

S1-Guideline on Bacterial Vaginosis in Gynecology and Obstetrics

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S1-Leitlinie Bakterielle Vaginose in Gynäkologie und Geburtshilfe

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1 Introduction

Normal vaginal flora of sexually mature women consists of lactobacilli, transient or commensal anaerobic and aerobic bacteria, and *Candida* species from skin and gut flora. The normal pH value of vaginal flora is 3.8–4.4. Lactobacilli predominate in the vaginal flora, primarily *L. crispatus*, *L. gasseri*, *L. iners* and *L. jensenii*, depending on ethnicity. Normal pH values differ significantly between ethnicities, ranging from 3.8 to 5.2. Some healthy women have no lactobacilli in their vaginal flora; the percentage of women with lactobacilli differs depending on ethnicity; the percentage for Caucasian women is around 9%, for Hispanic and African women the percentage is more than 30% with normal pH values of around 5 [14,43]. At present, there are no studies into this issue for Europe.

Bacterial vaginosis (BV) is the most common microbiological disorder of the vaginal flora in sexually mature women. The prevalence of BV differs, depending on ethnicity; it is 2.4 times more common in women of African descent [13]. In Europe, the prevalence ranges from 5% in women who come for regular gynecological check-ups to more than 30% in women treated in clinics for sexually transmitted diseases. The reported prevalence in pregnant women is between 7 and 22% [3,9,15,22,25].

Psychosocial stress is a significant risk factor for BV [39]. There are also indications of gene–environment interactions as additional risk factors and of gene polymorphisms affecting genetic control of the individual's immune response to BV [35]. The abnormal oral conditions of periodontal disease have been found to be significantly correlated with disturbed vaginal flora in BV [46]. Vitamin D deficiency was identified as a risk factor for BV in pregnant women [13].

Microarray technology has been used to detect microorganisms associated with BV; in addition

to *Gardnerella* (*G.*) *vaginalis* and *Atopobium* (*A.*) *vaginae*, bacteria of the genera *Megasphaera*, *Dialister*, *Mobiluncus*, *Prevotella*, *Leptotrichia*, *Sneathia*, *Peptostreptococcus* and others were identified [4,14]. There are several strains of *G. vaginalis* with different properties [6].

The most common cause of BV is probably sexual transmission [6,7,10,42,53]. Vitamin D deficiency is also being discussed as a risk factor for BV [13]. Typically, the numbers of H₂O₂-producing lactobacilli species are reduced in women with BV compared to women without BV [8].

Only around 50% of affected women present with characteristic symptoms such as increased homogeneous vaginal discharge or a fishy odor on alkalization [26]. Many women with BV remain asymptomatic. Increased vaginal discharge can cause vulvar irritation.

2 Diagnosis

According to the diagnostic criteria proposed in some studies, BV is diagnosed when at least three of the following four findings are present at gynecological examination [1]:

- ▶ thin, homogeneous vaginal discharge
- ▶ vaginal pH greater than 4.5
- ▶ amine odor (particularly after alkalization with 10% potassium hydroxide solution [KOH])
- ▶ >20% clue cells on wet mount. (Gardner and Dukes [11] called the typical epithelial cells covered with bacteria found on vaginal wet mount “clue cells” because they offered a clue to the diagnosis of BV).

Alternatively, BV can be diagnosed by Gram-staining vaginal fluid (Nugent score) [40].

Typical findings for BV are decreased concentrations of certain facultative anaerobic *Lactobacillus* strains and 1000-fold higher concentrations of anaerobic microorganisms. *G. vaginalis* is also present at 100-fold higher concentrations. Thus,

Bibliography

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bacterial vaginosis is characterized by a microbial imbalance, with a marked shift towards increased concentrations of anaerobic microorganisms (see above) to the detriment of facultative anaerobic flora, particularly *Lactobacillus* species [4, 8, 16, 21, 30, 36]. Particularly *L. iners*, which is present in normal vaginal microflora, was found to have a tendency to replace *L. crispatus* or *L. gasseri*, with higher concentrations of *L. iners* found in the vaginal fluid of women with BV [58, 63].

3 Bacterial Biofilm

A prominent feature of BV is a polymicrobial biofilm adherent to the vaginal epithelium. The biofilm consists mainly of *G. vaginalis* and *A. vaginae*, but lactobacilli are also present in lower concentrations [50]. Bacterial biofilms are typical for chronic and/or foreign body-associated infections. Guideline-directed treatment does not eliminate the biofilm in women with BV [51]. The biofilm has also been detected in epithelial cells in the urine of women with BV and of their sexual partners [53] and in individual samples of washed donor semen from sperm banks [54] but not in the (peri-)anal area of women with BV [52]. The biofilm has also been found on endometrial samples obtained during curettage and on fallopian tube epithelia of women with BV [55]. Since 1978, standard recommended therapy consists of oral metronidazole [42], but metronidazole does not reliably eliminate the biofilm, even when clinical impression, pH and wet mount appear to show a cure [51] (see also Recommendations for Treatment below).

4 Bacterial Vaginosis and Gynecology

Women with BV have a higher risk of infection ascending from the cervix to the endometrium (endometritis) to the adnexa (salpingitis, tubo-ovarian abscess) [17, 28, 55]. The risk also appears to be increased in women with an intrauterine device in situ [2, 9, 11]. Endometritis resulting from BV can lead to abnormal bleeding [32]. Women with BV also have a greater predisposition to urinary tract infection [11, 26, 28]. Infectious morbidity after hysterectomy is also increased [12, 20, 31, 48, 49, 60]. There are some indications that BV represents a risk factor for ascending infection after termination of pregnancy and for spontaneous abortion [21, 29, 55].

5 Bacterial Vaginosis and Obstetrics

Pregnant women with BV have a higher risk of premature rupture of membranes, preterm labor and preterm birth due to ascending infection. Fever during and after delivery is also more common (post-partum endometritis and wound infection). These infections have been found to be histologically correlated with chorioamnionitis and with positive microbiological cultures from amniotic membranes and placenta. Women after cesarean section are particularly at risk [5, 12, 15, 17–19, 33, 34, 37, 47, 55, 56, 59].

Numerous studies have investigated the treatment of BV during pregnancy [27, 33, 34, 41]. Treatment consists of the systemic administration of either metronidazole or vaginal clindamycin cream. Some studies have indicated that systemic antibiotic treatment of BV after the 1st trimester of pregnancy can reduce the

number of preterm births in high risk groups (e.g. previous history of preterm birth). Intravaginal application appears to be less suitable to reduce the risk of preterm births in high risk groups.

Studies in Thuringia and Vienna have shown that screening for BV even in pregnant women without a history of preterm birth followed by treatment (oral or intravaginal application) is effective; these findings are supported by the meta-analysis of Varma and Gupta [22, 23, 25, 56].

In the Erfurt trial for the prevention of preterm births, pregnant women regularly measured their vaginal pH themselves. The active participation of pregnant women meant that this strategy detected changes in pH levels very early on, allowing a number of risk factors relevant for late spontaneous abortion and preterm birth to be treated quickly. Statistical analysis of the trial data confirmed the positive impact and represented a breakthrough with regard to the availability of a feasible and universally applicable measure to prevent preterm birth [45].

Preventing preterm births with the help of screening to detect and treat genital infections, particularly BV, is one strategy to optimize and rationalize healthcare. The cost-benefit analyses of this approach carried out by various parties have been controversially discussed and their methodology has been criticized [45]. Regular determination of vaginal pH has been found to be widely accepted by pregnant women, making the method highly feasible [22, 23].

6 Recommendations for Diagnosis

The criteria listed above should be used to exclude BV in all women prior to placing an intrauterine device and prior to any intrauterine intervention. Women should receive comprehensive antibiotic treatment either prior to intervention or perioperatively. A predisposition to BV should be considered in patients with bleeding disorders or recurrent urinary tract infections.

Determination of vaginal pH and wet mount of vaginal flora are suggested for screening prior to a planned pregnancy or early on in pregnancy. These diagnostic measures can be used to exclude genital infections with a high level of confidence and with little expenditure of time. This applies particularly to women with a history of preterm birth. Antibiotic treatment is indicated if BV is diagnosed. Pregnant women with a history of preterm birth and a diagnosis of BV should receive systemic treatment with antibiotics. Wet mount evaluation should be done prior to terminating a pregnancy.

7 Recommendations for Treatment

Metronidazole and 2% clindamycin vaginal cream are two highly effective anaerobicides used to treat bacterial vaginosis. Non-pregnant women can be treated with oral metronidazole administered 2×500 mg/day for 7 days. Both a single dose administration of oral metronidazole and 2×2 g metronidazole administered at an interval of 48 hours have acceptable cure rates. Good results were also reported for intravaginal metronidazole applications consisting of $1-2 \times 500$ mg metronidazole vaginal tablets daily for 7 days. The daily application of 5 g of 2% clindamycin vaginal cream for 7 days is another effective alternative to treat BV. Placebo-controlled studies have reported similar cure rates for 1×2 g oral metronidazole and 2×1 g metronidazole applied

intravaginally on 2 consecutive days (89.9% vs. 92.5%), although compliance was significantly higher for vaginal applications [24]. Individual studies have reported recurrence rates after treatment with 10 mg dequalinium chloride vaginal tablets for 6 days [61] or 1 × 250 mg nifuratel vaginal tablets for 10 days [38] similar to the recurrence rates for standard metronidazole treatment.

7.1 Treatment of the bacterial biofilm

None of the recommended treatments eliminate the adherent bacterial biofilm [51]. There is currently no evidence-based treatment available that minimizes the risk of chronic manifestation and recurrence. The bacterial biofilm and its presence in the upper genital tract of women and in partners of women with BV appears to be the explanation for the fact that the cure rate after 3 months is only 60–70% and is even lower after 6 months [30, 57].

New findings from studies of probiotic *Lactobacillus* strains and clinical studies examining ways of lowering the recurrence rates of BV after standard treatment using probiotics or acidic applications to reduce vaginal pH have shown that these methods can be effective and found that they reduced the rate of BV recurrence by around half [24, 44, 62].

7.2 Treatment during pregnancy

Despite theoretical concerns and after consultation with the patient, metronidazole can be administered systemically as described above after the 1st trimester of pregnancy to treat pregnant women with BV. Alternatively, treatment can also consist of local vaginal application of 500–1000 mg metronidazole for 7 days. Oral clindamycin 2 × 300 mg/day for 7 days can be prescribed after the 1st trimester of pregnancy. Daily intravaginal application of 5 g of 2% clindamycin vaginal cream for 7 days has similar cure rates as metronidazole; the reported side-effects are minimal and there are no concerns about its application during pregnancy. However, studies have shown that treatment of BV during pregnancy as a prophylactic measure against preterm birth in high risk patients (e.g. previous history of preterm birth) is only effective if treatment is systemic [34].

7.3 Co-treatment of the partner

Although the typical bacterial biofilm found with BV has also been found in cells from the urine or sperm of male partners of women with BV, there is currently no scientific basis for routine co-treatment of partners, and it is therefore currently not recommended.

Consensus Process

These recommendations were compiled by the following members of the *Arbeitsgemeinschaft für Infektionen und Infektionsimmunologie* (Professional Society for Infections and Infection Immunology [AGII]) of the *Deutsche Gesellschaft für Gynäkologie und Geburtshilfe* (German Society for Gynecology and Obstetrics [DGGG e.V.]):

- ▶ Prof. Dr. med. Werner Mendling, Wuppertal (overall responsibility)
- ▶ Prof. Dr. med. Joachim Martius, Agatharied
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The present version was approved by all of the authors in July 2013.

The revision was confirmed in August 2013 by the Board of the *Deutsche Gesellschaft für Gynäkologie und Geburtshilfe* (German Society for Gynecology and Obstetrics).

The Guideline will remain valid until 08/2016.

Conflict of Interest

Prof. Mendling received royalties for scientific consultancy or lectures by Abbott GmbH & Co. KG Wiesbaden/Germany, Medinova AG Zürich/Switzerland, Pierre Fabre GmbH Freiburg/Germany, Dr. August Wolff GmbH & Co. KG Bielefeld/Germany and Polichem S.A. Lugano/Switzerland. Prof. Martius declares no conflict of interest. Prof. Hoyme received royalties by Dr. August Wolff GmbH & Co. KG Bielefeld.

References

- 1 Amsel R, Totten PA, Spiegel CA et al. Nonspecific vaginitis. Diagnostic criteria and microbial and epidemiologic associations. *Am J Med* 1983; 74: 14–22
- 2 Avonts D, Sercu M, Heyerick P et al. Incidence of uncomplicated genital infections in women using oral contraception or an intrauterine device: a prospective study. *Sex Transm Dis* 1990; 17: 23–29
- 3 Desseauve D, Chantrel J, Fruchart A et al. Prevalence and risk factors of bacterial vaginosis during the first trimester of pregnancy in a large French population-based study. *Eur J Obstet Gynecol Reprod Biol* 2012; 163: 30–34
- 4 Dols JAM, Smit PW, Kort R et al. Microarray-based identification of clinically relevant vaginal bacteria in relation to bacterial vaginosis. *Am J Obstet Gynecol* 2011; 204: 305.e1–305.e7
- 5 Donders GGG, van Bulk B, Caudron J et al. Relationship of bacterial vaginosis and mycoplasmas to the risk of spontaneous abortion. *Am J Obstet Gynecol* 2000; 183: 431–437
- 6 Elsner P, Hartmann AA. Gardnerella vaginalis in the male upper genital tract: a possible source of reinfection of the female partner. *Sex Transm Dis* 1987; 14: 122–123
- 7 Eren AM, Zozaya M, Taylor CM et al. Exploring the diversity of Gardnerella vaginalis in the genitourinary tract microbiota of monogamous couples through subtle nucleotide variation. *PLoS One* 2011; 6: e26732
- 8 Eschenbach DA, Davick PR, Williams BL et al. Prevalence of hydrogen peroxide-producing *Lactobacillus* species in normal women and women with bacterial vaginosis. *J Clin Microbiol* 1989; 27: 251–256
- 9 Eschenbach DA, Hillier S, Critchlow C et al. Diagnosis and clinical manifestations of bacterial vaginosis. *Am J Obstet Gynecol* 1988; 158: 819–828
- 10 Gardner HC, Dukes CD. Haemophilus vaginalis vaginitis: a newly defined specific infection previously classified non-specific vaginitis. *Am J Obstet Gynecol* 1955; 69: 962–976
- 11 Harmanli OH, Cheng GY, Nyirjesy P et al. Urinary tract infections in women with bacterial vaginosis. *Obstet Gynecol* 2000; 95: 710–712
- 12 Hauth JC, MacPherson C, Carey JC et al. Early pregnancy threshold vaginal pH and Gram stain scores predictive of subsequent preterm birth in asymptomatic women. *Am J Obstet Gynecol* 2003; 188: 831–835
- 13 Hensel KJ, Randis TM, Gelber SE et al. Pregnancy-specific association of vitamin D deficiency and bacterial vaginosis. *Am J Obstet Gynecol* 2011; 204: 41.e1–41.e9
- 14 Hickley RJ, Zhou X, Pierson JD et al. Understanding vaginal microbiome completely from an ecological perspective. *Transl Res* 2012; 160: 267–282
- 15 Hillier S, Holmes KK. Bacterial Vaginosis. In: Holmes KK, Sparling PF, Mardh P-A, Lemon SM, Stamm WE, Wasserheit JN, eds. Sexually transmitted Diseases. New York: McGraw-Hill; 1999: 563–586
- 16 Hillier SL, Critchlow CW, Stevens CE et al. Microbiological, epidemiological and clinical correlations of vaginal colonisation by *Mobiluncus* species. *Genitourin Med* 1991; 67: 26–31
- 17 Hillier SL, Kiviat NB, Hawes SE et al. Role of bacterial vaginosis-associated microorganisms in endometritis. *Am J Obstet Gynecol* 1996; 175: 435–441

- 18 Hillier SL, Martius J, Krohn M et al. A case-control study of chorioamnionic infection and histologic chorioamnionitis in prematurity. *N Engl J Med* 1988; 319: 972–978
- 19 Hillier SL, Nugent RP, Eschenbach DA et al. Association between bacterial vaginosis and preterm delivery of a low-birthweight infant. *N Engl J Med* 1995; 333: 1737–1742
- 20 Hooton TM, Fihn SD, Johnson C et al. Association between bacterial vaginosis and acute cystitis in women using diaphragmas. *Arch Intern Med* 1989; 149: 1932–1936
- 21 Hoyne UB, Eschenbach DA. Bakterielle Vaginose. Mikrobiologie, Diagnostik, Therapie und mögliche Komplikationen. *Dtsch Med Wochenschr* 1985; 110: 349–352
- 22 Hoyne UB, Möller U, Saling E. Aktuelle Aspekte der Thüringer Frühgeburtenvermeidungs-Aktion 2000. *Zentralbl Gynäkol* 2003; 125: 107–111
- 23 Hoyne UB, Schwalbe N, Saling E. Die Effizienz der Thüringer Frühgeburtenvermeidungs-Aktion 2000 wird durch die Perinatalstatistik der Jahre 2001–2003 bestätigt. *Geburtsh Frauenheilk* 2005; 65: 284–288
- 24 Hoyne UB, Brandt M, May TW et al. Sequentielle intravaginale Gabe von Metronidazol und Milchsäure zur Behandlung und Rezidivprävention bei bakterieller Vaginose. *Geburtsh Frauenheilk* 2009; 69: 395–400
- 25 Kiss H, Petricevic L, Husslein P. Prospective randomized controlled trial of an infection screening program to reduce the rate of preterm delivery. *BMJ* 2004; 329: 371–375
- 26 Klebanoff MA, Schwabke JR, Zhang J et al. Vulvovaginal symptoms in women with bacterial vaginosis. *Obstet Gynecol* 2004; 104: 267–272
- 27 Lamont RF, Duncan SLB, Mandal D et al. Intravaginal clindamycin to reduce preterm birth in women with abnormal genital tract flora. *Obstet Gynecol* 2003; 101: 516–522
- 28 Larsson PG, Bergmann B, Forsum U et al. Treatment of bacterial vaginosis in women with vaginal bleeding complications or discharge and harboring *Mobiluncus*. *Gynecol Obstet Invest* 1990; 29: 296–300
- 29 Larsson PG, Bergmann B, Forsum U et al. *Mobiluncus* and clue cells as predictors of PID after first trimester abortion. *Acta Obstet Gynecol Scand* 1989; 68: 217–220
- 30 Larsson PG, Forsum U. Bacterial vaginosis – a disturbed bacterial flora and treatment enigma. *APMIS* 2005; 113: 305–316
- 31 Larsson PG, Platz-Christensen JJ, Forsum U et al. Clue cells in predicting infections after abdominal hysterectomy. *Obstet Gynecol* 1991; 77: 450–452
- 32 Lefevre JC, Averous S, Bauriaud R et al. Lower genital tract infections in women: comparison of clinical and epidemiological findings with microbiology. *Sex Transm Dis* 1988; 15: 110–113
- 33 Leitich H, Bodner-Adler B, Brunbauer M et al. Bacterial vaginosis as a risk factor for preterm delivery: a meta-analysis. *Am J Obstet Gynecol* 2003; 189: 139–147
- 34 Leitich H, Brunbauer M, Bodner-Adler B et al. Antibiotic treatment of bacterial vaginosis in pregnancy: a meta-analysis. *Am J Obstet Gynecol* 2003; 188: 752–758
- 35 Macones GA, Parry S, Elkousy M et al. A polymorphism in the promoter region of TNF and bacterial vaginosis: preliminary evidence of gene-environment interaction in the etiology of spontaneous preterm birth. *Am J Obstet Gynecol* 2004; 190: 1504–1508
- 36 Martius J, Krohn M, Hillier SL et al. Relationship of vaginal *Lactobacillus* species, cervical *Chlamydia trachomatis*, and bacterial vaginosis to preterm birth. *Obstet Gynecol* 1988; 71: 89–95
- 37 McGregor JA, French JL, Seo K. Premature rupture of the membranes and bacterial vaginosis. *Am J Obstet Gynecol* 1993; 169: 463–466
- 38 Mendling W, Caserini M, Palmieri R. A randomized, controlled study to assess the efficacy and safety of nifuratel vaginal tablets on bacterial vaginosis. *Sex Transm Infect* 2013; 9 (Suppl. 1): A28
- 39 Nansel TR, Riggs MA, Yu K-F et al. The association of psychosocial stress and bacterial vaginosis in a longitudinal cohort. *Am J Obstet Gynecol* 2006; 194: 381–386
- 40 Nugent RP, Krohn MA, Hillier SL. Reliability of diagnosing bacterial vaginosis is improved by a standardized method of Gram stain interpretation. *J Clin Microbiol* 1991; 29: 297–301
- 41 Okun N, Gronau KA, Hannah ME. Antibiotics for bacterial vaginosis or trichomoniasis in pregnancy: a systematic review. *Obstet Gynecol* 2005; 105: 857–868
- 42 Pheifer TA, Forsyth PS, Durfee MA et al. Nonspecific vaginitis: role of *Haemophilus vaginalis* and treatment with metronidazole. *N Engl J Med* 1978; 298: 1429–1434
- 43 Ravel J, Gajer P, Abdo Z et al. Vaginal microbiome of reproductive-age women. *Proc Natl Acad Sci USA* 2011; 108 (Suppl. 1): 4680–4687
- 44 Reid G, Burton J, Devillard E. The rationale for probiotics in female urogenital healthcare. *MedGenMed* 2004; 29: 49
- 45 Saling E. Problems in prevention of preterm birth – regrettable contradictions. *J Perin Med* 2011; 39: 223–225
- 46 Sanu O, Lamont RF. Periodontal disease and bacterial vaginosis as genetic and environmental markers for the risk of spontaneous preterm labor and preterm birth. *J Matern Fetal Neonatal Med* 2011; 24: 1476–1485
- 47 Simhan HN, Caritis SN, Krohn M et al. The vaginal inflammatory milieu and the risk of early premature preterm rupture of membranes. *Am J Obstet Gynecol* 2005; 192: 213–218
- 48 Soper DE. Bacterial vaginosis and postoperative infections. *Am J Obstet Gynecol* 1993; 169: 467–469
- 49 Soper DE, Bump RC, Hurt WG. Bacterial vaginosis and trichomoniasis vaginitis are risk factors for cuff cellulitis after abdominal hysterectomy. *Am J Obstet Gynecol* 1990; 163: 1016–1023
- 50 Swidsinski A, Mendling W, Loening-Baucke V et al. Adherent biofilms in bacterial vaginosis. *Obstet Gynecol* 2005; 106: 1013–1023
- 51 Swidsinski A, Mendling W, Loening-Baucke V et al. An adherent *Gardnerella vaginalis* biofilm persists on the vaginal epithelium after standard therapy with oral metronidazole. *Am J Obstet Gynecol* 2008; 198: 97.e1–97.e6
- 52 Swidsinski A, Dörffel Y, Loening-Baucke V et al. Dissimilarity in the occurrence of bifidobacteriaceae in vaginal and perianal microbiota in women with bacterial vaginosis. *Anaerobe* 2010; 16: 478–482
- 53 Swidsinski A, Dörffel Y, Loening-Baucke V et al. *Gardnerella* biofilm involves females and males and is sexually transmitted. *Gynecol Obstet Invest* 2010; 70: 256–263
- 54 Swidsinski A, Dörffel Y, Loening-Baucke V et al. Desquamated epithelial cells covered with a polymicrobial biofilm typical for bacterial vaginosis are present in randomly selected cryopreserved donor semen. *FEMS Immunol Med Microbiol* 2010; 59: 399–404
- 55 Swidsinski A, Verstraelen H, Loening-Baucke V et al. Presence of a polymicrobial endometrial biofilm in patients with bacterial vaginosis. *PLoS One* 2013; 8: e53997
- 56 Varma R, Gupta JK. Antibiotic treatment of bacterial vaginosis in pregnancy: Multiple meta-analysis and dilemmas in interpretation. *Eur J Obstet Gynecol Reprod Biol* 2006; 124: 10–14
- 57 Verstraelen H, Verhelst R. Bacterial vaginosis: an update on diagnosis and treatment. *Expert Rev Anti Infect Ther* 2009; 7: 1109–1124
- 58 Verstraelen H, Verhelst R, Claeys G et al. Longitudinal analysis of the vaginal microflora in pregnancy suggests that *L. crispatus* promotes the stability of the normal vaginal microflora and that *L. gasseri* and/or *L. iners* are more conducive to the occurrence of abnormal vaginal microflora. *BMC Microbiol* 2009; 9: 116
- 59 Watts DH, Eschenbach DA, Kenny GE. Early postpartum endometritis: the role of bacteria, genital mycoplasmas, and *Chlamydia trachomatis*. *Obstet Gynecol* 1989; 73: 52–60
- 60 Watts DH, Krohn M, Hillier SL et al. Bacterial vaginosis as a risk factor for post-cesarean endometritis. *Obstet Gynecol* 1990; 75: 52–58
- 61 Weissenbacher ER, Donders G, Unzeitig V et al.; Fluomizin Study Group. A comparison of dequalinium chloride vaginal tablets (Fluomizin®) and clindamycin vaginal cream in the treatment of bacterial vaginosis: a single-blind, randomized clinical trial of efficacy and safety. *Gynecol Obstet Invest* 2012; 73: 8–15
- 62 Ya W, Reifer C, Miller LE. Efficacy of vaginal probiotic capsules for recurrent bacterial vaginosis: a double blind, randomized, placebo-controlled study. *Am J Obstet Gynecol* 2010; 203: 120.e1–120.e6
- 63 Zozaya-Hinchliffe M, Lillis R, Martin DH et al. Quantitative PCR assessments of bacterial species in women with and without bacterial vaginosis. *J Clin Microbiol* 2010; 48: 1812–1819