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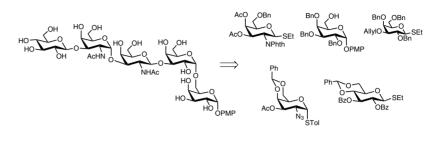
Paper

Efficient Synthesis of the Pentasaccharide Repeating Unit of the O-Antigenic Polysaccharide of *Escherichia coli* O166 Strain

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Abstract An efficient strategy has been developed for the synthesis of the pentasaccharide repeating unit of the cell-wall polysaccharide of *Escherichia coli* O166 strain through sequential stereoselective glycosylations of monosaccharide intermediates. All the glycosylation steps were high-yielding with high stereoselectivities.

Key words carbohydrates, glycosylations, glycosides, stereoselectivity, oligosaccharides

Diarrheal outbreaks and gastrointestinal complications are important health problems in developing countries.¹ In general, intake of contaminated food and water and a lack of adequate sanitation are the leading causes of enteric disorders.² Recently, gastrointestinal infections have also become significant health hazards in developed countries.³ Among the several enteropathogenic microbes that are responsible for the diarrheal infections, pathogenic strains of Escherichia coli merit particular attention. These are associated with several gastrointestinal infections, particularly 'travelers' diarrhea' and they can be classified into several pathotypes, such as enteropathogenic, enterohemorrhagic, enterotoxigenic, enteroinvasive, enteroaggregative, or diffusely adherent.⁴ The O166 strain of *E. coli* is generally classed as belonging to the enteroaggregative pathotype and causes diarrhea in human by producing a heat-stable enterotoxin.⁵ E. coli O166 has also been isolated from the environment and from cattle, and has also been classified as an enterohemorrhagic strain.⁶ In 1996, E. coli O166 was identified as the cause of an outbreak of diarrhea in Japan.⁷

The target pentasaccharide **1** was synthesized as its 4methoxyphenyl glycoside by a series of stereoselective sequential glycosylation reactions of suitably functionalized monosaccharide intermediates. For this purpose, the monosaccharide intermediates **2**, **3**,¹⁰ **4**,¹¹ **5**,¹² and **6**¹³ were prepared by following the methods previously reported (Figure 1).

Cell-wall polysaccharides of virulent strains of bacteria play crucial roles in the initial stages of bacterial infections in hosts. As result, researchers have focused attention on the characterization of cell-wall O-antigenic polysaccharides from several bacterial strains. Recently, the structure of the pentasaccharide repeating unit of the O-antigenic polysaccharide of E. coli O166 was reported by Ali et al.8 This pentasaccharide contains D-glucose, D-galactose, and *N*-acetyl-D-galactosamine moieties. As the result of the acceleration in the failure of antibiotics to act on multidrugresistant strains of bacteria, the development of alternative approaches to the control of bacterial infections is currently a major area in drug-discovery research.⁹ It is therefore relevant to develop therapeutics based on glycoconjugate derivatives related to cell-wall polysaccharide O-antigens, because of their involvement in the process of bacterial infection. Sufficient quantities of oligosaccharides free of biological impurities are required for biological studies, and these cannot readily be isolated from natural sources. Therefore, the development of efficient strategies for chemical synthesis of these oligosaccharides would be extremely useful in providing access to significant quantities of pure oligosaccharides with appropriate structures. In this context, an efficient synthesis of the pentasaccharide repeating unit of the O-antigenic polysaccharide of E. coli O166 has been developed.

Syn<mark>thesis</mark>

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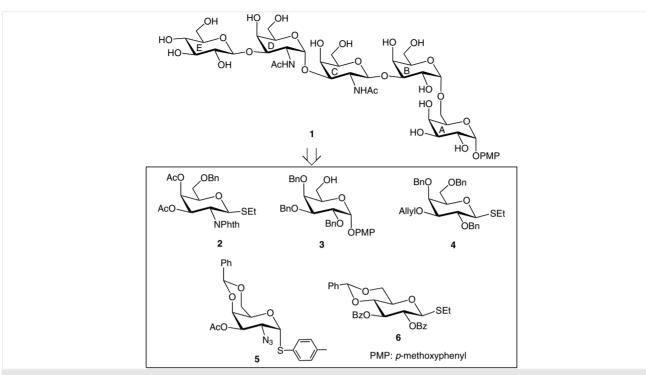


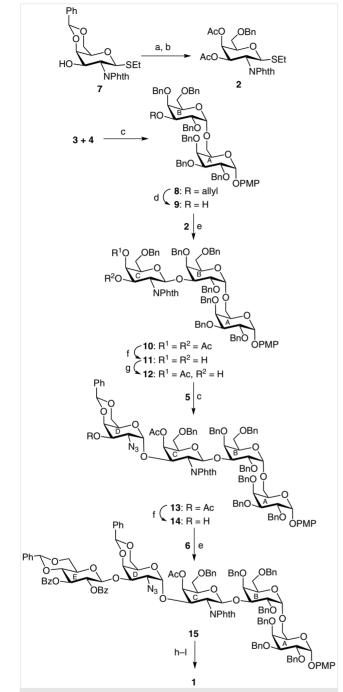
Figure 1 Structures of the synthesized pentasaccharide repeating unit of the O-antigenic polysaccharide of E. coli O166 (1) and various monosaccharide intermediates (2–6)

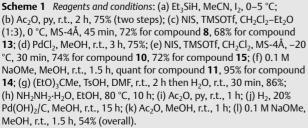
Treatment of the known ethyl 4,6-*O*-benzylidene-2-deoxy-2-(*N*-phthalimido)-1-thio- β -D-galactopyranoside (**7**)¹⁴ with triethylsilane in the presence of molecular iodine,¹⁵ followed by acetylation with acetic anhydride and pyridine¹⁶ gave ethyl 3,4-di-*O*-acetyl-6-*O*-benzyl-2-deoxy-2-(*N*-phthalimido)-1-thio- β -D-galactopyranoside (**2**) in 75% overall yield (Scheme 1).

Stereoselective 1,2-cis glycosylation of the D-galactosyl donor $\mathbf{3}^{10}$ with the D-galactosyl acceptor $\mathbf{4}^{11}$ in the presence of N-iodosuccinimide and trimethylsilyl trifluoromethanesulfonate^{17,18} in dichloromethane-diethyl ether gave the disaccharide derivative 8 in 72% yield, together with a minor quantity (~8%) of another isomer. Disaccharide 8 was purified by column chromatography and its stereoselective formation was confirmed by spectroscopic analysis. Removal of the allyl ether group from disaccharide 8 by using palladium(II) chloride¹⁹ gave the partially deprotected disaccharide 9 in 75% yield. Stereoselective 1,2-trans glycosylation of disaccharide 9 with the D-galactosamine donor 2 in the presence of N-iodosuccinimide and trimethylsilyl trifluoromethanesulfonate^{17,18} gave the trisaccharide derivative **10** in 74% yield. The exclusive formation of compound 10 was confirmed by NMR spectroscopy. De-O-acetylation of compound 10 with sodium methoxide²⁰ gave the trisaccharide diol derivative 11, which was selectively 4-O-acetylated through the formation of an ortho ester²¹ and subsequent acidic hydrolysis to give trisaccharides 12 in 86% overall yield. Stereoselective 1,2-cis glycosylation of trisaccharide

12 with the D-galactosamine derivative 5¹² as a glycosyl donor in the presence of N-iodosuccinimide and trimethylsilyl trifluoromethanesulfonate^{17,18} in dichloromethane-diethyl ether gave the tetrasaccharide derivative 13 in 68% yield, together with a minor quantity of the trans-glycosylation product (~10%), which was separated by column chromatography. Stereoselective formation of the tetrasaccharide derivative 13 was confirmed by spectroscopic analysis. Selective removal²² of the 3-O-acetyl group from compound 13 by using sodium methoxide left the internally located 4-O-acetyl group unaffected and gave the tetrasaccharide acceptor **14** in 95% vield. *N*-Iodosuccinimide–trimethylsilvl trifluoromethanesulfonate-mediated 1,2-trans-glycosylation of tetrasaccharide 14 with the D-glucose thioglycoside derivative **6**¹³ in dichloromethane gave the pentasaccharide derivative 15 in 72% yield. NMR spectroscopic analysis of compound 15 confirmed that it was formed exclusively. Finally, pentasaccharide 15 was subjected to a series of reactions to remove the protecting groups completely. These reactions included (a) removal of the N-phthaloyl group by treatment with hydrazine monohydrate,23 followed by acetylation of the resulting amine using acetic anhydride and pyridine; (b) removal of the benzyl ethers and benzylidene acetals and reduction of the azido group by hydrogenolysis over palladium(II) hydroxide/carbon,²⁴ followed by N-acetylation with acetic anhydride in methanol; and (c) removal of the acetyl and benzoyl groups by treatment with sodium methoxide to give the target pentaA. Si, A. K. Misra

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saccharide **1** in 54% overall yield. The formation of compound **1** was unambiguously confirmed by spectroscopic analysis.

In summary, a straightforward strategy has been developed for the synthesis of the pentasaccharide repeating unit of the O-antigen of *Escherichia coli* O166 by a series of sequential stereoselective glycosylations of monosaccharide intermediates. The glycosylation steps were high-yielding and gave an excellent stereochemical outcome. Similar reaction conditions were used in each of the glycosylation reactions.

All reactions were monitored by TLC on silica gel coated plates. TLC spots were visualized by spraying the plates with ceric sulfate [2% $Ce(SO_4)_2$ in 2 N H₂SO₄] and warming the sprayed TLC plates on a hot-plate. Silica gel (230–400 mesh) was used for column chromatography. NMR spectra were recorded on Bruker Avance 500 MHz spectrometer with CDCl₃ as solvent and TMS as the internal reference, unless stated otherwise. Chemical shifts (δ =) are expressed in ppm. Complete assignment of the ¹H and ¹³C NMR spectra was carried out by means of a standard set of NMR experiments, e.g. ¹H NMR, ¹³C NMR, ¹³C DEPT 135, 2D COSY, and 2D HSQC. MALDI mass spectra were recorded on a Bruker mass spectrometer. Optical rotations were recorded in a JASCO P-2000 polarimeter. Commercially available grades of organic solvents of adequate purity were used in all reactions.

Ethyl 3,4-Di-O-acetyl-6-O-benzyl-2-deoxy-2-(*N*-phthalimido)-1-thio-β-D-galactopyranoside (2)

Et₃SiH (1.8 mL, 11.27 mmol) and I₂ (250 mg, 0.98 mmol) were added sequentially to a solution of monosaccharide derivative **7** (2 g, 4.53 mmol) in MeCN (10 mL) at 0–5 °C, and the mixture was stirred at 0–5 °C for 40 min. The mixture was then diluted with CH₂Cl₂ (100 mL) and washed successively with 5% aq Na₂S₂O₃ (50 mL) and H₂O (100 mL), then dried (Na₂SO₄) and concentrated. A solution of the crude product in Ac₂O (5 mL) and pyridine (5 mL) was kept at r.t. for 2 h. The reagents were removed under reduced pressure and the crude product was purified by chromatography [silica gel, hexane–EtOAc (3:1)] to give a yellow oil; yield: 1.8 g (75%); $[\alpha]_D^{23}$ +43 (*c* 1.0, CHCl₃).

IR (neat): 3024, 1736, 1515, 1372, 1216, 1096, 766 cm⁻¹.

¹H NMR (500 MHz, CDCl₃): δ = 7.83–7.21 (m, 9 H, Ar-H), 5.79 (dd, *J* =11.0, 3.0 Hz, 1 H, H-3), 5.55 (d, *J* =2.0 Hz, 1 H, H-4), 5.44 (d, *J* = 10.5 Hz, 1 H, H-1), 4.57 (d, *J* = 12.0 Hz, 1 H, PhCH₂), 4.53 (t, *J* = 10.5 Hz, 1 H, H-2), 4.43 (d, *J* = 12.0 Hz, 1 H, PhCH₂), 4.07–4.04 (m, 1 H, H-5), 3.59–3.56 (m, 1 H, H-6_a), 3.50–3.46 (m, 1 H, H-6_b), 2.71–2.63 (m, 2 H, SCH₂CH₃), 2.08–1.82 (2 s, 6 H, 2 COCH₃), 1.26 (t, *J* = 7.4 Hz, 3 H, SCH₂CH₃).

 ^{13}C NMR (125 MHz, CDCl₃): δ = 170.0, 169.5 (2 COCH₃), 167.8, 167.3 (PhthCO), 137.6–123.6 (m, Ar-C), 81.5 (C-1), 75.9 (C-3), 73.5 (PhCH₂), 69.0 (C-4), 67.6 (C-6), 67.4 (C-5), 50.3 (C-2), 24.4 (SCH₂CH₃), 20.7, 20.5 (2 COCH₃), 14.9 (SCH₂CH₃).

ESI-MS: 550.1 [M + Na]+.

Anal. Calcd for $C_{27}H_{29}NO_8S$ (527.16): C, 61.47; H, 5.54. Found: C, 61.30; H, 5.70.

4-Methoxyphenyl (3-O-Allyl-2,4,6-tri-O-benzyl-α-D-galactopyranosyl)-(1→6)-2,3,4-tri-O-benzyl-α-D-galactopyranoside (8)

MS-4Å (3 g) were added to a solution of compound **3** (1.5 g, 2.69 mmol) and compound **4** (1.6 g, 3.0 mmol) in anhyd 1:3 CH₂Cl₂–Et₂O (20 mL), and the mixture was cooled to 0 °C under argon. The cooled mixture was treated with NIS (0.7 g, 3.11 mmol) and TMSOTf (15 μ L) then stirred at 0 °C for 45 min. The mixture was then diluted with CH₂Cl₂ (100 mL) and washed successively with 5% aq Na₂S₂O₃ (50 mL), sat. aq NaHCO₃ (100 mL), and H₂O (100 mL). The organic phase was then dried (Na₂SO₄) and concentrated under reduced pressure to give a crude product that was purified by chromatography [silica gel, hexane–EtOAc (4:1)] to give a colorless oil; yield: 2 g (72%); [α]_D²³ +60 (c 1.0, CHCl₃).

IR (neat): 3025, 2363, 1719, 1509, 1387, 1216, 1097, 758 cm⁻¹.

¹H NMR (500 MHz, CDCl₃): δ = 7.39–7.20 (m, 30 H, Ar-H), 6.98 (d, *J* = 9.0 Hz, 2 H, Ar-H), 6.74 (d, *J* = 9.0 Hz, 2 H, Ar-H), 5.96–5.86 (m, 1 H, CH=CH₂), 5.37 (d, *J* = 2.5 Hz, 1 H, H-1_A), 5.32–5.28 (m, 1 H, CH=CH₂), 5.15–5.13 (m, 1 H, CH=CH₂), 4.95–4.67 (7 d, *J* = 11.0 Hz each, 7 H, PhCH₂), 4.65 (d, *J* = 3.5 Hz, 1 H, H-1_B), 4.58–4.34 (5 d, *J* = 11.0 Hz each, 5 H, PhCH₂), 4.14–4.11 (m, 5 H, H-2_A, H-5_A, H-5_B, OCH₂=CH), 3.94 (br s, 1 H, H-4_A), 3.90–3.88 (m, 2 H, H-2_B, H-3_A), 3.85 (br s, 1 H, H-4_B), 3.71–3.65 (m, 1 H, H-6_{aA}), 3.67 (s, 2 H, OCH₃), 3.58–3.53 (m, 2 H, H-3_B, H-6_{aB}), 3.49–3.46 (dd, *J* = 12.0, 5.5 Hz, 1 H, H-6_{bB}), 3.41–3.39 (dd, *J* = 12.0, 4.5 Hz, 1 H, H-6_{bA}).

¹³C NMR (125 MHz, CDCl₃): δ = 154.8–114.4 (m, Ar-C, CH₂=CH), 98.0 (C-1_B), 96.9 (C-1_A), 78.9 (C-5_A), 78.7 (C-5_B), 76.4 (C-3_B), 76.1 (C-3_A), 75.4 (C-4_B), 74.7 (2 C, 2 PhCH₂), 74.6 (C-4_A), 73.6 (PhCH₂), 73.4 (PhCH₂), 73.3 (2 C, 2 PhCH₂), 71.4 (OCH₂), 70.3 (C-2_A), 69.1 (C-2_B), 68.6 (C-6_B), 67.4 (C-6_A), 55.3 (OCH₃).

MALDI-MS: 1051.4 [M + Na]⁺.

Anal. Calcd for $C_{64}H_{68}O_{12}$ (1028.47): C, 74.69; H, 6.66. Found: C, 74.54; H, 6.80.

4-Methoxyphenyl (2,4,6-Tri-O-benzyl-α-D-galactopyranosyl)- (1→6)-2,3,4-tri-O-benzyl-α-D-galactopyranoside (9)

PdCl₂ (125 mg, 0.70 mmol) was added to a solution of compound **8** (1.8 g, 1.75 mmol) in anhyd MeOH (25 mL), and the mixture was stirred at r.t. for 3 h. The mixture was then concentrated under reduced pressure and purified by chromatography [silica gel, hexane–EtOAc (4:1)] to give a colorless oil; yield: 1.3 g (75%); $[\alpha]_D^{23}$ +95 (*c* 1.0, CHCl₃).

IR (neat): 3020, 2362, 1722, 1599, 1513, 1426, 1217, 1046, 927, 761 $\rm cm^{-1}.$

¹H NMR (500 MHz, CDCl₃): δ = 7.39–7.24 (m, 30 H, Ar-H), 6.97 (d, *J* = 9.0 Hz, 2 H, Ar-H), 6.72 (d, *J* = 9.0 Hz, 2 H, Ar-H), 5.33 (d, *J* = 2.5 Hz, 1 H, H-1_A), 4.96–4.70 (6 d, *J* = 11.0 Hz each, 6 H, PhCH₂), 4.67 (d, *J* = 2.5 Hz, 1 H, H-1_B), 4.60–4.34 (6 d, *J* = 11.0 Hz each, 6 H, PhCH₂), 4.13–4.10 (m, 3 H, H-2_A, H-5_A, H-5_B), 3.92 (br s, 1 H, H-4_A), 3.89–3.83 (m, 2 H, H-2_B, H-3_A), 3.79 (br s, 1 H, H-4_B), 3.70 (s, 3 H, OCH₃), 3.68–3.66 (m, 2 H, H-3_B, H-6_{aA}), 3.52–3.48 (m, 2 H, H-6_{abB}), 3.35–3.32 (m, 1 H, H-6_{bA}).

¹³C NMR (125 MHz, CDCl₃): δ = 155.1–114.4 (m, Ar-C), 97.5 (C-1_B), 97.1 (C-1_A), 78.9 (C-5_A), 77.3 (C-5_B), 76.7 (C-4_B), 76.4 (C-3_A), 75.4 (C-4_A), 75.1 (PhCH₂), 74.6 (PhCH₂), 73.4 (2 C, 2 PhCH₂), 73.3 (PhCH₂), 72.7 (PhCH₂), 70.2 (C-3_B), 69.9 (C-2_A), 68.9 (C-2_B), 68.6 (C-6_B), 67.3 (C-6_A), 55.4 (OCH₃).

MALDI-MS: 1011.4 [M + Na]+.

Anal. Calcd for $C_{61}H_{64}O_{12}$ (988.44): C, 74.07; H, 6.52. Found: C, 73.87; H, 6.70.

4-Methoxyphenyl [3,4-Di-O-acetyl-6-O-benzyl-2-deoxy-2-(N-phthalimido)- β -D-galactopyranosyl]-(1 \rightarrow 3)-(2,4,6-tri-O-benzyl- α -D-galactopyranosyl)-(1 \rightarrow 6)-2,3,4-tri-O-benzyl- α -D-galactopyranoside (10)

MS-4Å (1 g) were added to a solution of compound **9** (1.2 g, 1.21 mmol) and compound **2** (960 mg, 1.82 mmol) in anhyd CH₂Cl₂ (10 mL), and the mixture was cooled to -20 °C under argon. NIS (560 mg, 2.49 mmol) and TMSOTf (10 μ L) were added, and the cooled mixture was stirred at -20 °C for 30 min. The mixture was then diluted with CH₂Cl₂ (100 mL) and washed successively with 5% aq Na₂S₂O₃ (50 mL), sat. aq NaHCO₃ (100 mL), and H₂O (100 mL). The organic phase was dried (Na₂SO₄) and concentrated under reduced pressure to give a crude product that was purified by chromatography [silica gel, hexane–EtOAc (4:1)] to give white solid; yield: 1.3 g (74%); mp 74–75 °C (EtOH); [α]_D²³+33 (*c* 1.0, CHCl₃).

IR (KBr): 3020, 2362, 1719, 1610, 1511, 1216, 1098, 1400, 761 cm⁻¹.

¹H NMR (500 MHz, CDCl₃): δ = 7.48–6.77 (m, 43 H, Ar-H), 5.87 (dd, *J* = 11.0, 3.5 Hz, 1 H, H-3_c), 5.58 (d, *J* = 3.0 Hz, 1 H, H-4_c), 5.55 (d, *J* = 8.0 Hz, 1 H, H-1_c), 5.28 (d, *J* = 2.5 Hz, 1 H, H-1_A), 4.95–4.58 (7d, *J* = 11.5 Hz each, 7 H, PhCH₂), 4.56 (t, *J* = 8.5 Hz each, 1 H, H-2_c), 4.51 (d, *J* = 11.5 Hz, 1 H, PhCH₂), 4.38–4.35 (m, 3 H, PhCH₂), 4.33 (br s, 1 H, H-1_B), 4.26–4.16 (m, 3 H, PhCH₂), 4.09–3.99 (m, 4 H, H-2_A, H-3_B, H-5_A, H-5_B), 3.98–3.92 (m, 2 H, H-3_A, H-4_A), 3.87–3.82 (m, 2 H, H-4_B, H-5_C), 3.75 (s, 3 H, OCH₃), 3.65 (dd, *J* = 10.0, 3.5 Hz, 1 H, H-2_B), 3.56–3.51 (m, 3 H, H-6_{aA}, H-6_{abc}), 3.42–3.28 (m, 2 H, H-6_{abb}), 3.24 (dd, *J* = 12.0, 5.5 Hz, 1 H, H-6_{bA}), 2.02, 1.84 (2 s, 6 H, 2 COCH₃).

¹³C NMR (125 MHz, CDCl₃): δ = 169.5, 169.4 (2 COCH₃), 167.7, 167.4 (PhthCO), 155.0–114.4 (m, Ar-C), 99.7 (C-1_c), 97.7 (C-1_A), 97.6 (C-1_B), 78.8 (2 C, C-5_A, C-5_B), 76.7 (C-3_A), 76.3 (C-2_B), 75.5 (C-3_B), 75.2 (C-2_A), 74.5 (PhCH₂), 74.4 (PhCH₂), 73.4 (PhCH₂), 73.3 (PhCH₂), 73.2 (PhCH₂), 73.1 (PhCH₂), 72.6 (PhCH₂), 71.6 (C-4_B), 69.9 (C-5_C), 69.0 (2 C, C-4_A, C-6_B), 67.9 (C-3_C), 67.4 (C-4_C), 67.2 (C-6_A), 67.1 (C-6_C), 55.5 (OCH₃), 51.9 (C-2_C), 20.6, 20.5 (2 COCH₃).

MALDI-MS: 1476.5 [M + Na]⁺.

Anal. Calcd for $C_{86}H_{87}NO_{20}$ (1453.58): C, 71.01; H, 6.03. Found: C, 70.84; H, 5.85.

4-Methoxyphenyl [4-O-Acetyl-6-O-benzyl-2-deoxy-2-(N-phthalimido)- β -D-galactopyranosyl]-(1 \rightarrow 3)-(2,4,6-tri-O-benzyl- α -D-galactopyranosyl)-(1 \rightarrow 6)-2,3,4-tri-O-benzyl- α -D-galactopyranoside (12)

A solution of compound **10** (1.2 g, 0.82 mmol) in 0.1 M methanolic NaOMe (20 mL) was stirred at r.t. for 1.5 h, then neutralized with Dowex 50W X8 (H⁺) resin, filtered, and concentrated under reduced pressure. A solution of the crude product **11**in DMF (5 mL) was treated with MeC(OEt)₃ (0.6 mL, 3.27 mmol) and TsOH (100 mg) and then stirred at r.t. for 2 h. H₂O (2 mL) was added and the mixture was stirred at r.t. for a further 30 min. The mixture was diluted with H₂O (100 mL) and extracted with CH₂Cl₂ (100 mL). The organic layer was washed successively with sat. aq NaHCO₃ (100 mL) and H₂O (100 mL), then dried (Na₂SO₄) and concentrated. The crude product was purified by chromatography [silica gel, hexane–EtOAc (4:1)] to give a colorless oil; yield: 1 g (86%, two steps); $[\alpha]_D^{23} + 35 (c 1.0, CHCl_3)$.

IR (neat): 2929, 2365, 1719, 1629, 1386, 1227, 1099, 768 cm⁻¹.

¹H NMR (500 MHz, CDCl₃): δ = 7.46–6.75 (m, 43 H, Ar-H), 5.48 (d, *J* = 8.5 Hz, 1 H, H-1_c), 5.44 (d, *J* = 3.0 Hz, 1 H, H-4_c), 5.27 (d, *J* = 2.0 Hz, 1 H, H-1_A), 4.93–4.49 (10 d, *J* = 11.5 Hz each, 10 H, PhCH₂), 4.41 (d, *J* = 2.5 Hz, 1 H, H-1_B), 4.36–4.32 (m, 4 H, H-2_c, PhCH₂), 4.22 (d, *J* = 11.5 Hz, 1 H, PhCH₂), 4.09–4.01 (m, 4 H, H-2_A, H-3_B, H-5_A, H-5_B), 3.98 (dd, *J* = 10.5, 3.0 Hz, 1 H, H-3_c), 3.96–3.87 (m, 3 H, H-3_A, H-4_A, H-4_B), 3.82–

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3.80 (m, 1 H, H-5_c), 3.72 (s, 3 H, OCH₃), 3.67 (dd, J = 10.5, 3.0 Hz, 1 H, H-2_B), 3.58–3.51 (m, 3 H, H-6_{abc}, H-6_{aA}), 3.42–3.35 (m, 1 H, H-6_{aB}), 3.34–3.26 (m, 2 H, H-6_{bA}, H-6_{bB}), 2.05 (s, 3 H, COCH₃).

¹³C NMR (125 MHz, CDCl₃): δ = 171.5 (COCH₃), 167.7, 167.4 (PhthCO), 155.0–114.4 (m, Ar-C), 99.9 (C-1_c), 97.8 (C-1_B), 97.7 (C-1_A), 78.8 (C-5_A), 78.7 (C-5_B), 76.7 (C-3_A), 76.3 (C-2_B), 75.6 (C-3_B), 75.2 (C-2_A), 74.6 (PhCH₂), 74.4 (PhCH₂), 73.5 (PhCH₂), 73.3 (PhCH₂), 73.2 (PhCH₂), 73.1 (PhCH₂), 72.8 (PhCH₂), 71.8 (C-4_B), 70.2 (C-4_A), 69.9 (C-3_c), 69.1 (C-4_c), 69.0 (C-6_B), 67.8 (C-6_A), 67.2 (C-5_c), 67.1 (C-6_c), 55.5 (OCH₃), 54.8 (C-2_c), 20.8 (COCH₃).

MALDI-MS: 1434.5 [M + Na]+.

Anal. Calcd for $C_{84}H_{85}NO_{19}$ (1411.57): C, 71.42; H, 6.07. Found: C, 71.25; H, 6.27.

MS-4Å (2 g) were added to a solution of compound **12** (900 mg, 0.64 mmol) and compound **5** (700 mg, 1.58 mmol) in anhyd 1:3 CH₂Cl₂–Et₂O (15 mL), and reaction mixture was cooled to 0 °C under argon. The cooled mixture was treated with NIS (360 g, 1.6 mmol) and TMSOTf (5 μ L) then stirred at 0 °C for 45 min. The mixture was then diluted with CH₂Cl₂ (100 mL) and washed successively with 5% aq Na₂S₂O₃ (50 mL), sat. aq NaHCO₃ (100 mL), and H₂O (100 mL), then dried (Na₂SO₄) and concentrated under reduced pressure. The crude product was purified by chromatography [silica gel, hexane–EtOAc (4:1)] to a white solid; yield: 750 mg (68%); mp 93–94 °C (EtOH); $[\alpha]_D^{23}$ +101 (*c* 1.0, CHCl₃).

IR (KBr): 2937, 1749, 1510, 1371, 1229, 1088, 1050, 827, 759 cm⁻¹.

¹H NMR (500 MHz, CDCl₃): δ = 7.49–6.76 (m, 48 H, Ar-H), 5.66 (d, *J* = 3.0 Hz, 1 H, H-4_c), 5.49 (d, *J* = 8.0 Hz, 1 H, H-1_c), 5.28 (d, *J* = 2.0 Hz, 1 H, H-1_A), 5.23 (s, 1 H, PhCH), 5.11 (d, *J* = 3.5 Hz, 1 H, H-1_D), 4.96 (d, *J* = 11.5 Hz, 1 H, PhCH₂), 4.86 (d, *J* = 11.5 Hz, 1 H, PhCH₂), 4.82 (dd, *J* = 10.0, 3.0 Hz, 1 H, H-3_D), 4.81–4.70 (m, 4 H, PhCH₂), 4.69–4.58 (m, 3 H, H-2_c, PhCH₂), 4.48 (d, *J* = 11.5 Hz, 1 H, PhCH₂), 4.40–4.23 (5 d, *J* = 11.5 Hz each, 5 H, PhCH₂), 4.09–4.04 (m, 5 H, H-2_A, H-3_B, H-4_D, H-5_A, H-5_B), 3.99–3.90 (m, 4 H, H-2_D, H-3_A, H-3_C, H-4_A), 3.87 (br s, 1 H, H-4_B), 3.82–3.80 (m, 1 H, H-5_C), 3.73 (s, 3 H, OCH₃), 3.70 (dd, *J* = 10.0, 3.0 Hz, 1 H, H-2_B), 3.60–3.50 (m, 4 H, H-6_{abc}, H-6_{abD}), 3.47–3.37 (m, 2 H, H-6_{aA}, H-6_{aB}), 3.18–3.17 (m, 1 H, H-5_D), 2.11, 2.06 (2 s, 6 H, 2 COCH₃).

¹³C NMR (125 MHz, CDCl₃): δ = 170.1, 169.9 (2 COCH₃), 167.7, 167.4 (PhthCO), 155.0–114.5 (m, Ar-C), 100.6 (PhCH), 99.8 (C-1_c), 97.8 (2 C, C-1_A, C-1_B), 97.0 (C-1_D), 78.8 (2 C, C-5_A, C-5_B), 76.4 (C-3_A), 76.3 (C-2_B), 75.5 (C-2_A), 75.2 (C-3_B), 74.5 (2 C, 2 PhCH₂), 73.6 (PhCH₂), 73.5 (PhCH₂), 73.3 (2 C, PhCH₂), 73.1 (C-4_A), 73.0 (C-4_D), 72.9 (PhCH₂), 72.6 (C-4_B), 69.8 (C-3_D), 69.7 (C-3_C), 69.2 (C-4_C), 69.1 (C-6_D), 68.3 (2 C, C-6_A, C-6_B), 67.0 (C-6_C), 66.4 (C-5_C), 62.9 (C-5_D), 57.2 (C-2_D), 55.5 (OCH₃), 53.4 (C-2_C), 20.9, 20.7 (2 COCH₃).

MALDI-MS: 1751.6 [M + Na]+.

Anal. Calcd for $C_{99}H_{100}N_4O_{24}$ (1728.67): C, 68.74; H, 5.83. Found: C, 68.60; H, 6.00.

4-Methoxyphenyl (2-Azido-4,6-O-benzylidene-2-deoxy- α -D-galactopyranosyl)-(1 \rightarrow 3)-[4-O-acetyl-6-O-benzyl-2-deoxy-2-(*N*-phthalimido)- β -D-galactopyranosyl]-(1 \rightarrow 3)-(2,4,6-tri-O-benzyl- α -D-galactopyranosyl)-(1 \rightarrow 6)-2,3,4-tri-O-benzyl- α -D-galactopyranoside (14)

A solution of compound **10** (700 mg, 0.40 mmol) in 0.1 M methanolic NaOMe (20 mL) was stirred at r.t. for 1.5 h. The mixture was then neutralized with Dowex 50W X8 (H⁺), filtered, and concentrated under reduced pressure. The crude product was passed through a short pad of silica gel with elution by hexane–EtOAc (2:1) to give a white solid; yield: 650 mg (95%); mp 95–96 °C (EtOH); $[\alpha]_D^{23}$ +61 (c 1.0, CHCl₃).

IR (KBr): 2927, 1746, 1508, 1456, 1369, 1233, 1109, 1055, 987, 826, 739 $\rm cm^{-1}.$

¹H NMR (500 MHz, CDCl₃): δ = 7.45–6.76 (m, 48 H, Ar-H), 5.66 (d, *J* = 3.0 Hz, 1 H, H-4_c), 5.47 (d, *J* = 8.5 Hz, 1 H, H-1_c), 5.27 (s, 1 H, PhCH), 5.26 (d, *J* = 2.5 Hz, 1 H, H-1_A), 5.09 (d, *J* = 3.0 Hz, 1 H, H-1_D), 4.95–4.69 (6 d, *J* = 11.5 Hz each, 6 H, PhCH₂), 4.65–4.58 (m, 4 H, H-2_c, PhCH₂), 4.45 (d, *J* = 11.5 Hz, 1 H, PhCH₂), 4.40 (d, *J* = 11.5 Hz, 1 H, PhCH₂), 4.38 (d, *J* = 3.0 Hz, 1 H, H-1_B), 4.35–4.22 (3 d, *J* = 11.5 Hz each, 3 H, PhCH₂), 4.36 (d, *J* = 3.0 Hz, 1 H, H-1_B), 4.35–4.22 (3 d, *J* = 11.5 Hz each, 3 H, PhCH₂), 4.07–4.00 (m, 4 H, H-2_A, H-3_B, H-5_A, H-5_B), 3.99–3.95 (m, 2 H, H-3_C, H-4_D), 3.92–3.84 (m, 2 H, H-3_A, H-3_D), 3.82–3.75 (m, 3 H, H-4_A, H-4_B, H-5_C), 3.73 (s, 3 H, OCH₃), 3.66 (d, *J* = 10.0, 3.0 Hz, 1 H, H-2_B), 3.62–3.60 (m, 1 H, H-6_{aD}), 3.58–3.48 (m, 4 H, H-2_D, H-6_{abc}, H-6_{bD}), 3.43–3.35 (m, 2 H, H-6_{aA}, H-6_{aB}), 3.32–3.25 (m, 2 H, H-6_{bA}, H-6_{bB}), 3.12–3.11 (m, 1 H, H-5_D), 2.10 (s, 3 H, COCH₃).

¹³C NMR (125 MHz, CDCl₃): δ = 171.2 (COCH₃), 167.7, 167.4 (PhthCO), 155.2–114.5 (m, Ar-C), 101.1 (PhCH), 99.8 (C-1_c), 97.7 (2 C, C-1_A, C-1_B), 96.6 (C-1_D), 78.8 (2 C, C-5_A, C-5_B), 76.7 (C-5_C), 76.3 (C-3_A), 75.5 (C-2_B), 75.1 (C-2_A), 74.9 (C-3_B), 74.5 (2 C, 2 PhCH₂), 73.6 (PhCH₂), 73.3 (PhCH₂), 73.1 (2 C, 2 PhCH₂), 72.5 (PhCH₂), 72.4 (C-4_A), 72.3 (C-4_D), 69.8 (C-4_B), 69.1 (C-3_D), 69.0 (C-6_D), 68.5 (C-6_A), 68.3 (C-6_B), 67.6 (C-3_C), 67.0 (C-6_C), 66.0 (C-4_C), 63.1 (C-5_D), 60.7 (C-2_D), 55.5 (OCH₃), 53.3 (C-2_C), 20.7 (COCH₃).

MALDI-MS: 1709.6 [M + Na]+.

Anal. Calcd for $C_{97}H_{98}N_4O_{23}$ (1686.66): C, 69.03; H, 5.85. Found: C, 68.84; H, 6.00.

4-Methoxyphenyl (2,3-O-Benzoyl-4,6-O-benzylidene- β -D-glucopy-ranosyl)-(1 \rightarrow 3)-(2-azido-4,6-O-benzylidene-2-deoxy- α -D-galactopyranosyl)-(1 \rightarrow 3)-[4-O-acetyl-6-O-benzyl-2-deoxy-2-(*N*-phthal-imido)- β -D-galactopyranosyl]-(1 \rightarrow 3)-(2,4,6-tri-O-benzyl- α -D-galactopyranosyl)-(1 \rightarrow 6)-2,3,4-tri-O-benzyl- α -D-galactopyranoside (15)

MS-4Å (500 mg) were added to a solution of compound **14** (600 mg, 0.36 mmol) and compound **6** (225 mg, 0.43 mmol) in anhyd CH₂Cl₂ (5 mL), and the mixture was cooled to – 20 °C under argon. NIS (100 mg, 0.44 mmol) and TMSOTf (3 μ L) were added and the mixture was stirred at –20 °C for 30 min. The mixture was then diluted with CH₂Cl₂ (50 mL) and washed successively with 5% aq Na₂S₂O₃ (25 mL), sat. aq NaHCO₃ (50 mL), and H₂O (50 mL). The organic phase was dried (Na₂SO₄) and concentrated under reduced pressure to give a crude product that was purified by chromatography [silica gel, hexane–EtOAc (4:1)] to give a white solid; yield: 550 mg (72%); mp 162–163 °C (EtOH); [α]_D²³ +72 (*c* 1.0, CHCl₃).

IR (KBr): 2932, 1776, 1748, 1720, 1509, 1388, 1229, 1107, 1081, 1052, 1029, 998, 827, 738, 722 $\rm cm^{-1}.$

¹H NMR (500 MHz, CDCl₃): δ = 7.89–6.70 (m, 63 H, Ar-H), 5.64 (t, *J* = 8.5 Hz, 1 H, H-2_E), 5.56 (d, *J* = 3.0 Hz, 1 H, H-4_C), 5.44 (d, *J* = 8.5 Hz, 1 H, H-1_C), 5.40 (t, *J* = 9.0 Hz, 1 H, H-3_E), 5.37 (s, 1 H, PhCH), 5.21 (d, *J* = 2.5 Hz, 1 H, H-1_A), 5.19 (s, 1 H, PhCH), 4.97 (d, *J* = 3.0 Hz, 1 H, H-1_D), 4.95

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(d, J = 8.5 Hz, 1 H, H-1_E), 4.91–4.52 (8 d, J = 11.5 Hz each, 8 H, PhCH₂), 4.51 (t, J = 8.5 Hz, 1 H, H-2_c), 4.41 (d, J = 11.5 Hz, 1 H, PhCH₂), 4.34 (d, J = 3.0 Hz, 1 H, H-1_B), 4.32–4.27 (m, 4 H, PhCH₂), 4.26–4.21 (m, 1 H, H-6_{aE}), 4.17 (d, J = 11.5 Hz, 1 H, PhCH₂), 4.02–3.96 (m, 5 H, H-2_A, H-3_B, H-5_A, H-5_B, H-5_E), 3.92–3.89 (m, 3 H, H-3_A, H-3_C, H-4_D), 3.85–3.80 (m, 2 H, H-3_D, H-4_A), 3.78–3.62 (m, 6 H, H-2_B, H-2_D, H-4_B, H-4_E, H-5_C, H-6_{bE}), 3.72 (s, 3 H, OCH₃), 3.55–3.46 (m, 4 H, H-6_{aA}, H-6_{abc}, H-6_{abc}), 3.43–3.40 (m, 1 H, H-6_{bD}), 3.38–3.32 (m, 1 H, H-6_{aB}), 3.30–3.20 (m, 2 H, H-6_{bA}, H-6_{bB}), 3.06–3.05 (m, 1 H, H-5_D), 1.95 (s, 3 H, COCH₃).

¹³C NMR (125 MHz, CDCl₃): δ = 171.0 (COCH₃), 167.7, 167.4 (PhthCO), 165.7, 165.3 (2 PhCO), 155.1–114.5 (m, Ar-C), 102.1 (C-1_E), 101.6 (PhCH), 100.4 (PhCH), 99.7 (C-1_C), 97.7 (C-1_A), 97.6 (C-1_B), 97.2 (C-1_D), 78.8 (2 C, C-5_A, C-5_B), 78.4 (C-5_E), 76.7 (C-5_C), 76.3 (C-3_A), 75.4 (2 C, C-2_A, C-2_B), 75.2 (C-3_B), 74.9 (C-3_E), 74.5 (2 C, 2 PhCH₂), 73.6 (PhCH₂), 73.1 (2 C, 2 PhCH₂), 72.7 (C-2_E), 72.5 (PhCH₂), 72.3 (2 C, C-4_A, C-4_E), 72.2 (C-4_D), 69.8 (C-4_B), 69.1 (C-3_D), 69.0 (C-6_D), 68.6 (C-6_E), 68.2 (C-6_A), 68.0 (C-6_B), 67.0 (C-6_C), 66.4 (2 C, C-3_C, C-4_C), 63.5 (C-5_D), 59.2 (C-2_D), 55.5 (OCH₃), 53.5 (C-2_C), 20.7 (COCH₃).

MALDI-MS: 2167.7 [M + Na]+.

Anal. Calcd for $C_{124}H_{120}N_4O_{30}\ (2144.80)$: C, 69.39; H, 5.64. Found: C, 69.23; H, 5.85.

4-Methoxyphenyl (β -D-glucopyranosyl)- $(1\rightarrow 3)$ -(2-acetamido-2-deoxy- α -D-galactopyranosyl)- $(1\rightarrow 3)$ -(2-acetamido-2-deoxy- β -D-galactopyranosyl)- $(1\rightarrow 3)$ - $(\alpha$ -D-galactopyranosyl)- $(1\rightarrow 6)$ - α -D-galactopyranoside (1)

NH₂NH₂·H₂O (0.3 mL) was added to a solution of compound 15 (500 mg, 0.23 mmol) in EtOH (15 mL), and the mixture was stirred at 80 °C for 10 h. The solvents were removed under reduced pressure, the crude product was dissolved in Ac₂O (2 mL) and pyridine (2 mL), and the solution was kept at r.t. for 1 h. The solvents were removed under reduced pressure to give an acetylated product that was passed through a short pad of silica gel with EtOAc (50 mL) as eluent. A solution of the purified acetylated in MeOH (10 mL) was treated with 20% $Pd(OH)_2/C(100 \text{ mg})$ under a positive pressure of H₂ with stirring at r.t. for 15 h. The mixture was then filtered through a bed of Celite that was washed MeOH (30 mL), and the filtrate was concentrated to half of its original volume. Ac₂O (1 mL) was added and the mixture was stirred at r.t. for 1 h. The solvents were removed under reduced pressure and a solution of the crude product in 0.1 M methanolic NaOMe (10 mL) was stirred at r.t. for 1.5 h. The mixture was neutralized with Dowex 50W X8 (H⁺), filtered, and concentrated to give the crude product that was purified by chromatography [Sephadex LH-20 gel, MeOH-H₂O (3:1)] to give **1** as a white powder; 125 mg (54%); $[\alpha]_{D}^{23}$ +99 (c 1.0, H₂O).

IR (KBr): 3020, 2928, 1744, 1377, 1220, 1069, 764 cm⁻¹.

¹H NMR (500 MHz, D₂O): δ = 7.02, 6.86 (2 d, *J* = 9.0 Hz each, 4 H, Ar-H), 5.45 (d, *J* = 4.0 Hz, 1 H, H-1_A), 4.95 (d, *J* = 3.5 Hz, 1 H, H-1_D), 4.70 (d, *J* = 3.5 Hz, 1 H, H-1_B), 4.51 (d, *J* = 8.0 Hz, 1 H, H-1_C), 4.37 (d, *J* = 7.5 Hz, 1 H, H-1_E), 4.28–4.24 (m, 1 H, H-2_D), 4.16–4.09 (m, 2 H, H-3_A, H-4_B), 4.02 (br s, 1 H, H-4_C), 3.98–3.90 (m, 5 H, H-2_A, H-2_C, H-3_C, H-4_A, H-4_D), 3.86–3.81 (m, 1 H, H-3_D), 3.77–3.56 (m, 13 H, H-2_B, H-3_B, H-3_E, H-5_B, H-6_{abA}, H-6_{abC}, H-6_{abC}, H-6_{aE}), 3.68 (s, 3 H, OCH₃), 3.55–3.47 (m, 3 H, H-5_A, H-5_C, H-6_{bE}), 3.35 (t, *J* = 8.5 Hz, 1 H, H-4_E), 3.31–3.27 (m, 2 H, H-5_D, H-5_E), 3.14 (t, *J* = 9.0 Hz, 1 H, H-2_E), 1.90, 1.89 (2 s, 6 H, 2 CO-CH₃).

¹³C NMR (125 MHz, D₂O): δ = 174.8, 174.6 (2 COCH₃), 154.7–115.0 (m, Ar-C), 104.3 (C-1_E), 102.6 (C-1_C), 98.2 (2 C, C-1_A, C-1_B), 93.8 (C-1_D), 79.5 (C-5_A), 77.4 (C-5_B), 75.8 (2 C, C-5_C, C-5_E), 75.6 (2 C, C-3_D, C-4_E), 74.7 (2 C, C-3_E, C-3_E), 72.9 (2 C, C-2_E, C-5_D), 71.1 (C-4_D), 70.0 (2 C, C-3_C, C-3_C), 71.1 (C-4_D), 70.0 (2 C, C-3_C), 72.6 (2 C, C-3_C), 73.6 (2 C, C-3_C), 73.6 (2 C, C-3_C), 73.6 (2 C, C-3_C), 74.7 (2 C, C-3_E), 72.9 (2 C, C-2_E, C-5_D), 71.1 (C-4_D), 70.0 (2 C, C-3_C), 73.6 (2 C, C-3_C), 73.6

 $\begin{array}{l} {\rm C-4_B},\ 69.0\ ({\rm C-3_A}),\ 68.6\ ({\rm C-4_A}),\ 68.0\ ({\rm C-2_B}),\ 67.5\ ({\rm C-2_A}),\ 67.1\ ({\rm C-6_A}),\\ {\rm 63.5\ ({\rm C-4_C}),\ 61.0\ ({\rm C-6_D}),\ 60.7\ (2\ C,\ {\rm C-6_E},\ {\rm -6.5\ (C-6_E)},\ 56.1\ ({\rm OCH_3}),\\ {\rm 51.0\ ({\rm C-2_C}),\ 48.1\ ({\rm C-2_D}),\ 22.5,\ 22.3\ (2\ {\rm COCH_3}). \end{array}$

ESI-MS: 1039.3 [M + Na]+.

Anal. Calcd for $C_{41}H_{64}N_2O_{27}$ (1016.37): C, 48.42; H, 6.34. Found: C, 48.24; H, 6.50.

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Supporting Information

Supporting information for this article is available online at http://dx.doi.org/10.1055/s-0034-1378911.

References

- (1) Thapar, N.; Sanderson, I. R. Lancet 2004, 363, 641.
- (2) Ashbolt, N. J. Toxicology 2004, 198, 229.
- (3) Guerrant, R. L.; Hughes, J. M.; Lima, N. L.; Crane, J. *Rev. Infect. Dis.* **1990**, *12*, S41.
- (4) Nataro, J. P.; Kaper, J. B. Clin. Microbiol. Rev. 1998, 11, 142.
- (5) McVeigh, A.; Fasano, A.; Scott, D. A.; Jelacic, S.; Moseley, S. L.; Robertson, D. C.; Savarino, S. J. *Infect. Immun.* **2000**, *68*, 5710.
- (6) Hussein, H. S.; Sakuma, T. J. Dairy Sci. 2005, 88, 450.
- (7) Zhou, Z.; Ogasawara, J.; Nishikawa, Y.; Seto, Y.; Helander, A.; Hase, A.; Iritani, N.; Nakamura, H.; Arikawa, K.; Kai, A.; Kamata, Y.; Hoshi, H.; Haruki, K. *Epidemiol. Infect.* **2002**, *128*, 363.
- (8) Ali, T.; Weintraub, A.; Widmalm, G. Carbohydr. Res. 2007, 342, 274.
- (9) Alanis, A. J. Arch. Med. Res. 2005, 36, 697.
- (10) Yan, S.; Ding, N.; Zhang, W.; Wang, P.; Li, Y.; Li, M. *Carbohydr*. *Res.* **2012**, *354*, 6.
- (11) Das, S. K.; Ghosh, R.; Roy, N. J. Carbohydr. Chem. 1993, 12, 693.
- (12) Santra, A.; Ghosh, T.; Misra, A. K. *Tetrahedron: Asymmetry* **2012**, 23, 1385.
- (13) Ziegler, T.; Eckhardt, E.; Strayle, J.; Herzog, H. Carbohydr. Res. **1994**, 253, 167.
- (14) Panchadhayee, R.; Misra, A. K. *Tetrahedron: Asymmetry* **2009**, 20, 1550.
- (15) Panchadhayee, R.; Misra, A. K. Synlett 2010, 1193.
- (16) Hudson, C. S.; Dale, I. K. J. Am. Chem. Soc. 1915, 37, 1264.
- (17) Konradsson, P.; Udodong, U. E.; Fraser-Reid, B. *Tetrahedron Lett.* **1990**, *31*, 4313.
- (18) Veeneman, G. H.; van Leeuwen, S. H.; van Boom, J. H. *Tetrahedron Lett.* **1990**, *31*, 1331.
- (19) Ogawa, T.; Yamamoto, H. Agric. Biol. Chem. 1985, 49, 475.
- (20) Zemplén, G. Ber. Dtsch. Chem. Ges. 1926, 59, 1254.
- (21) Field, R. A.; Otter, A.; Fu, W.; Hindsgaul, O. *Carbohydr. Res.* **1995**, 276, 347.
- (22) Kumar, R.; Maulik, P. R.; Misra, A. K. *Glycoconjugate J.* **2008**, *25*, 511.
- (23) Lee, H.-H.; Schwartz, D. A.; Harris, J. F.; Carver, J. P.; Krepinsky, J. J. Can. J. Chem. **1986**, 64, 1912.
- (24) Pearlman, W. M. Tetrahedron Lett. 1967, 8, 1663.