

Familial Hemophagocytic Lymphohistiocytosis Presenting as Hydrops Fetalis

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Am J Perinatol Rep 2015;5:e22–e24.

Abstract

Keywords

- ▶ familial hemophagocytic lymphohistiocytosis
- ▶ hydrops fetalis
- ▶ hepatosplenomegaly
- ▶ cytopenia
- ▶ immunosuppressive chemotherapy

Background Familial hemophagocytic lymphohistiocytosis (FLH) is an autosomal recessive disorder of immune regulation that leads to a hyperinflammatory syndrome. Fetal onset FHL is extremely rare and is considered to be the most severe form of FHL.

Case We report a preterm case of FHL that presented as hydrops fetalis. The infant was treated with a chemotherapy regimen based on the HLH-2004 protocol from the third day of life. However, he had persistent cytopenia and died on the 18th day of life due to bacteremia. The detection of defective perforin expression in the patient's natural killer cells and mutations in the *PRF1* gene resulted in a molecular diagnosis of FHL.

Conclusion We suggest that early diagnosis and the development of an appropriate immunosuppressive strategy that can induce and maintain remission until hematopoietic stem cell transplantation can be performed are required to improve the outcomes of fetal onset FHL.

Hemophagocytic lymphohistiocytosis (HLH) is caused by the prolonged and excessive activation of T-cells and macrophages.¹ HLH encompasses several entities; that is, primary familial HLH (FHL), which has an estimated incidence of 1 in 50,000 births, and a secondary form of HLH associated with infection, malignancies, and rheumatological disorders.² FHL is a rare autosomal recessive disorder in which four different gene defects have been identified: *PRF1* (FHL2), *UNC13D* (FHL3), *STX11* (FHL4), and *STXBP2* (FHL5).^{1,3}

Most children with FHL are born healthy and become ill in the first 2 to 6 months of life.⁴ Neonatal presentation is rare,⁵ and the diagnosis of the condition is frequently delayed. In fact, it is often missed completely or only diagnosed at autopsy.⁶ In addition, fetal onset FHL is extremely rare and is considered to be the most severe form of FHL.^{4,6} Here, we report a preterm case of fetal onset FHL2.

Case Report

Our patient was a male newborn infant with a gestational age of 36^{5/7} weeks and a body weight of 2,665 g. He was the third child of unrelated healthy parents and had healthy brothers. Maternal screening tests for toxoplasmosis, rubella, cytomegalovirus, human immunodeficiency virus, and human parvovirus produced negative results. The patient was delivered by emergency cesarean section because of fetal ascites and a nonreassuring fetal status. His Apgar scores were 4 and 7 at 1 and 5 minutes, respectively, and he was intubated and placed on mechanical ventilation due to respiratory distress. In addition, he had diffusely distributed small purple and red spots on his trunk and extremities (→Fig. 1). His liver and spleen had descended 2 and 1 cm below the right and left costal margins, respectively. A blood test produced the

received
October 9, 2014
accepted after revision
November 25, 2014
published online
March 4, 2015

DOI <http://dx.doi.org/10.1055/s-0034-1544110>.
ISSN 2157-6998.

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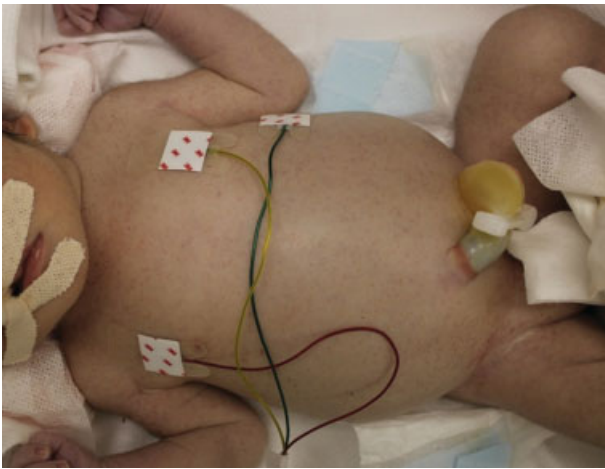


Fig. 1 At birth, diffusely distributed small purple and red spots were present on the infant's trunk and extremities.

following findings: hemoglobin, 10.1 g/dL; white blood cell count, $2.5 \times 10^9/L$, including 6.0% immature neutrophils, 12.0% segmented neutrophils, 15.5% monocytes, 65.0% lymphocytes, and 0.5% atypical lymphocytes; platelet count, $14 \times 10^9/L$; total protein, 4.2 g/dL; total bilirubin, 3.49 mg/dL; aspartate aminotransferase (AST), 144 IU/L; alanine aminotransferase (ALT), 31 IU/L; lactate dehydrogenase (LDH), 917 IU/L; ferritin, 4,176 ng/mL; and C-reactive protein, 2.23 mg/dL. An abdominal ultrasound examination detected hepatosplenomegaly and ascites. We diagnosed the patient with a congenital bacterial or viral infection, and he was treated with antibiotics, gamma globulins, and an exchange transfusion. However, we could not specify the cause of his infection, and his cytopenia did not improve despite repeated transfusions of platelets, red cells, and fresh frozen plasma.

The serum concentrations of cytokines were measured before the exchange transfusion. The patient's serum interleukin (IL)-6, IL-8, and tumor necrosis factor α levels were 133.0 pg/mL (normal: < 4 pg/mL), 65.5 pg/mL (normal: < 2 pg/mL), and 40.5 pg/mL (normal: 0.6–2.8 pg/mL), respectively. Laboratory tests performed on blood samples obtained on the first day of life showed significantly elevated serum levels of AST (7,173 IU/L), ALT (1,233 IU/L), LDH (14,730 IU/L), ferritin (115,360 ng/mL), and soluble IL-2 receptor (5,400 IU/mL, normal: 145–519 U/mL). In addition, the patient's serum triglyceride and β_2 -microglobulin levels were 107 mg/dL (normal: 40–150 mg/dL) and 9.2 mg/L (normal: 0.68–1.65 mg/L), respectively. The patient's natural killer (NK) cell function was markedly reduced (0%, normal: 18–40%). On the third day, a bone marrow aspiration biopsy detected hemophagocytosis together with increased numbers of macrophages and histiocytes, which were consistent with HLH.

We treated the patient from the third day of life with a chemotherapy regimen based on the HLH-2004 protocol.⁷ We intravenously administered dexamethasone (0.5 mg/kg/d) and cyclosporine A (2 mg/kg/d), but the patient's condition deteriorated. In addition, etoposide was administered intravenously at a dose of 100 mg/m² twice a week. Although the patient's edema improved slightly, leukopenia ($< 0.5 \times 10^9/L$)

and thrombocytopenia ($< 50 \times 10^9/L$) persisted. On the 16th day of life, the infant developed bacteremia because of *Pseudomonas aeruginosa*, and he died on the 18th day of life. Flow cytometric analysis revealed an absence of perforin expression in his NK cells (**Fig. 2**), and postmortem DNA analysis identified mutations in the *PRF1* gene (c.1A $>$ G and c.1090_1091delCT).

Discussion

Neonatal HLH has been described in the previous studies of pediatric HLH^{5,8}; however, the characteristics of neonatal HLH have not been well defined. The clinical findings of neonatal HLH are very similar to those of congenital infections. In a nationwide Japanese survey, Suzuki et al indicated that neonates with hepatomegaly, thrombocytopenia, and elevated LDH levels should be examined further for potential HLH.⁶ Maruyama et al reviewed six case reports involving seven premature infants with congenital HLH.⁹ Four of the patients were diagnosed with FHL; however, a definitive diagnosis of FHL was not obtained in the other three cases. Ascites and hepatosplenomegaly were detected in almost all the cases, and hydrops fetalis was also seen in four cases. Our patient presented with hydrops fetalis, ascites, and hepatosplenomegaly. In addition, cytopenia, hyperferritinemia, and elevated LDH levels were facilitated the early diagnosis of neonatal HLH in this case.

Previous reports have shown that untreated FHL is uniformly fatal within 2 years of diagnosis.⁵ Isaacs reviewed the cases of 72 fetuses and neonates with HLH and found that they exhibited an overall survival rate of 26%.⁸ Among the latter patients, only 3 of the 34 (9%) patients who were diagnosed with FHL had survived compared with 10 of the 17 (59%) patients with infection-associated hemophagocytic syndrome. Immunosuppressive chemotherapy results in the control of FHL in some cases; however, remission is rarely sustained. Most patients suffer an early death unless they undergo hematopoietic stem cell transplantation (HSCT), which appears to be the only curative approach.⁸ Our patient died on the 18th day of life despite early diagnosis and immunosuppressive chemotherapy. Suzuki et al described 20 cases of

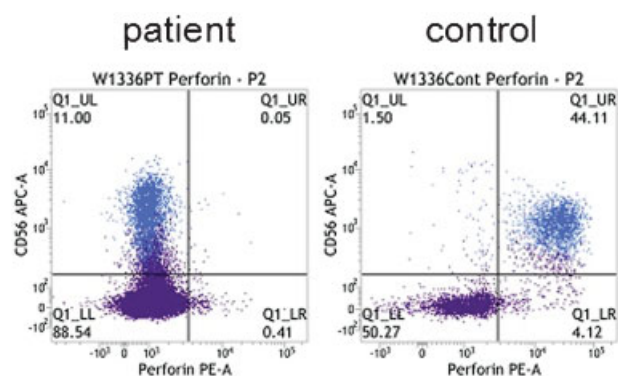


Fig. 2 Fluorescence-activated cytometric analysis of perforin expression in CD3-peripheral lymphocytes obtained from the patient and a healthy control. Perforin expression was markedly reduced in the patient's CD56+ cells.

neonatal HLH that occurred in Japan during a 10-year period.⁶ Of the three patients with genetically confirmed FHL, two died without receiving HSCT on the 29th and 114th day of life because of progression of the disease, respectively, whereas one patient who was treated with HSCT survived. Although the HLH-2004 protocol proved effective inducing remission in FHL, it may be insufficient for the treatment of neonatal onset FHL. Other less myelotoxic immunotherapies, such as antithymocyte globulin, could be combined with pre-HSCT therapy. A study involving a mouse model of FHL suggested that targeted immunotherapy might play a role in treating FHL, as improved recovery and survival were observed in mice treated with anti-interferon-gamma monoclonal antibodies.¹⁰ Recently, a humanized monoclonal anti-CD 52 antibody, CAMPATH-1H, has been used to treat refractory FHL.¹¹ The deletion of T-cells and B-cells with CAMPATH-1H could be considered as an optional treatment for neonatal onset FHL. Also, mesenchymal stem cells, which are known to display multipotency and robust anti-inflammatory and regenerative properties, have been used to treat a range of immune-mediated conditions including graft versus host disease.¹² We hope that the clinical application of these novel immunosuppressive strategies will help induce and maintain remission in FHL neonates until HSCT can be performed.

Conflicts of Interest

The authors have no conflicts of interest to declare.

Acknowledgments

We thank the patient's family for their kind cooperation and for giving the permission to publish photographs of the patient. We also thank Dr. Keiko Wada, Dr. Hitomi Sakai, and Dr. Aiko Kozaki for their help with the clinical management of our patient.

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