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Endoscopic characteristics to differentiate sessile serrated lesion and microvesicular hyperplastic polyp from goblet cell-rich hyperplastic polyp


Affiliations below.

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Conflict of Interest: The authors declare that they have no conflict of interest.

Abstract:

Backgrounds
Among colorectal serrated polyps (SPs), sessile serrated lesions (SSLs) and hyperplastic polyps (HPs) show a similar endoscopic appearance. However, the endoscopic distinctions between the two categories, microvesicular HP (MVHP) and goblet cell-rich HP (GCHP) are not well understood. Therefore, we compared the endoscopic features of SSL, MVHP, and GCHP.

Methods
This retrospective, cross-sectional study was conducted at the Toyoshima Endoscopy Clinic. We examined the polyp size, location, Paris classification type, mucus cap, indistinct border, expanded crypt opening, varicose microvascular vessels, and JNET classification type. Multivariable analysis of each endoscopic finding using a binomial logistic regression model determined the factors that predicted SP histology.

Results
A total of 670 SPs were enrolled in this study, comprising 159 SSLs, 361 MVHPs, and 150 GCHPs. On comparing the SSL + MVHP group and GCHP, a mucus cap (partial regression coefficient 1.705), expanded crypt opening (1.828), and varicose microvascular vessels (1.270) were more often observed in SSL + MVHP group compared with GCHP. In the comparison between MVHP and GCHP, a mucus cap (1.564), expanded crypt opening (1.802), and varicose microvascular vessels (1.288) were more often found in MVHP in contrast to GCHP. When comparing SSL and MVHP, SSLs were more likely to be in the proximal colon (0.662) and were larger (0.198) than MVHPs. No significant differences were observed in other endoscopic findings.

Conclusions
SSL and MVHP have endoscopic appearances that differ from those of GCHP. Considering MVHP and GCHP as distinct entities may aid in the endoscopic diagnosis of SPs.

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INTRODUCTION

Globally, colorectal cancer (CRC) is the most common type of gastrointestinal cancer. Conventional adenomas and sessile serrated lesions (SSLs) are precursors of CRC [1,2]. The risk of CRC can be reduced by removing conventional adenomas and SSLs; therefore, the detection and diagnosis of these precursor lesions during colonoscopy are crucial [3-5]. New CRCs evolving via serrated polyps (SPs), namely the serrated pathway, account for 25%–30% [6,7]. Colorectal SPs are histologically characterized by a serrated epithelial architecture. Currently there is an understanding of the different types of colorectal SPs and their biological characteristics, including SSL, hyperplastic polyps (HPs), and traditional serrated adenomas (TSAs) [8].

An overall distortion of the normal crypt architecture is the characteristic histological feature of SSL. Crypt architectural changes observed in SSLs are as follows: 1) crypts grow along the muscularis mucosa, 2) the crypt base dilates in contrast to superficial serrations in HP, 3) crypts asymmetrically proliferate, and 4) they contain a mixture of goblet cells with microvesicular mucin droplets. HPs are identified by exclusion if the architectural criteria for the SSL are not met. The overall architecture of the HP is unchanged compared to that of the normal colonic mucosa, and the crypts remained evenly spaced. HP has two histological variants, microvesicular HP (MVHP) and goblet cell-rich HP (GCHP), based on crypt architecture and mucin type. MVHP has funnel-shaped crypts with serrations limited to the upper two-thirds, with mucin type of mixed microvesicular and goblet cells. GCHP is characterized by elongated crypts that resemble enlarged normal crypts with little to no serration and goblet cell-predominant mucin (Figure 1) [9,10].

The serrated pathway includes a sequence of genetic and epigenetic alterations that lead to the development of sporadic CRCs. Activating mutations in BRAF in MVHP and SSL, or KRAS in GCHP are thought to initiate the development of SPs and are mutually exclusive[6,8,11-13].

Among SPs, SSLs and HPs show a similar appearance on endoscopy, such as a flat appearance and a similar color to that of healthy colonic mucosa [14-16]. A previous study comparing the endoscopic appearance of SSLs and HPs (including both MVHPs and GCHPs) found that SSLs were larger and more
frequently showed a mucus cap and an indistinct border than HPs [17]. However, the differences in endoscopic appearance between the two categories, MVHP and GCHP, are poorly understood. Therefore, we divided the HPs into two distinct categories (MVHP and GCHP) and compared the endoscopic features of the SSL, MVHP, and GCHP.

METHODS

Study overview

This retrospective cross-sectional study was conducted at the Toyoshima Endoscopy Clinic, a specialized outpatient endoscopy clinic located in an urban area of Japan. The patients were enrolled between September 2022 and February 2023. When patients had multiple SPs, they were treated individually. Indications for screening patients included, evaluation of symptoms, investigation for abnormal laboratory findings including fecal immunochemical tests, and surveillance [18].

Ethics

This study was conducted in accordance with ethical guidelines for medical studies in Japan and received approval from the ethics committee of the Certified Institutional Review Board of Yoyogi Mental Clinic (certificate number: RKK227). We have published the study protocol on our clinic’s website (https://www.ichou.com/?p=7125), allowing patients to opt out of the study if they so desired. Participants provided written consent to participate in the study prior to undergoing endoscopy. This study complied with the guidelines of the Declaration of Helsinki.

Colonoscopy

Three expert endoscopists (T.N., S.Y., and O.T.) performed the colonoscopies. Toyoshima Endoscopy Clinic has incorporated an EVIS X1 video system center (CV-1500; Olympus Co., Tokyo, Japan) featuring a 4 K resolution ultrahigh-definition liquid crystal display monitor (OEV321UH; Olympus Co., Tokyo, Japan). The clinic utilized colonoscopes (PCF-H290Z, CF-HQ290Z, or CF-XZ1200; Olympus Co., Tokyo, Japan) for the procedures. The T-File System (STS-Medic Inc., Tokyo, Japan) was used for
managing endoscopic reports and images. Endoscopic sedation was administered using pethidine, midazolam, and/or propofol with a targeted depth of sedation set at “moderate sedation” (or conscious sedation). Pancolonic chromoendoscopy utilizing 0.05% indigo carmine was routinely conducted. The observation modes included white-light imaging and/or texture and color enhancement imaging (TXI) [19-21]. All detected SPs were gently washed with water and observed using narrow-band imaging (NBI) with magnification.

Clinically significant SPs (CSSPs) were defined as all SSLs, TSAs, HPs ≥ 10 mm, and HPs > 5 mm in the proximal colon [2]. In this study, the endoscopists removed the polyps suspected to be CSSPs[18]. The HP was eligible for resection if it was > 5 mm in the proximal colon or ≥ 10mm in the distal colorectum. When the polyp was suspected to be an SSL owing to its mucus cap, indistinct border, expanded crypt opening, and/or varicose microvascular vessels, it was resected even if it was ≤ 5 mm in the proximal colon or < 10 mm in the distal colorectum. Diminutive (i.e., ≤ 5 mm) polyps in the distal colorectum, which were predicted with high confidence to be HPs, were not resected. TSAs were not included in this study because the endoscopic appearance of TSAs is distinct from that of other SPs [6].

**Colorectal polyp**

Endoscopic reports included the location, size, shape, and endoscopic appearance of each SP. The proximal colon was delineated as extending from the cecum to the descending colon. Polyp size was measured by comparing it to the thickness or width of a snare or forceps. The endoscopic morphology was assessed according to the Paris classification [22], indistinct borders [17], presence of a mucus cap [23], expanded crypt opening, varicose microvascular vessels [24,25], and Japan NBI expert team (JNET) classification [26]. The mucus cap was defined as rich mucus covering, while indistinct borders referred to vague demarcations of the lesion border [23]. An expanded crypt opening, also referred to as a dark spot within the crypt or corresponding to Kudo pit pattern type II-Open, was defined as the heterogeneous expansion of nearby crypts [27]. Varicose microvascular vessels were defined as those thicker than meshed capillary vessels and meandering as varicose veins, which differed from the capillary pattern of the mucosal vascular
network. The lengths of the varicose microvascular vessels varied, and their location was inconsistent on the lesion surface [24]. Representative images are shown in Figure 1.

An expert gastrointestinal pathologist (Emeritus Professor Hidenobu Watanabe) diagnosed SPs using hematoxylin and eosin staining. Data were extracted from the Toyoshima Clinic Endoscopy Database.

**Statistical analysis**

We divided the SPs into two groups: a microvesicular mucin group and goblet cell-rich mucin group (*i.e.*, SSL + MVHP versus GCHP). We then assessed the differences in endoscopic appearance between the two groups. Next, a subgroup analysis was conducted for HPs (*i.e.*, MVHP versus GCHP). Finally, we performed a subgroup analysis of the microvesicular mucin group (*i.e.*, SSL versus MVHP).

We examined the means and standard deviations for each continuous variable (patient age and polyp size) and frequencies for each categorical variable (patient sex, proximal polyp location, Paris classification type 0-II, mucus cap, indistinct border, expanded crypt opening, varicose microvascular vessels, and JNET classification type 1).

We performed univariable and multivariable analyses of each endoscopic finding using a binomial logistic regression model to determine the factors that predicted the histology of SP, reported as partial regression coefficients. The multivariable analysis was restricted to observations with no missing data. The effects model consisted of variables that were statistically significant in univariable analysis. Two-tailed *P* values <0.05 were considered statistically significant.

We assigned 1 point to the significant variables in the multivariable analysis and developed the sum of these points as the endoscopic SSL/MVHP score. The receiver operating characteristic (ROC) curve was constructed to predict SSL and MVHP other than GCHP in SPs, and the area under the curve (AUC), sensitivity, specificity, and positive predictive value (PPV) of the endoscopic SSL/MVHP score were measured. The optimal cut-off value of the ROC curve was determined using the Youden index. Calculations were performed using BellCurve for Excel version 4.05 (Social Survey Research Information Co., Ltd., Tokyo, Japan).
RESULTS

A total of 670 lesions comprising 159 SSLs, 361 MVHPs, and 150 GCHPs were included in this study. The characteristics of the SPs are listed in Table 1. Mean age was 59.0 years, and 44.8% of the patients were men. SPs located in the proximal colon accounted for 70.3%. The majority of SPs showed Paris 0-IIa morphology and JNET classification 1. The mean polyp size was 6.42 mm. The frequencies of mucus cap, indistinct border, expanded crypt opening, and varicose microvascular vessels were 51.0%, 48.5%, 44.8%, and 27.0%, respectively.

Comparison between SSL + MVHP and GCHP groups

Table 2 shows the effect of endoscopic appearance on the histological diagnosis of SP in univariable and multivariable analyses. The mucus cap (partial regression coefficient 1.705, 95% confidence interval [CI] 1.141–2.269), expanded crypt opening (1.828, 1.159–2.496), and varicose microvascular vessels (1.270, 0.590–1.949) were more frequently observed in the SSL+MVHP group than in the GCHP group. The mucus cap, expanded crypt opening, and varicose microvascular vessels were each assigned 1 point, and the sum of the points was defined as the endoscopic SSL/MVHP score. The ROC curve for the endoscopic SSL/MVHP score is shown in Figure 2A. Of the endoscopic SP score, the AUC was 0.83 (95% CI 0.81–0.86) and the optimal cut-off value was 1; the sensitivity, specificity, and PPV were 81.5%, 74.7%, 91.8%, respectively.

Comparison between MVHP and GCHP

Univariable and multivariable analyses of the differences in endoscopic appearance between MVHP and GCHP are shown in Table 3. Similar to the above analysis, the mucus cap (partial regression coefficient 1.564, 95% CI 0.988–2.139), expanded crypt opening (1.802, 1.127–2.477), and varicose microvascular vessels (1.288, 0.596–1.980) were more often found in MVHP than in GCHP. The AUC of the endoscopic SSL/MVHP score was 0.80 (95% CI 0.76–0.83) and the optimal cut-off value was 1 (Figure 2B). The sensitivity, specificity, and PPV were 76.7%, 74.7%, and 87.9%, respectively.
Comparison between SSL and MVHP

Table 4 presents the results of univariable and multivariable analyses comparing the endoscopic findings for SSL and MVHP. SSLs were more likely to be located in the proximal colon (partial regression coefficient 0.662, 95% CI 0.087–1.237) and larger (0.198, 0.134–0.262) than MVHPs. No significant differences were observed in the other endoscopic findings.

DISCUSSION

We found that the SSL and MVHP groups exhibited distinct endoscopic appearances compared to the GCHP group. Furthermore, a sub-analysis of HPs revealed that MVHP and GCHP presented with different endoscopic findings. These differences were consistent when comparing the combined group of SSL + MVHP group with the GCHP group. Adhesion of the mucus, expanded crypt opening, and varicose microvascular vessels were independently and more frequently observed in SSL and MVHP than in GCHP. Furthermore, a comparison between SSL and MVHP showed that SSL were more prevalent in the proximal colon and had a larger diameter. However, there were no significant differences in other endoscopic features. These distinctions in endoscopic appearance may be attributed to histological and molecular similarities between SSL and MVHP, unlike in GCHP. Histologically, the mucin types in SSL and MVHP are mixed microvesicular and goblet cells, whereas GCHP is goblet cell-predominant. This property of mucus may contribute to endoscopic mucus adhesion [28]. Similar to MVHP, SSL is characterized by bland cytology and crypts with prominent serrations. Although an SSL is identified when at least one crypt shows unequivocal distortion according to the updated WHO criteria, the majority of SSL crypts lack an abnormal architecture, and most crypts resemble those seen in MVHP [6,8]. Regarding molecular features, more than 90% of SSLs and 70–80% of MVHPs have BRAF mutations, whereas SSLs and MVHPs do not have KRAS mutations. In contrast, > 90% of GCHPs have KRAS mutations, but no GCHPs have BRAF mutations. Given that SSL and MVHP are recognized as the BRAF serrated pathway, GCHP is thought to be the KRAS serrated pathway and is mutually exclusive [8,13]. The similarities between SSL and MVHP may be reflected in their endoscopic appearances [13]. In the endoscopic diagnosis of SPs, including the use of
artificial intelligence, a more accurate diagnosis could be made if SPs were evaluated based on three categories: SSL, MVHP, and GCHP, instead of classifying them into two categories: SSL and HP.

In contrast to HPs, SSLs are frequently covered with a mucus cap [6]. An expanded crypt opening is believed to correspond to crypt dilation, which is an important histological feature of SSLs [23]. Varicose microvascular vessels are defined as thickened vessels that differ from the capillary pattern of the mucosal vascular network and are inconsistently located on the lesion surface [24]. Thus, mucus cap, expanded crypt opening, and varicose microvascular vessels have been reported as specific findings of SSL; however, this study demonstrated that they are also associated with MVHP. Recent advances in endoscopy (e.g., improved image quality and image enhancement with TXI) may lead to the identification of these findings in MVHPs (Figure 1) [20,29].

In this study, the differences between SSL and MVHP were their localization and size. Pai et al. reported that the majority (75–90%) of HPs are found in the distal colon and rectum, whereas SSLs have a predilection for the proximal colon (70–80%). SSLs are characterized by a larger size and distal HPs are usually small (< 5 mm). Furthermore, SSL is a known precursor of CRC, although HP, particularly proximal MVHP, is a probable precursor of SSL [6,8,17]. These results are in agreement with those of the present study. Collectively, among the SPs, the mucus cap, expanded crypt opening, and varicose microvascular vessels are predictors of not only SSL but also MVHP. Among the SPs with these endoscopic appearances, large polyps in the proximal colon are more likely to be SSLs. Endoscopic appearance, size, and location could allow the prediction of the type of SP [8].

The endoscopist’s level of confidence in the optical diagnosis of a colorectal lesion is an important factor in its application to clinical practice. The majority of lesions have typical endoscopic features that enable a high confidence prediction of histology [30]. However, our findings suggest that optical diagnosis performance may be decreased in the differentiation of SSLs from MVHPs. The confidence level may increase if the SPs are categorized as the SSL+MVHP group versus the GCHP group instead of SSLs versus HPs. At least, the individual endoscopic diagnosis of MVHP and GCHP is warranted. Currently, computer-aided diagnosis (CADx) using artificial intelligence (AI) is progressing in endoscopic diagnosis [31]. Learning HPs as separate MVHPs or GCHPs would enable the differential diagnosis of MVHPs and
GCHPs. Our findings are particularly promising for the development of future AI CADx modules and may substantially contribute to the field.

Pathologically, making the differential diagnosis of SPs is also challenging. Singh et al. [32] reported that nearly one-fifth of previously diagnosed HPs in the proximal colon and those > 5 mm in size were histologically reclassified as SSLs on reassessment by other pathologists. Since MVHPs in particular bear histological similarities to SSLs [9,10], pathologists should carefully differentiate proximal MVHPs and MVHPs > 5 mm, from SSLs. When pathologists arbitrarily diagnose MVHP and GCHP separately, the chance of misdiagnosing SSLs as HPs might be reduced.

The varicose microvascular vessels have two entities, as follows: the dilated and branching vessels (DBVs) reported by Yamada et al. [33] and the thick and branching vessels (TBVs) reported by Yamashina et al.[34]. DBVs are defined as thickened capillary vessels with branching on the surface and differ from the “meshed capillary vessels” in the Sano et al. capillary pattern classification [35]. DBVs are usually dark brown, which indicates that the vessels are in the superficial layers. Yamashina et al. described that TBV was dark green and much thicker than meshed capillary vessels. The dark green color in NBI indicates that the vessels are in the deeper layers. The sensitivities of varicose microvascular vessels, DBVs, and TBVs for SSL are reportedly 57.9%, 65%, and 45.1%, respectively. Their specificities for SSL were 87.8%, 76%, and 68.9%, respectively. Our study used the diagnostic criteria for varicose microvascular vessels (Figures 1C and 1G (red arrows) represent TBVs and DBVs, respectively). Varicose microvascular vessels should be assessed individually for DBVs and TBVs, and that is a future agenda.

The present study has some limitations. Firstly, this had a retrospective design and was limited to a single center with expert endoscopists. However, the data source was well-controlled. Future prospective investigations involving multiple centers, including non-expert practitioners are warranted. Secondly, we used NBI and TXI as image-enhancing modalities (Olympus Corporation). To further validate our results it is necessary to verify these findings using other image enhancement methods, such as blue LASER imaging (BLI) and linked color imaging (LCI, Fujifilm Corporation) is required [36]. Thirdly, mutations in BRAF and KRAS may contribute to differences in endoscopic appearance. However, this study did not explore the association between molecular profiles and endoscopic features. Fourthly, the present study was conducted
in routine clinical practice, and polyp resection was limited to those suspected to be CSSPs [2]. Therefore, certain small HPs in the distal colorectum were excluded from this study. Conversely, all endoscopically diagnosed SSLs were removed, including the small ones in the distal colorectum. If all HPs were also resected, it may have highlighted the larger and more frequent occurrence of SSLs in the proximal colon.

In conclusions, SSL and MVHP had distinct endoscopic appearances including mucus cap, expanded crypt opening, and varicose microvascular vessels, compared to GCHP. There were no differences in endoscopic findings between SSL and MVHP, other than their location and size. Thus, interestingly, MVHP and GCHP, while belonging to the same HP category, displayed different endoscopic appearances. Conversely, SSL and MVHP despite belonging to different histopathological categories, demonstrated striking endoscopic similarities. This study results build upon our current understanding of SPs, particularly for distinguishing between JNET type I lesions. It offers the differentiating variables of SSL, MVHP, and GCHP, advancing the endoscopist’s ability to distinguish among them.

Diagnosing HP as an individual category (i.e., MVHP or GCHP), rather than diagnosing it inclusively, is vital in clinical practice for both endoscopists and pathologists. Although MVHPs are categorized into HPs according to WHO classification [7], they may be precursors to SSLs and share a very similar endoscopic appearance. Differentiating MVHPs from GCHPs would influence pathological awareness, endoscopic therapeutic strategies, and surveillance interval recommendations. More evidence from longitudinal studies is needed to determine appropriate therapeutic strategies for SPs.

REFERENCES


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**Tables**

**Table 1. Characteristics of serrated polyps.**

<table>
<thead>
<tr>
<th></th>
<th>Total</th>
<th>SSL</th>
<th>MVHP</th>
<th>GCHP</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>N</strong></td>
<td>670</td>
<td>159</td>
<td>361</td>
<td>150</td>
</tr>
<tr>
<td><strong>Age, mean+-SD, years</strong></td>
<td>59.0+-11.3</td>
<td>57.9+-10.8</td>
<td>58.6+-11.2</td>
<td>61.1+-11.8</td>
</tr>
<tr>
<td><strong>Male sex, %</strong></td>
<td>44.8</td>
<td>34.6</td>
<td>47.9</td>
<td>48.0</td>
</tr>
<tr>
<td><strong>Proximal colon, %</strong></td>
<td>70.3</td>
<td>86.8</td>
<td>63.4</td>
<td>69.3</td>
</tr>
<tr>
<td><strong>Paris classification, type 0-II, %</strong></td>
<td>98.4</td>
<td>97.5</td>
<td>98.3</td>
<td>99.3</td>
</tr>
<tr>
<td><strong>Size, mean+-SD, mm</strong></td>
<td>6.42+-3.97</td>
<td>9.65+-4.96</td>
<td>5.80+-3.15</td>
<td>4.45+-2.18</td>
</tr>
<tr>
<td><strong>Mucus cap, %</strong></td>
<td>51.0</td>
<td>81.1</td>
<td>53.7</td>
<td>12.7</td>
</tr>
<tr>
<td><strong>Indistinct border, %</strong></td>
<td>48.5</td>
<td>76.7</td>
<td>49.3</td>
<td>16.7</td>
</tr>
<tr>
<td><strong>Expanded crypt opening, %</strong></td>
<td>44.8</td>
<td>69.2</td>
<td>49.3</td>
<td>8.0</td>
</tr>
<tr>
<td><strong>Varicose microvascular vessels, %</strong></td>
<td>27.0</td>
<td>45.3</td>
<td>26.9</td>
<td>8.0</td>
</tr>
<tr>
<td><strong>JNET classification, type 1, %</strong></td>
<td>93.4</td>
<td>95.6</td>
<td>93.6</td>
<td>90.7</td>
</tr>
</tbody>
</table>

SSL, sessile serrated lesion; MVHP, microvesicular hyperplastic polyp; GCHP, goblet cell-rich hyperplastic polyp; SD, standard deviation; JNET, Japan narrow band imaging expert team.
Table 2. Comparison of endoscopic appearance between SSL + MVHP and GCHP groups.

<table>
<thead>
<tr>
<th></th>
<th>Univariable analysis</th>
<th></th>
<th></th>
<th>Multivariable analysis</th>
<th></th>
<th></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Partial regression coefficient</td>
<td>95% CI</td>
<td>P value</td>
<td>Partial regression coefficient</td>
<td>95% CI</td>
<td>Degree of freedom</td>
<td>P value</td>
</tr>
<tr>
<td>Proximal location</td>
<td>0.059</td>
<td>-0.336, 0.454</td>
<td>0.77</td>
<td></td>
<td></td>
<td>1</td>
<td></td>
</tr>
<tr>
<td>Polyp size</td>
<td>0.273</td>
<td>0.194, 0.353</td>
<td>&lt; 0.001</td>
<td>0.074</td>
<td>-0.014, 0.162</td>
<td>1</td>
<td>0.10</td>
</tr>
<tr>
<td>Paris classification, type 0-II</td>
<td>-1.072</td>
<td>-3.136, 0.992</td>
<td>0.31</td>
<td></td>
<td></td>
<td>1</td>
<td></td>
</tr>
<tr>
<td>Mucus cap</td>
<td>2.425</td>
<td>1.912, 2.938</td>
<td>&lt; 0.001</td>
<td>1.705</td>
<td>1.141, 2.269</td>
<td>1</td>
<td>&lt; 0.001</td>
</tr>
<tr>
<td>Indistinct border</td>
<td>1.920</td>
<td>1.456, 2.383</td>
<td>&lt; 0.001</td>
<td>0.395</td>
<td>-0.182, 0.971</td>
<td>1</td>
<td>0.18</td>
</tr>
<tr>
<td>Expanded crypt opening</td>
<td>2.659</td>
<td>2.044, 3.273</td>
<td>&lt; 0.001</td>
<td>1.828</td>
<td>1.159, 2.496</td>
<td>1</td>
<td>&lt; 0.001</td>
</tr>
<tr>
<td>Varicose microvascular vessels</td>
<td>1.711</td>
<td>1.094, 2.329</td>
<td>&lt; 0.001</td>
<td>1.270</td>
<td>0.590, 1.9849</td>
<td>1</td>
<td>&lt; 0.001</td>
</tr>
<tr>
<td>JNET classification, type 1</td>
<td>0.520</td>
<td>-0.143, 1.182</td>
<td>0.12</td>
<td></td>
<td></td>
<td>1</td>
<td></td>
</tr>
</tbody>
</table>

P values were calculated using binomial logistic regression analysis.

SSL, sessile serrated lesion; MVHP, microvesicular hyperplastic polyp; GCHP, goblet cell-rich hyperplastic polyp; CI, confidence interval; JNET, Japan narrow band imaging expert team.
Table 3. Comparison of endoscopic appearance between MVHP and GCHP.

<table>
<thead>
<tr>
<th></th>
<th>Univariable analysis</th>
<th></th>
<th>Multivariable analysis</th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Partial regression coefficient</td>
<td>95% CI</td>
<td>P value</td>
<td>Partial regression coefficient</td>
</tr>
<tr>
<td>Proximal location</td>
<td>-0.265</td>
<td>-0.673, 0.143</td>
<td>0.20</td>
<td></td>
</tr>
<tr>
<td>Polyp size</td>
<td>0.197</td>
<td>0.113, 0.280</td>
<td>&lt; 0.001</td>
<td>0.014</td>
</tr>
<tr>
<td>Paris classification, type 0-II</td>
<td>-0.924</td>
<td>-3.049, 1.202</td>
<td>0.39</td>
<td></td>
</tr>
<tr>
<td>Mucus cap</td>
<td>2.081</td>
<td>1.557, 2.604</td>
<td>&lt; 0.001</td>
<td>1.564</td>
</tr>
<tr>
<td>Indistinct border</td>
<td>1.582</td>
<td>1.105, 2.058</td>
<td>&lt; 0.001</td>
<td>0.434</td>
</tr>
<tr>
<td>Expanded crypt opening</td>
<td>2.415</td>
<td>1.790, 3.040</td>
<td>&lt; 0.001</td>
<td>1.802</td>
</tr>
<tr>
<td>Varicose microvascular vessels</td>
<td>1.441</td>
<td>0.807, 2.075</td>
<td>&lt; 0.001</td>
<td>1.288</td>
</tr>
<tr>
<td>JNET classification, type 1</td>
<td>0.414</td>
<td>-0.280, 1.108</td>
<td>0.24</td>
<td></td>
</tr>
</tbody>
</table>

P values were calculated using binomial logistic regression analysis.

MVHP, microvesicular hyperplastic polyp; GCHP, goblet cell-rich hyperplastic polyp; CI, confidence interval; JNET, Japan narrow band imaging expert team.
Table 4. Comparison of endoscopic appearance between SSL and MVHP.

<table>
<thead>
<tr>
<th></th>
<th>Partial regression coefficient</th>
<th>95% CI</th>
<th>P value</th>
<th>Partial regression coefficient</th>
<th>95% CI</th>
<th>Degree of freedom</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Proximal location</td>
<td>1.332</td>
<td>0.825, 1.838</td>
<td>&lt; 0.001</td>
<td>0.6620</td>
<td>0.087, 1.237</td>
<td>1</td>
<td>0.024</td>
</tr>
<tr>
<td>Polyp size</td>
<td>0.250</td>
<td>0.191, 0.308</td>
<td>&lt; 0.001</td>
<td>0.198</td>
<td>0.134, 0.262</td>
<td>1</td>
<td>&lt; 0.001</td>
</tr>
<tr>
<td>Paris classification, type 0-II</td>
<td>-0.423</td>
<td>-1.702, 0.856</td>
<td>0.52</td>
<td>0.521</td>
<td>-0.008, 1.049</td>
<td>1</td>
<td>0.054</td>
</tr>
<tr>
<td>Mucus cap</td>
<td>1.309</td>
<td>0.861, 1.757</td>
<td>&lt; 0.001</td>
<td>0.521</td>
<td>-0.008, 1.049</td>
<td>1</td>
<td>0.054</td>
</tr>
<tr>
<td>Indistinct border</td>
<td>1.221</td>
<td>0.799, 1.643</td>
<td>&lt; 0.001</td>
<td>0.113</td>
<td>-0.429, 0.655</td>
<td>1</td>
<td>0.68</td>
</tr>
<tr>
<td>Expanded crypt opening</td>
<td>0.836</td>
<td>0.442, 1.231</td>
<td>&lt; 0.001</td>
<td>0.132</td>
<td>-0.352, 0.617</td>
<td>1</td>
<td>0.59</td>
</tr>
<tr>
<td>Varicose microvascular vessels</td>
<td>0.812</td>
<td>0.423, 1.201</td>
<td>&lt; 0.001</td>
<td>0.299</td>
<td>-0.155, 0.754</td>
<td>1</td>
<td>0.20</td>
</tr>
<tr>
<td>JNET classification, type 1</td>
<td>0.390</td>
<td>-0.477, 1.258</td>
<td>0.38</td>
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</tr>
</tbody>
</table>

P values were calculated using binomial logistic regression analysis.

SSL, sessile serrated lesion; MVHP, microvesicular hyperplastic polyp; CI, confidence interval; JNET, Japan narrow band imaging expert team.
FIGURE LEGENDS

Figure 1. **Endoscopic and histological images of serrated polyps.** A, B, C, D) Sessile serrated lesion; E, F, G, H) Microvesicular hyperplastic polyp; I, J, K, L) Goblet cell-rich hyperplastic polyp. A, E, I) White-light imaging; B, F, J) Texture and color enhancement imaging with indigo carmine dye; C, G, K) Narrow-band imaging with magnification. Black and red arrows show expanded crypt opening and varicose microvascular vessels, respectively. An EVIS X1 video system center (CV-1500) and a colonoscope (CF-XZ1200; Olympus Corporation) were used. D, H, L) Hematoxylin and eosin stain.

Figure 2. **ROC curve to predict histology of serrated polyp.** ROC curve was based on the endoscopic SSL/MVHP score. The mucus cap, expanded crypt opening, and varicose microvascular vessels were each assigned 1 point, and the sum of the points was defined as the endoscopic SSL/MVHP score. A) ROC curve to predict SSL (n = 159) and MVHP (n = 361) group from GCHP (n = 150); B) ROC curve to predict MVHP from GCHP. ROC, receiver operating characteristic; TPF, true positive fraction; FPF, false positive fraction; SSL, sessile serrated lesion; MVHP, microvesicular hyperplastic polyp; GCHP, goblet cell-rich hyperplastic polyp.