking the

PAR-1 pathway or by reducing MMP-1 levels in the maternal circulation.

Traditional Chinese medicine treatment

In animals, cells and even clinical trials, some Chinese herbal preparations and their active ingredients have shown effective therapeutic benefits. Apocynin, Astragaloside IV (AsIV) and Fisetin could reduce PE-like symptoms such as hypertension and proteinuria, possibly by reducing inflammation through the TLR4/NF-B pathway [140, 141, 142]. In placental tissue from pre-eclamptic rats, astragalus injection can successfully inhibit the expression of sFlt-1, NFAT-5 and NFκB and increase the expression of PIGF and MMP-9 [110]. By controlling NLRP1 and NLRP3 inflammosomes in monocytes and activating the TLR4/NFκB pathway, silybin (SB) may be able to treat PE. However, its safety is questionable [143, 144].

Physiologic substances for PE

In addition to medication, the development of PE may also be regulated by physiological substances. One study found decreased vagus nerve activity in women complicated by PE [145]. ACh greatly reduced p38 MAPK and NFkB phosphorylation, as well as hypoxia-induced ROS production and subsequent apoptosis. According to Wang Zheng et al., ACh therapy can increase the activity of the vagus nerve and may be beneficial in the treatment of PE [146, 147]. The increase in estrogen and progesterone (PG) during pregnancy allows the uterus and placenta to improve vascularisation. PG regulates the expression of inflammation-related genes and proteins by inhibiting NFkB activation in macrophages, exerts an immunomodulatory effect on monocytes in pre-eclamptic women. Immunomodulation may be an alternative treatment for PE [148, 149]. Endogenous melatonin production is inhibited by hypoxia/reoxygenation (H/R), which reduces syncytiotrophoblast vitality. To prevent H/R-induced damage, exogenous melatonin therapy may be an option. This would increase placental cell survival and benefit fetal outcomes [150].

Drug carriers

Early pregnancy is the period of placental formation and also the susceptibility period for fetal teratogenesis. Poor placental formation is an irreversible process. Therefore, the treatment of PE is mainly focused on improving symptoms. Many NF κ B inhibitors cannot be used directly to prevent and cure PE because many small-molecule drugs may have unknown effects on the fetus across the placental barrier in early pregnancy. To stabilise the maternal circulation and avoid placental metastasis, Adrian C. Eddy and colleagues have developed a drug delivery method based on the bioengineered protein ELP (elastin-like polypeptide). The usefulness of this system for small molecule, peptide and protein therapies used during pregnancy is enhanced by the adaptability of ELP to fuse with various therapeutic agents [151].

Conclusion

NFkB affects the ability of trophoblasts to proliferate, invade and migrate by affecting the expression of MMPs, which may lead to inadequate remodelling of the spiral arteries and changes in placental villous morphology and function in early pregnancy. Note that the NFkB activation pathway in the placenta of women with PE in the third trimester differs from that in other cells and tissues. This difference may be related to the development of the P53/RSK1 complex [124] and may be due to the prolonged inflammatory state of the maternal placenta. However, the period of activation and transformation of the NFkB pathway is still unknown. It is interesting to note that cellular metabolic reprogramming is closely linked to TLR4/NFkB signaling. When the NFkB signaling pathway is activated, the energy metabolism within cells shifts from aerobic metabolism to glycolysis, which may also be related to the occurrence of PE [133].

Drugs targeting NFkB have shown therapeutic effects on PE in a large number of experiments, but considering their impact on the fetus, their use in the human body is still very cautious. In the future, it may be possible to reduce the impact of drugs on the fetus by constructing drug carriers, or to make drugs only act on the mother by affecting placental immunity and drug metabolism.

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Contributors' Statement

Yaxi Li prepared the manuscript. Jing Wang, Xiaolei Liang provided writing guidance. Qinying Zhu, Xue Qin, Yi Li revised the manuscript. Ruifen He, Junhong Du provided drawing guidance. All authors have approved the final manuscript.

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Conflict of Interest

The authors declare that they have no conflict of interest.

References

- Filipek A, Jurewicz E. Preeclampsia a disease of pregnant women. Postepy Biochem 2018; 64: 232–229. doi:10.18388/pb.2018_146
- [2] Wang Y, Li B, Zhao Y. Inflammation in Preeclampsia: Genetic Biomarkers, Mechanisms, and Therapeutic Strategies. Front Immunol 2022; 13: 883404. doi:10.3389/fimmu.2022.883404
- [3] Williams D. Long-term complications of preeclampsia. Semin Nephrol 2011; 31: 111–122. doi:10.1016/j.semnephrol.2010.10.010
- [4] Beksac MS, Tanacan A, Ozten G et al. Low-dose low-molecular-weight heparin prophylaxis against obstetrical complications in pregnancies with metabolic and immunological disorder-associated placental inflammation. J Matern Fetal Neonatal Med 2022; 35: 1546–1553. doi:10.1080/1 4767058.2020.1760834
- [5] Kupka E, Hesselman S, Hastie R et al. Low-dose aspirin use in pregnancy and the risk of preterm birth: a Swedish register-based cohort study. Am J Obstet Gynecol 2023; 228: 336.e1–336.e9. doi:10.1016/j.ajog.2022.0 9.006
- [6] Romagano MP, Sherman LS, Shadpoor B et al. Aspirin-Mediated Reset of Preeclamptic Placental Stem Cell Transcriptome – Implication for Stabilized Placental Function. Stem Cell Rev Rep 2022; 18: 3066–3082. doi:1 0.1007/s12015-022-10419-8
- [7] Tomimori-Gi K, Katsuragi S, Kodama Y et al. Low-dose aspirin therapy improves decidual arteriopathy in pregnant women with a history of preeclampsia. Virchows Arch 2022; 481: 713–720. doi:10.1007/s00428-0 22-03388-3
- [8] Amylidi-Mohr S, Kubias J, Neumann S et al. Reducing the Risk of Preterm Preeclampsia: Comparison of Two First Trimester Screening and Treatment Strategies in a Single Centre in Switzerland. Geburtshilfe Frauenheilkd 2021; 81: 1354–1361. doi:10.1055/a-1332-1437
- [9] Lu F, Gong H, Lei H et al. Downregulation of cathepsin C alleviates endothelial cell dysfunction by suppressing p38 MAPK/NF-κB pathway in preeclampsia. Bioengineered 2022; 13: 3019–3028. doi:10.1080/2165597 9.2021.2023994
- [10] Ding Y, Yang X, Han X et al. Ferroptosis-related gene expression in the pathogenesis of preeclampsia. Front Genet 2022; 13: 927869. doi:10.3 389/fgene.2022.927869
- [11] Tang R, Xiao G, Jian Y et al. The Gut Microbiota Dysbiosis in Preeclampsia Contributed to Trophoblast Cell Proliferation, Invasion, and Migration via IncRNA BC030099/NF-κB Pathway. Mediators Inflamm 2022; 2022: 6367264. doi:10.1155/2022/6367264
- [12] Lee S, Shin J, Kim JS et al. Targeting TBK1 Attenuates LPS-Induced NLRP3 Inflammasome Activation by Regulating of mTORC1 Pathways in Trophoblasts. Front Immunol 2021; 12: 743700. doi:10.3389/fimmu.2021.743 700
- [13] Logan MK, Lett KE, McLaurin DM et al. Coilin as a regulator of NF-kB mediated inflammation in preeclampsia. Biol Open 2022; 11: bio059326. doi:10.1242/bio.059326
- [14] Sakowicz A, Bralewska M, Pietrucha T et al. Canonical, Non-Canonical and Atypical Pathways of Nuclear Factor κb Activation in Preeclampsia. Int J Mol Sci 2020; 21: 5574. doi:10.3390/ijms21155574
- [15] Armistead B, Kadam L, Drewlo S et al. The Role of NFkappaB in Healthy and Preeclamptic Placenta: Trophoblasts in the Spotlight. Int J Mol Sci 2020; 21: 1775. doi:10.3390/ijms21051775
- [16] Oeckinghaus A, Ghosh S. The NF-kappaB family of transcription factors and its regulation. Cold Spring Harb Perspect Biol 2009; 1: a000034. doi:10.1101/cshperspect.a000034
- [17] Min C, Eddy SF, Sherr DH et al. NF-kappaB and epithelial to mesenchymal transition of cancer. J Cell Biochem 2008; 104: 733–744. doi:10.1002/j cb.21695

- [18] Thomsen LCV, Melton PE, Tollaksen K et al. Refined phenotyping identifies links between preeclampsia and related diseases in a Norwegian preeclampsia family cohort. J Hypertens 2015; 33: 2294–2302. doi:10.109 7/HJH.000000000000696
- [19] Johnson MP, Fitzpatrick E, Dyer TD et al. Identification of two novel quantitative trait loci for pre-eclampsia susceptibility on chromosomes 5q and 13q using a variance components-based linkage approach. Mol Hum Reprod 2007; 13: 61–67. doi:10.1093/molehr/gal095
- [20] Silva Carmona A, Mendieta Zerón H. NF-kB and SOD expression in preeclamptic placentas. Turk J Med Sci 2016; 46: 783–788. doi:10.3906/s ag-1503-75
- [21] Svensson J, Jenmalm MC, Matussek A et al. Macrophages at the Fetal-Maternal Interface Express Markers of Alternative Activation and Are Induced by M-CSF and IL-10. J Immunol 2011; 187: 3671–3682. doi:10.4 049/jimmunol.1100130
- [22] Choi H, Yang S-W, Joo J-S et al. Sialylated IVIg binding to DC-SIGN+ Hofbauer cells induces immune tolerance through the caveolin-1/NF-kB pathway and IL-10 secretion. Clin Immunol 2023; 246: 109215. doi:10.1 016/j.clim.2022.109215
- [23] Robertson SA, Care AS, Moldenhauer LM. Regulatory T cells in embryo implantation and the immune response to pregnancy. J Clin Invest 2018; 128: 4224–4235. doi:10.1172/JCI122182
- [24] Zhang X, Wei H. Role of Decidual Natural Killer Cells in Human Pregnancy and Related Pregnancy Complications. Front Immunol 2021; 12: 728291. doi:10.3389/fimmu.2021.728291
- [25] Gnainsky Y, Granot I, Aldo PB et al. Local injury of the endometrium induces an inflammatory response that promotes successful implantation. Fertil Steril 2010; 94: 2030–2036. doi:10.1016/j.fertnstert.2010.02.022
- [26] Kim C, Cathey AL, Watkins DJ et al. Adverse birth outcomes are associated with circulating matrix metalloproteinases among pregnant women in Puerto Rico. J Reprod Immunol 2023; 159: 103991. doi:10.1016/j.jri.2 023.103991
- [27] Jing M, Chen X, Qiu H et al. Insights into the immunomodulatory regulation of matrix metalloproteinase at the maternal-fetal interface during early pregnancy and pregnancy-related diseases. Front Immunol 2022; 13: 1067661. doi:10.3389/fimmu.2022.1067661
- [28] Mukherjee I, Dhar R, Singh S et al. Oxidative stress-induced impairment of trophoblast function causes preeclampsia through the unfolded protein response pathway. Sci Rep 2021; 11: 18415. doi:10.1038/s41598-0 21-97799-y
- [29] Sakowicz A. The role of NFκB in the three stages of pregnancy implantation, maintenance, and labour: a review article. BJOG 2018; 125: 1379–1387. doi:10.1111/1471-0528.15172
- [30] Singh N, Herbert B, Sooranna G et al. Is there an inflammatory stimulus to human term labour? PLoS One 2021; 16: e0256545. doi:10.1371/jou rnal.pone.0256545
- [31] Condon JC, Hardy DB, Kovaric K et al. Up-regulation of the progesterone receptor (PR)-C isoform in laboring myometrium by activation of nuclear factor-kappaB may contribute to the onset of labor through inhibition of PR function. Mol Endocrinol 2006; 20: 764–775. doi:10.1210/me.2005-0 242
- [32] Singh N, Herbert B, Sooranna GR et al. Is myometrial inflammation a cause or a consequence of term human labour? J Endocrinol 2017; 235: 69–83. doi:10.1530/JOE-17-0318
- [33] Li R, Xie J, Xu W et al. LPS-induced PTGS2 manipulates the inflammatory response through trophoblast invasion in preeclampsia via NF-kappaB pathway. Reprod Biol 2022; 22: 100696. doi:10.1016/j.repbio.2022.100 696
- [34] Somerset DA, Zheng Y, Kilby MD et al. Normal human pregnancy is associated with an elevation in the immune suppressive CD25+ CD4+ regulatory T-cell subset. Immunology 2004; 112: 38–43. doi:10.1111/j.1365-2 567.2004.01869.x

- [35] Ding H, Dai Y, Lei Y et al. Upregulation of CD81 in trophoblasts induces an imbalance of Treg/Th17 cells by promoting IL-6 expression in preeclampsia. Cell Mol Immunol 2019; 16: 302–312. doi:10.1038/s41423-0 18-0186-9
- [36] Ruan Q, Chen YH. Nuclear factor-κB in immunity and inflammation: the Treg and Th17 connection. Adv Exp Med Biol 2012; 946: 207–221. doi:1 0.1007/978-1-4614-0106-3_12
- [37] Miller D, Motomura K, Galaz J et al. Cellular immune responses in the pathophysiology of preeclampsia. J Leukoc Biol 2022; 111: 237–260. doi:10.1002/JLB.5RU1120-787RR
- [38] Gill N, Leng Y, Romero R et al. The immunophenotype of decidual macrophages in acute atherosis. Am J Reprod Immunol 2019; 81: e13098. doi:10.1111/aji.13098
- [39] King AE, Critchley HO, Kelly RW. The NF-kappaB pathway in human endometrium and first trimester decidua. Mol Hum Reprod 2001; 7: 175– 183. doi:10.1093/molehr/7.2.175
- [40] Rabalski AJ, Gyenis L, Litchfield DW. Molecular Pathways: Emergence of Protein Kinase CK2 (CSNK2) as a Potential Target to Inhibit Survival and DNA Damage Response and Repair Pathways in Cancer Cells. Clin Cancer Res 2016; 22: 2840–2847. doi:10.1158/1078-0432.CCR-15-1314
- [41] Huang WC, Hung MC. Beyond NF-κB activation: nuclear functions of IκB kinase α. J Biomed Sci 2013; 20: 3. doi:10.1186/1423-0127-20-3
- [42] Sakowicz A, Bralewska M, Pietrucha T et al. The Preeclamptic Environment Promotes the Activation of Transcription Factor Kappa B by P53/ RSK1 Complex in a HTR8/SVneo Trophoblastic Cell Line. Int J Mol Sci 2021; 22: 10200. doi:10.3390/ijms221910200
- [43] Vaughan JE, Walsh SW. Activation of NF-kB in placentas of women with preeclampsia. Hypertens Pregnancy 2012; 31: 243–251. doi:10.3109/1 0641955.2011.642436
- [44] Li G, Wei W, Suo L et al. Low-Dose Aspirin Prevents Kidney Damage in LPS-Induced Preeclampsia by Inhibiting the WNT5A and NF-κB Signaling Pathways. Front Endocrinol (Lausanne) 2021; 12: 639592. doi:10.3389/f endo.2021.639592
- [45] Litang Z, Hong W, Weimin Z et al. Serum NF-kBp65, TLR4 as Biomarker for Diagnosis of Preeclampsia. Open Med (Wars) 2017; 12: 399–402. doi:10.1515/med-2017-0057
- [46] Armistead B, Kadam L, Drewlo S et al. The Role of NFkB in Healthy and Preeclamptic Placenta: Trophoblasts in the Spotlight. Int J Mol Sci 2020; 21: 1775. doi:10.3390/ijms21051775
- [47] Mercnik MH, Schliefsteiner C, Fluhr H et al. Placental macrophages present distinct polarization pattern and effector functions depending on clinical onset of preeclampsia. Front Immunol 2022; 13: 1095879. doi:10.3389/fimmu.2022.1095879
- [48] Shaha S, Patel K, Riddell M. Cell polarity signaling in the regulation of syncytiotrophoblast homeostasis and inflammatory response. Placenta 2023; 141: 26–34. doi:10.1016/j.placenta.2022.11.007
- [49] Tanacan A, Beksac MS, Orgul G et al. Impact of extractable nuclear antigen, anti-double stranded DNA, antiphospholipid antibody, and anticardiolipin antibody positivity on obstetrical complications and pregnancy outcomes. Hum Antibodies 2019; 27: 135–141. doi:10.3233/hab-180359
- [50] Chappell S, Morgan L. Searching for genetic clues to the causes of preeclampsia. Clin Sci (Lond) 2006; 110: 443–458. doi:10.1042/CS2005032
 3
- [51] Parthiban PS, Mahendra J, Logaranjani A et al. Association between specific periodontal pathogens, Toll-like receptor-4, and nuclear factor-κB expression in placental tissues of pre-eclamptic women with periodontitis. J Investig Clin Dent 2018. doi:10.1111/jicd.12265
- [52] Le QA, Akhter R, Coulton KM et al. Periodontitis and Preeclampsia in Pregnancy: A Systematic Review and Meta-Analysis. Matern Child Health J 2022; 26: 2419–2443. doi:10.1007/s10995-022-03556-6

- [53] Konopka T, Zakrzewska A. Periodontitis and risk for preeclampsia a systematic review. Ginekol Pol 2020; 91: 158–164. doi:10.5603/GP.2020.0 024
- [54] León R, Silva N, Ovalle A et al. Detection of Porphyromonas gingivalis in the amniotic fluid in pregnant women with a diagnosis of threatened premature labor. J Periodontol 2007; 78: 1249–1255. doi:10.1902/jop.2 007.060368
- [55] Figuero E, Han YW, Furuichi Y. Periodontal diseases and adverse pregnancy outcomes: Mechanisms. Periodontol 2000 2020; 83: 175–188. doi:10.1111/prd.12295
- [56] Mahendra J, Parthiban PS, Mahendra L et al. Evidence Linking the Role of Placental Expressions of Peroxisome Proliferator-Activated Receptor-γ and Nuclear Factor-Kappa B in the Pathogenesis of Preeclampsia Associated With Periodontitis. J Periodontol 2016; 87: 962–970. doi:10.1902/j op.2016.150677
- [57] Hernández HG, Hernández-Castañeda AA, Pieschacón MP et al. ZNF718, HOXA4, and ZFP57 are differentially methylated in periodontitis in comparison with periodontal health: Epigenome-wide DNA methylation pilot study. J Periodontal Res 2021; 56: 710–725. doi:10.1111/jre.12868
- [58] Chopra A, Radhakrishnan R, Sharma M. Porphyromonas gingivalis and adverse pregnancy outcomes: a review on its intricate pathogenic mechanisms. Crit Rev Microbiol 2020; 46: 213–236. doi:10.1080/1040841X.2 020.1747392
- [59] Thomas C, Timofeeva I, Bouchoucha E et al. Oral and periodontal assessment at the first trimester of pregnancy: The PERISCOPE longitudinal study. Acta Obstet Gynecol Scand 2023; 102: 669–680. doi:10.1111/aog s.14529
- [60] Wen X, Fu X, Zhao C et al. The bidirectional relationship between periodontal disease and pregnancy *via* the interaction of oral microorganisms, hormone and immune response. Front Microbiol 2023; 14: 1070917. doi:10.3389/fmicb.2023.1070917
- [61] Chen X, Li P, Liu M et al. Gut dysbiosis induces the development of preeclampsia through bacterial translocation. Gut 2020; 69: 513–522. doi:1 0.1136/gutjnl-2019-319101
- [62] Chang Y, Chen Y, Zhou Q et al. Short-chain fatty acids accompanying changes in the gut microbiome contribute to the development of hypertension in patients with preeclampsia. Clin Sci (Lond) 2020; 134: 289– 302. doi:10.1042/CS20191253
- [63] Altemani F, Barrett HL, Gomez-Arango L et al. Pregnant women who develop preeclampsia have lower abundance of the butyrate-producer Coprococcus in their gut microbiota. Pregnancy Hypertens 2021; 23: 211–219. doi:10.1016/j.preghy.2021.01.002
- [64] Lv LJ, Li SH, Li SC et al. Early-Onset Preeclampsia Is Associated With Gut Microbial Alterations in Antepartum and Postpartum Women. Front Cell Infect Microbiol 2019; 9: 224. doi:10.3389/fcimb.2019.00224
- [65] Xue P, Zheng M, Gong P et al. Single administration of ultra-low-dose lipopolysaccharide in rat early pregnancy induces TLR4 activation in the placenta contributing to preeclampsia. PLoS One 2015; 10: e0124001. doi:10.1371/journal.pone.0124001
- [66] Tang R, Xiao G, Jian Y et al. The Gut Microbiota Dysbiosis in Preeclampsia Contributed to Trophoblast Cell Proliferation, Invasion, and Migration via IncRNA BC030099/NF-kappaB Pathway. Mediators Inflamm 2022; 2022: 6367264. doi:10.1155/2022/6367264
- [67] Apicella C, Ruano CSM, Thilaganathan B et al. Pan-Genomic Regulation of Gene Expression in Normal and Pathological Human Placentas. Cells 2023; 12: 578. doi:10.3390/cells12040578
- [68] Svensson J, Jenmalm MC, Matussek A et al. Macrophages at the fetal-maternal interface express markers of alternative activation and are induced by M-CSF and IL-10. J Immunol 2011; 187: 3671–3682. doi:10.4049/jim munol.1100130
- [69] Sakowicz A, Pietrucha T, Rybak-Krzyszkowska M et al. Double hit of NEMO gene in preeclampsia. PLoS One 2017; 12: e0180065. doi:10.137 1/journal.pone.0180065

- [70] Olarerin-George AO, Anton L, Hwang Y-C et al. A functional genomics screen for microRNA regulators of NF-kappaB signaling. BMC Biol 2013; 11: 19. doi:10.1186/1741-7007-11-19
- [71] Fan Y, Dong Z, Zhou G et al. Elevated miR-23a impairs trophoblast migration and invasiveness through HDAC2 inhibition and NF-kB activation. Life Sci 2020; 261: 118358. doi:10.1016/j.lfs.2020.118358
- [72] Zhou G, Li Z, Hu P et al. miR-219a suppresses human trophoblast cell invasion and proliferation by targeting vascular endothelial growth factor receptor 2 (VEGFR2). J Assist Reprod Genet 2021; 38: 461–470. doi:10.1 007/s10815-020-02022-y
- [73] Sheedy FJ, Palsson-McDermott E, Hennessy EJ et al. Negative regulation of TLR4 via targeting of the proinflammatory tumor suppressor PDCD4 by the microRNA miR-21. Nat Immunol 2010; 11: 141–147. doi:10.103 8/ni.1828
- [74] Dai Y, Diao Z, Sun H et al. MicroRNA-155 is involved in the remodelling of human-trophoblast-derived HTR-8/SVneo cells induced by lipopolysaccharides. Hum Reprod 2011; 26: 1882–1891. doi:10.1093/humrep/der 118
- [75] Kim S, Lee K-S, Choi S et al. NF-kB-responsive miRNA-31–5p elicits endothelial dysfunction associated with preeclampsia via down-regulation of endothelial nitric-oxide synthase. J Biol Chem 2018; 293: 18989–19000. doi:10.1074/jbc.RA118.005197
- [76] Yin A, Chen Q, Zhong M et al. MicroRNA-138 improves LPS-induced trophoblast dysfunction through targeting RELA and NF-κB signaling. Cell Cycle 2021; 20: 508–521. doi:10.1080/15384101.2021.1877927
- [77] Park M, Choi S, Kim S et al. NF-κB-responsive miR-155 induces functional impairment of vascular smooth muscle cells by downregulating soluble guanylyl cyclase. Exp Mol Med 2019; 51: 1–12. doi:10.1038/s12276-0 19-0212-8
- [78] Tay Y, Rinn J, Pandolfi PP. The multilayered complexity of ceRNA crosstalk and competition. Nature 2014; 505: 344–352. doi:10.1038/nature12986
- [79] Cipolla GA, de Oliveira JC, Salviano-Silva A et al. Long Non-Coding RNAs in Multifactorial Diseases: Another Layer of Complexity. Noncoding RNA 2018; 4: 13. doi:10.3390/ncrna4020013
- [80] Cheng D, Jiang S, Chen J et al. The Increased IncRNA MIR503HG in Preeclampsia Modulated Trophoblast Cell Proliferation, Invasion, and Migration via Regulating Matrix Metalloproteinases and NF-κB Signaling. Dis Markers 2019; 2019: 4976845. doi:10.1155/2019/4976845
- [81] Johnson JD, Louis JM. Does race or ethnicity play a role in the origin, pathophysiology, and outcomes of preeclampsia? An expert review of the literature. Am J Obstet Gynecol 2022; 226: S876–S885. doi:10.1016/ j.ajog.2020.07.038
- [82] Jiang L, Tang K, Magee LA et al. A global view of hypertensive disorders and diabetes mellitus during pregnancy. Nat Rev Endocrinol 2022; 18: 760–775. doi:10.1038/s41574-022-00734-y
- [83] Millar LK, Stollberg J, DeBuque L et al. Fetal membrane distention: determination of the intrauterine surface area and distention of the fetal membranes preterm and at term. Am J Obstet Gynecol 2000; 182: 128– 134. doi:10.1016/s0002-9378(00)70501-1
- [84] Padron JG, Norman Ing ND, Ng PoK et al. Stretch Causes Cell Stress and the Downregulation of Nrf2 in Primary Amnion Cells. Biomolecules 2022; 12: 766. doi:10.3390/biom12060766
- [85] Lim R, Barker G, Lappas M. The transcription factor Nrf2 is decreased after spontaneous term labour in human fetal membranes where it exerts anti-inflammatory properties. Placenta 2015; 36: 7–17. doi:10.1 016/j.placenta.2014.11.004
- [86] Gurbuz RH, Atilla P, Orgul G et al. Impaired Placentation and Early Pregnancy Loss in Patients with MTHFR Polymorphisms and Type-1 Diabetes Mellitus. Fetal Pediatr Pathol 2019; 38: 376–386. doi:10.1080/1551381 5.2019.1600623

- [87] Qin Y, Bily D, Aguirre M et al. Understanding PPARγ and Its Agonists on Trophoblast Differentiation and Invasion: Potential Therapeutic Targets for Gestational Diabetes Mellitus and Preeclampsia. Nutrients 2023; 15: 2459. doi:10.3390/nu15112459
- [88] Grimaldi B, Kohan-Ghadr H-R, Drewlo S. The Potential for Placental Activation of PPARy to Improve the Angiogenic Profile in Preeclampsia. Cells 2022; 11: 3514. doi:10.3390/cells11213514
- [89] Ju Y, Gu L, Hu M et al. Andrographolide exerts a neuroprotective effect by regulating the LRP1-mediated PPARγ/NF-κB pathway. Eur J Pharmacol 2023; 951: 175756. doi:10.1016/j.ejphar.2023.175756
- [90] Polvani S, Tarocchi M, Galli A. PPARγ and Oxidative Stress: Con(β) Catenating NRF2 and FOXO. PPAR Res 2012; 2012: 641087. doi:10.115 5/2012/641087
- [91] Psilopatis I, Vrettou K, Fleckenstein FN et al. The Role of Peroxisome Proliferator-Activated Receptors in Preeclampsia. Cells 2023; 12: 647. doi:1 0.3390/cells12040647
- [92] McCarthy FP, Drewlo S, Kingdom J et al. Peroxisome proliferator-activated receptor-γ as a potential therapeutic target in the treatment of preeclampsia. Hypertension 2011; 58: 280–286. doi:10.1161/HYPER-TENSIONAHA.111.172627
- [93] Kadam L, Kohan-Ghadr HR, Drewlo S. The balancing act PPAR-y's roles at the maternal-fetal interface. Syst Biol Reprod Med 2015; 61: 65–71. doi:10.3109/19396368.2014.991881
- [94] Nair AR, Silva SD, Agbor LN et al. Endothelial PPARγ (Peroxisome Proliferator-Activated Receptor-γ) Protects From Angiotensin II-Induced Endothelial Dysfunction in Adult Offspring Born From Pregnancies Complicated by Hypertension. Hypertension 2019; 74: 173–183. doi:10.1161/ HYPERTENSIONAHA.119.13101
- [95] Kadam L, Kilburn B, Baczyk D et al. Rosiglitazone blocks first trimester invitro placental injury caused by NF-kB-mediated inflammation. Sci Rep 2019; 9: 2018. doi:10.1038/s41598-018-38336-2
- [96] Wang J, Yang W, Xiao W et al. The association between smoking during pregnancy and hypertensive disorders of pregnancy: A systematic review and meta-analysis. Int J Gynaecol Obstet 2022; 157: 31–41. doi:10.1002/ ijgo.13709
- [97] Conde-Agudelo A, Althabe F, Belizán JM et al. Cigarette smoking during pregnancy and risk of preeclampsia: a systematic review. Am J Obstet Gynecol 1999; 181: 1026–1035. doi:10.1016/s0002-9378(99)70341-8
- [98] Xu H, Shi Q, Mo Y et al. Downregulation of α7 nicotinic acetylcholine receptors in peripheral blood monocytes is associated with enhanced inflammation in preeclampsia. BMC Pregnancy Childbirth 2019; 19: 188. doi:10.1186/s12884-019-2340-5
- [99] Liu Y, Yang J, Bao J et al. Activation of the cholinergic anti-inflammatory pathway by nicotine ameliorates lipopolysaccharide-induced preeclampsia-like symptoms in pregnant rats. Placenta 2017; 49: 23–32. doi:10.1 016/j.placenta.2016.11.003
- [100]Sharentuya N, Tomimatsu T, Mimura K et al. Nicotine suppresses interleukin-6 production from vascular endothelial cells: a possible therapeutic role of nicotine for preeclampsia. Reprod Sci 2010; 17: 556–563. doi:10.1177/1933719110362594
- [101]Lan R, Yang Y, Song J et al. Fas regulates the apoptosis and migration of trophoblast cells by targeting NF-κB. Exp Ther Med 2021; 22: 1055. doi:1 0.3892/etm.2021.10489
- [102] Tan W, Fu H, Zhou X et al. ANKRD37 inhibits trophoblast migration and invasion by regulating the NF-κB pathway in preeclampsia. J Gene Med 2022; 24: e3416. doi:10.1002/jgm.3416
- [103]Xie Y, Li X, Lv D et al. TREM-1 amplifies trophoblastic inflammation via activating NF-κB pathway during preeclampsia. Placenta 2021; 115. doi:10.1016/j.placenta.2021.09.016
- [104] Huang Z, Du G, Huang X et al. The enhancer RNA Inc-SLC4A1-1 epigenetically regulates unexplained recurrent pregnancy loss (URPL) by activating CXCL8 and NF-kB pathway. EBioMedicine 2018; 38: 162–170. doi:10.1016/j.ebiom.2018.11.015

- [105]Xue P, Fan W, Diao Z et al. Up-regulation of PTEN via LPS/AP-1/NF-κB pathway inhibits trophoblast invasion contributing to preeclampsia. Mol Immunol 2020; 118: 182–190. doi:10.1016/j.molimm.2019.12.018
- [106]Li R, Xie J, Xu W et al. LPS-induced PTGS2 manipulates the inflammatory response through trophoblast invasion in preeclampsia via NF-κB pathway. Reprod Biology 2022; 22: 100696. doi:10.1016/j.repbio.2022.100 696
- [107]Li S, Li A, Zhai L et al. Suppression of FPR2 expression inhibits inflammation in preeclampsia by improving the biological functions of trophoblast via NF-κB pathway. J Assist Reprod Genet 2022; 39: 239–250. doi:10.100 7/s10815-022-02395-2
- [108] Furmento VA, Marino J, Blank VC et al. The granulocyte colony-stimulating factor (G-CSF) upregulates metalloproteinase-2 and VEGF through PI3K/Akt and Erk1/2 activation in human trophoblast Swan 71 cells. Placenta 2014; 35: 937–946. doi:10.1016/j.placenta.2014.09.003
- [109]Cabral-Pacheco GA, Garza-Veloz I, Castruita-De la Rosa C et al. The Roles of Matrix Metalloproteinases and Their Inhibitors in Human Diseases. Int J Mol Sci 2020; 21: 9739. doi:10.3390/ijms21249739
- [110]Zhu Q, Wu X, Long Q et al. Mechanism of astragalus injection to relieve symptoms of preeclampsia rat model by inhibiting MMP-9/sFlt-1/TNF- α . Altern Ther Health Med 2023; 29: 125–131
- [111]Staun-Ram E, Goldman S, Gabarin D et al. Expression and importance of matrix metalloproteinase 2 and 9 (MMP-2 and -9) in human trophoblast invasion. Reprod Biol Endocrinol 2004; 2: 59. doi:10.1186/1477-7827-2-59
- [112]Bahabayi A, Yang N, Xu T et al. Expression of Matrix Metalloproteinase-2,-7,-9 in Serum during Pregnancy in Patients with Pre-Eclampsia: A Prospective Study. Int J Environ Res Public Health 2022; 19: 14500. doi:10.3 390/ijerph192114500
- [113] Wu L, Zhao K-Q, Wang W et al. Nuclear receptor coactivator 6 promotes HTR-8/SVneo cell invasion and migration by activating NF-κB-mediated MMP9 transcription. Cell Prolif 2020; 53: e12876. doi:10.1111/cpr.128 76
- [114]Oh SY, Hwang JR, Choi M et al. Autophagy regulates trophoblast invasion by targeting NF-κB activity. Sci Rep 2020; 10: 14033. doi:10.1038/s4159 8-020-70959-2
- [115]Liu J, Song G, Meng T et al. UL16-Binding Protein 1 Induced HTR-8/SVneo Autophagy via NF- κ B Suppression Mediated by TNF- α Secreted through uNK Cells. Biomed Res Int 2020; 2020: 9280372. doi:10.1155/2020/928 0372
- [116] Oh SY, Hwang JR, Choi M et al. Autophagy regulates trophoblast invasion by targeting NF- κB activity. Sci Rep 2020; 10: 14033. doi:10.1038/s4159 8-020-70959-2
- [117] Trocoli A, Djavaheri-Mergny M. The complex interplay between autophagy and NF-κB signaling pathways in cancer cells. Am J Cancer Res 2011; 1: 629–649
- [118]Copetti T, Demarchi F, Schneider C. p65/RelA binds and activates the beclin 1 promoter. Autophagy 2009; 5: 858–859. doi:10.4161/auto.882 2
- [119]Cheng C, Zhang J, Li X et al. NPRC deletion mitigated atherosclerosis by inhibiting oxidative stress, inflammation and apoptosis in ApoE knockout mice. Signal Transduct Target Ther 2023; 8: 290. doi:10.1038/s41392-0 23-01560-y
- [120] Chaabani R, Bejaoui M, Ben Jeddou I et al. Effect of the Non-steroidal Anti-inflammatory Drug Diclofenac on Ischemia-Reperfusion Injury in Rat Liver: A Nitric Oxide-Dependent Mechanism. Inflammation 2023; 46: 1221–1235. doi:10.1007/s10753-023-01802-9
- [121]Kim JH, Kim JY, Park M et al. NF-κB-dependent miR-31/155 biogenesis is essential for TNF-α-induced impairment of endothelial progenitor cell function. Exp Mol Med 2020; 52: 1298–1309. doi:10.1038/s12276-020-0478-x

- [122] Villalobos-Labra R, Liu R, Spaans F et al. Placenta-derived extracellular vesicles from preeclamptic and healthy pregnancies impair ex vivo vascular endothelial function. Biosci Rep 2022; 42: BSR20222185. doi:10.104 2/BSR20222185
- [123] Russell TM, Richardson DR. The good Samaritan glutathione-S-transferase P1: An evolving relationship in nitric oxide metabolism mediated by the direct interactions between multiple effector molecules. Redox Biol 2023; 59: 102568. doi:10.1016/j.redox.2022.102568
- [124]Sakowicz A. The Targeting of Nuclear Factor Kappa B by Drugs Adopted for the Prevention and Treatment of Preeclampsia. Int J Mol Sci 2022; 23: 2881. doi:10.3390/ijms23052881
- [125]Zuo Q, Zou Y, Huang S et al. Aspirin reduces sFlt-1-mediated apoptosis of trophoblast cells in preeclampsia. Mol Hum Reprod 2021; 27: gaaa089. doi:10.1093/molehr/gaaa089
- [126]Kim J, Lee KS, Kim J-H et al. Aspirin prevents TNF-α-induced endothelial cell dysfunction by regulating the NF-κB-dependent miR-155/eNOS pathway: Role of a miR-155/eNOS axis in preeclampsia. Free Radic Biol Med 2017; 104: 185–198. doi:10.1016/j.freeradbiomed.2017.01.010
- [127] Wu Y, Kang F, Yang Y et al. The protective effect of magnesium sulfate on placental inflammation via suppressing the NF-kB pathway in a preeclampsia-like rat model. Pregnancy Hypertens 2023; 31: 4–13. doi:10.1 016/j.preghy.2022.11.004
- [128]Shi CX, Qi QH, Xu J et al. Protective effect of magnesium sulfate on cranial nerves in preeclampsia rats through NF-κB/ICAM-1 pathway. Eur Rev Med Pharmacol Sci 2020; 24: 2785–2794. doi:10.26355/eurrev_20200 3_20639
- [129]Kovo M, Mevorach-Zussman N, Khatib N et al. The Effects of Magnesium Sulfate on the Inflammatory Response of Placentas Perfused With Lipopolysaccharide: Using the Ex Vivo Dual-Perfused Human Single-Cotyledon Model. Reprod Sci 2018; 25: 1224–1230. doi:10.1177/19337 19117737845
- [130]Ma Y, Yang Y, Lv M et al. 1,25(OH)2D3 alleviates LPS-induced preeclampsia-like rats impairment in the protective effect by TLR4/NF-kB pathway. Placenta 2022; 130: 34–41. doi:10.1016/j.placenta.2022.10.012
- [131]Brownfoot FC, Hannan NJ, Cannon P et al. Sulfasalazine reduces placental secretion of antiangiogenic factors, up-regulates the secretion of placental growth factor and rescues endothelial dysfunction. EBioMedicine 2019; 41: 636–648. doi:10.1016/j.ebiom.2019.02.013
- [132]Hu J, Zhang J, Zhu B. Protective effect of metformin on a rat model of lipopolysaccharide-induced preeclampsia. Fundam Clin Pharmacol 2019; 33: 649–658. doi:10.1111/fcp.12501
- [133]Zhang Y, Liu W, Zhong Y et al. Metformin Corrects Glucose Metabolism Reprogramming and NLRP3 Inflammasome-Induced Pyroptosis via Inhibiting the TLR4/NF-kB/PFKFB3 Signaling in Trophoblasts: Implication for a Potential Therapy of Preeclampsia. Oxid Med Cell Longev 2021; 2021: 1806344. doi:10.1155/2021/1806344
- [134]Zhao Y, Zong F. Inhibiting USP14 ameliorates inflammatory responses in trophoblast cells by suppressing MAPK/NF-kB signaling. Immun Inflamm Dis 2021; 9: 1016–1024. doi:10.1002/iid3.465
- [135]Zhu YZ, Wu W, Zhu Q et al. Discovery of Leonuri and therapeutical applications: From bench to bedside. Pharmacol Ther 2018; 188: 26–35. doi:10.1016/j.pharmthera.2018.01.006
- [136]Zong F, Zhao Y. Alkaloid leonurine exerts anti-inflammatory effects via modulating MST1 expression in trophoblast cells. Immun Inflamm Dis 2021; 9: 1439–1446. doi:10.1002/iid3.493
- [137] Hu PF, Sun FF, Qian J. Leonurine Exerts Anti-Catabolic and Anti-Apoptotic Effects via Nuclear Factor kappa B (NF-kB) and Mitogen-Activated Protein Kinase (MAPK) Signaling Pathways in Chondrocytes. Med Sci Monit 2019; 25: 6271–6280. doi:10.12659/msm.916039
- [138]Estrada-Gutierrez G, Cappello RE, Mishra N et al. Increased expression of matrix metalloproteinase-1 in systemic vessels of preeclamptic women: a critical mediator of vascular dysfunction. Am J Pathol 2011; 178: 451– 460. doi:10.1016/j.ajpath.2010.11.003

- [139] Walsh SW, Nugent WH, Al Dulaimi M et al. Proteases Activate Pregnancy Neutrophils by a Protease-Activated Receptor 1 Pathway: Epigenetic Implications for Preeclampsia. Reprod Sci 2020; 27: 2115–2127. doi:10.100 7/s43032-020-00232-4
- [140]Sha H, Ma Y, Tong Y et al. Apocynin inhibits placental TLR4/NF-κB signaling pathway and ameliorates preeclampsia-like symptoms in rats. Pregnancy Hypertens 2020; 22: 210–215. doi:10.1016/j.preghy.2020.10.006
- [141]Tuerxun D, Aierken R, Zhang YM et al. Astragaloside IV alleviates lipopolysaccharide-induced preeclampsia-like phenotypes via suppressing the inflammatory responses. Kaohsiung J Med Sci 2021; 37: 236–244. doi:10.1002/kjm2.12313
- [142]Li Y, Liu Y, Chen J et al. Protective effect of Fisetin on the lipopolysaccharide-induced preeclampsia-like rats. Hypertens Pregnancy 2022; 41: 23– 30. doi:10.1080/10641955.2021.2013874
- [143]Esmaeil N, Anaraki SB, Gharagozloo M et al. Silymarin impacts on immune system as an immunomodulator: One key for many locks. Int Immunopharmacol 2017; 50: 194–201. doi:10.1016/j.intimp.2017.06.030
- [144]Matias ML, Gomes VJ, Romao-Veiga M et al. Silibinin Downregulates the NF-κB Pathway and NLRP1/NLRP3 Inflammasomes in Monocytes from Pregnant Women with Preeclampsia. Molecules 2019; 24: 1548. doi:10.3 390/molecules24081548
- [145]Eneroth-Grimfors E, Westgren M, Ericson M et al. Autonomic cardiovascular control in normal and pre-eclamptic pregnancy. Acta Obstet Gynecol Scand 1994; 73: 680–684. doi:10.3109/00016349409029402

- [146] Wang Z, Zhao G, Zibrila AI et al. Acetylcholine ameliorated hypoxia-induced oxidative stress and apoptosis in trophoblast cells via p38 MAPK/ NF-kB pathway. Mol Hum Reprod 2021; 27: gaab045. doi:10.1093/mol ehr/gaab045
- [147] Wang Z, Zibrila AI, Liu S et al. Acetylcholine ameliorated TNF-α-induced primary trophoblast malfunction via muscarinic receptors[†]. Biol Reprod 2020; 103: 1238–1248. doi:10.1093/biolre/ioaa158
- [148] Matias ML, Romao-Veiga M, Ribeiro VR et al. Progesterone and vitamin D downregulate the activation of the NLRP1/NLRP3 inflammasomes and TLR4-MyD88-NF-κB pathway in monocytes from pregnant women with preeclampsia. J Reprod Immunol 2021; 144: 103286. doi:10.1016/j.jri.2 021.103286
- [149]Pepe GJ, Albrecht ED. Regulation of functional differentiation of the placental villous syncytiotrophoblast by estrogen during primate pregnancy. Steroids 1999; 64: 624–627. doi:10.1016/s0039-128x(99)00043-4
- [150]Sagrillo-Fagundes L, Assunção Salustiano EM, Ruano R et al. Melatonin modulates autophagy and inflammation protecting human placental trophoblast from hypoxia/reoxygenation. J Pineal Res 2018; 65: e12520. doi:10.1111/jpi.12520
- [151]Eddy AC, Howell JA, Chapman H et al. Biopolymer-Delivered, Maternally Sequestered NF-κB (Nuclear Factor-κB) Inhibitory Peptide for Treatment of Preeclampsia. Hypertension 2020; 75: 193–201. doi:10.1161/HYPER-TENSIONAHA.119.13368