EFSUMB Guidelines on Interventional Ultrasound (INVUS), Part IV
EUS-guided interventions: General Aspects and EUS-guided Sampling (short version)

EFSUMB Leitlinien interventioneller Ultraschall (INVUS), Teil IV
Endosonografisch gestützte Interventionen: allgemeine Aspekte und endosonografisch gestützte Materialgewinnung (Kurzversion)

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- endoscopic ultrasound
- EUS-guided sampling
- EUS-guided fine-needle aspiration
- rapid onsite cytopathological evaluation
- cytopathological reporting
- quality indicators
- adverse events

Abstract
The fourth part of the European Federation of Societies for Ultrasound in Medicine and Biology (EFSUMB) Guidelines on Interventional Ultrasound describes general aspects of endoscopic ultrasound-guided diagnostic and therapeutic interventions and assesses the evidence for endoscopic ultrasound-guided sampling. Endoscopic ultrasound combines the most advanced high-resolution ultrasound imaging of lesions within the wall and in the vicinity of the gastrointestinal tract and safe and effective fine needle-based tissue acquisition from these lesions. The guideline addresses the indications, contraindications, techniques, adverse events, training and clinical impact of EUS-guided sampling. Advantages and drawbacks are weighed in comparison with image-guided percutaneous biopsy. Based on the most current evidence, clinical practice recommendations are given for crucial preconditions and steps of EUS-guided sampling as well as for safe performance. Additionally, the guideline deals with the principles and reliability of cytopathological reporting in endoscopic ultrasound-guided sampling (short version; the long version is published online).

Zusammenfassung

Introduction: Diagnostic and therapeutic EUS-guided interventions
This is the first of two guidelines (parts IV and V) within the framework of the European Federation of Societies for Ultrasound in Medicine and Biology (EFSUMB) Guidelines on Interventional Ultrasound describing endoscopic ultrasound (EUS)-guided diagnostic and therapeutic interventions.

Part IV deals with the indications and clinical impact of EUS-guided sampling1 and gives evidence-based recommendations for the safe and efficient performance of this technique based on the available evidence at the time of manuscript prepara-
Regulation. It is complemented by a guideline on EUS-guided therapeutic interventions (part V) [1], Methods of guideline development are described in the introduction to the EFSUMB Guidelines on Interventional Ultrasound (INVUS) [2]. Levels of evidence (LoE) and Grades of Recommendations (GoR) have been assigned according to the Oxford Centre for Evidence-based Medicine criteria (March 2009 edition) [http://www.cebm.net/oxford-centre-evidence-based-medicine-levels-evidence-march-2009].

General considerations

Endoscopic ultrasound (EUS; synonym: endosonography)² represents an interdisciplinary high-performance imaging method which combines endoscopy and ultrasound by using special transducers mounted at the tip of the endoscope [3 – 9]. With the introduction of EUS-guided fine-needle aspiration (EUS-FNA)³ in 1992 [10], it became possible to obtain a definite cytopathological or histopathological diagnosis of mediastinal, abdominal, retroperitoneal and pelvic lesions, establishing EUS-FNA as a crucial procedure. Further advances in 1996, allowed EUS-guided celiac plexus neurolysis [11], EUS-guided drainage of GI tract-adjacent fluid collections and pancreatic pseudocysts [12]. A first experience with endobronchial ultrasound-guided transbronchial fine-needle aspiration (EBUS-TBNA)⁴ was reported in 2003 [13].

Equipment and setup

Linear echoendoscopes have an ultrasound examination field in the longitudinal axis of the echoendoscope, allowing performance of needle-based diagnostic and therapeutic interventions through real-time visualization [14]. A wide variety of accessory equipment for EUS-guided interventions is commercially available [15]. For endobronchial ultrasound (EBUS) specifically designed linear echoendoscopes based on flexible bronchoscopes are available [16]. A therapeutic echoendoscope with a forward-viewing 90-degree curved array at the front of the scope has a potential advantage of allowing interventions with an axial application of force during needle insertion and stenting, with comparable imaging quality and equal performance in EUS-guided drainage of pancreatic pseudocysts being reported [17 – 19]. Both rigid (linear and curved-array) ultrasound probes with a biopsy channel as well as flexible linear and forward-viewing echoendoscopes can be used for transrectal EUS-guided interventions [20 – 27].

**Recommendation 1**

Linear echoendoscopes are indispensable for EUS-guided sampling and injection treatments (LoE 5, GoR D). Strong consensus (100%).

**Recommendation 2**

For one-step EUS-guided drainage procedures, a large channel therapeutic linear echoendoscope is recommended (LoE 5, GoR D). Strong consensus (100%).

**Patient preparation and monitoring**

Patient preparation is identical to flexible endoscopy, with fasting for 6 hours, detailed informed consent, management of anticoagulation and antiplatelet therapy, indication for prophylactic antibiotics, and need for sedation [28]. The American Society of Gastrointestinal Endoscopy (ASGE) has defined several evidence-based preprocedural quality indicators [29].

EUS (including EBUS) can be safely performed using conscious sedation (midazolam) perhaps combined with an opioid (fentanyl or pethidine) [30 – 32]. Alternatively, propofol sedation is a more efficient approach, with better patient and operator satisfaction and a good safety profile, especially for lengthy interventions like therapeutic EUS [31, 33 – 41]. Endoscopist- and nurse-administered propofol sedation has been shown to be safe and effective in EUS and EUS-guided interventions (including EBUS-TBNA) in both average-risk and high-risk patients [36, 37, 42 – 44]. National legal restrictions in many countries do not permit non-anesthesiologist administration of propofol [45, 46].

**Recommendation 3**

Carefully monitored conscious or deep sedation should be routinely offered to all patients undergoing EUS-guided interventions (LoE 1b, GoR B). Strong consensus (100%).

**Recommendation 4**

Controlled propofol sedation should be used with due consideration of national legal regulations, and experience and training of the medical personnel (LoE 5, GoR D). Strong consensus (100%).

**Platforms for EUS-guided diagnostic and therapeutic interventions**

Needles for EUS-guided sampling are available with a size of 25 – 19 Gauge (G) [14, 15, 47 – 49]. “Histology needles” include: trucut needles, needles with side fenestration and reverse bevel technology [50 – 57] or with shark mouth-like needle tip geometry. Therapeutic EUS-guided interventions commence with puncture of the targeted lesion for initial access, followed by therapeutic injection or a guidewire-assisted drainage procedure. 19 G needles are normally used, allowing passage of 0.035 inch guidewires.

**Indications and contraindications**

EUS has evolved as a diagnostic and therapeutic procedure with substantial clinical impact, altering management in a number of patients [3, 29, 57 – 63]. Appropriate (evidence-based) indications are listed in Table 1 [3, 7 – 9, 29, 57, 64 – 69]. Absolute contraindications of EUS are similar with those of conventional advanced endoscopy procedures. Specific contraindications of EUS-guided sampling and therapeutic interventions are related to unacceptable risks of bleeding, infection, and perforation (Table 2; section 5) [8, 9, 14, 66, 70, 71], which should be assessed prior to the intervention on an individual basis weighing procedural risk vs. clinical impact.

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² Term is explained in the addendum on terminology (online version only).
³ Term is explained in the addendum on terminology (online version only).
⁴ Term is explained in the addendum on terminology (online version only).
Table 1  Established indications of endoscopic ultrasound.

<table>
<thead>
<tr>
<th>diagnostic evaluation (including EUS-guided sampling)</th>
<th>pancreatico-biliary disorders</th>
<th>oesophagogastrroduodenal disorders</th>
<th>mediastinal diseases</th>
<th>rectal diseases</th>
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</thead>
<tbody>
<tr>
<td>established indications of endoscopic ultrasound</td>
<td>obstructive jaundice/biliary stricture of unknown aetiology</td>
<td>subepithelial tumours</td>
<td>primary central lung/mediastinal tumours</td>
<td>fistulas, abscesses, extraluminal tumours, subepithelial tumours</td>
</tr>
<tr>
<td>EUS-guided therapeutic interventions</td>
<td>pancreatico-biliary cancers</td>
<td>drainage procedures</td>
<td>injection treatment</td>
<td>other interventions</td>
</tr>
<tr>
<td>T (tumour) and N (nodal) stage, M (metastatic) stage (celiac trunk lymph nodes, left adrenal, left liver lobe, ascites, etc.)</td>
<td>T and N stage (EUS and EBUS), M stage (abdominal lymph nodes, left adrenal, left liver lobe)</td>
<td>transmural drainage of (per-)pancreatic and peri-intestinal fluid collections (pseudocysts, walled-off pancreatic necroses, abscesses, postoperative fluid collections)</td>
<td>celiac plexus/ganglia neurolysis and block</td>
<td>placement of fiducials</td>
</tr>
<tr>
<td>pancreatic duct drainage</td>
<td>bile duct and gallbladder drainage</td>
<td>pancreatic duct drainage</td>
<td>glue or coil embolization of bleeding varices, visceral pseudoaneurysms, and other vascular lesions</td>
<td>ethanol ablation of small pancreatic insulinomas</td>
</tr>
<tr>
<td>perianastomotic masses</td>
<td>mediastinal lymphadenopathy</td>
<td>pancreatic duct drainage</td>
<td>ethanol ablation of small pancreatic insulinomas</td>
<td>other interventions</td>
</tr>
</tbody>
</table>

Table 2  Indications and contraindications of EUS-guided sampling.

<table>
<thead>
<tr>
<th>frequent/ established indications</th>
<th>less frequent/ emerging indications</th>
<th>contraindications</th>
</tr>
</thead>
<tbody>
<tr>
<td>pancreatic solid lesions</td>
<td>retroperitoneal masses</td>
<td>absolute</td>
</tr>
<tr>
<td>pancreatic cystic lesions</td>
<td>mediastinal masses</td>
<td>no impact on patient management</td>
</tr>
<tr>
<td>lymph nodes</td>
<td>perianastomotic masses</td>
<td>lack of informed consent</td>
</tr>
<tr>
<td>subepithelial tumours</td>
<td>adrenal gland masses</td>
<td>mediastinal cysts (risk of infection)</td>
</tr>
<tr>
<td>liver masses</td>
<td>relative</td>
<td></td>
</tr>
<tr>
<td>ascites and peritoneal nodules</td>
<td>severe coagulopathy</td>
<td></td>
</tr>
<tr>
<td>gastrointestinal wall thickening</td>
<td>continued oral anticoagulation</td>
<td></td>
</tr>
<tr>
<td>bile duct strictures/ lesions</td>
<td>continued treatment with ADP antagonists (clopidogrel, prasugrel, ticagrelor)</td>
<td></td>
</tr>
<tr>
<td>kidney masses</td>
<td>large vessels in the expected needle track</td>
<td></td>
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<tr>
<td>focal splenic lesions</td>
<td></td>
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</tbody>
</table>

Recommendation 5

As EUS is an advanced invasive procedure, it requires a proper indication, assessment of individual risks and contraindications, detailed informed consent, considerate management of antithrombotic therapy and antibiotic prophylaxis, and a pre-interventional team time-out\(^5\) (LoE 5, GoR D). Strong consensus (100%).

Education and training in endoscopic ultrasound

Preconditions for performing EUS

EUS has evolved into an advanced endoscopic procedure requiring structured training as supported by the ASGE core curriculum for EUS [72]. Certain conditions need to be attained prior to commencing an EUS program; state-of-the-art EUS equipment and accessories should be in place, previous EUS experience and training (>12 months) with proficiency in basic EUS should be available, the presence of a multidisciplinary team with expertise in gastroenterology, pulmonary medicine, oncology, surgery, imaging and pathology is desired.

Methods and models for training

Various approaches for structured training have been described. Support includes EUS textbooks and atlases, DVDs and online resources [7–9, 73–78]. Computer-based simulator training, EUS-guided sampling and therapeutic intervention training with animal models and supervised hands-on may facilitate improved learning for clinical application and may minimize false-negative sampling in patients [73, 76, 79–93]. Objective assessment tools with rating scales for various steps of EUS-guided sampling have been developed to measure competency for mediastinal staging of NSCLC [83, 85, 90, 94–96].

Learning curve and minimum number of supervised procedures

Reports on the skill acquisition of individuals for EUS [97, 98] and for EUS-FNA of solid pancreatic lesions [99, 100] challenge the ASGE recommendations that comprehensive competence in all procedures requires at least 150 supervised cases, including 50 EUS-FNA [101]. Also for EBUS-TBNA, studies show significant variation in learning curves [96] with an acceptable 90% accuracy achieved after performing 50–80 procedures [102–105].

Recommendation 6

Education and training for EUS procedures should be performed systematically in a standardized fashion. Various learning methods should be used, including textbooks and on-line resources, and EUS phantoms, before performing hands-on human cases (LoE 5, GoR D). Strong consensus (100%).

Recommendation 7

Standardized criteria and assessment tools should be developed and applied for assessing skills of individual trainees before credentialing competency (LoE 1b, GoR B). Strong consensus (100%).

\(^5\) Term is explained in the addendum on terminology (online version only).
Quality indicators and benchmarking

Useful quality indicators for EUS-guided sampling may be:

- Percentage of appropriate indications for performing EUS-guided sampling due to guidelines (Table 2) [29]
- Prospective documentation of adverse events after EUS-guided sampling (performance target: > 98 %) [29]
- Incidence of adverse events after EUS-guided sampling (performance targets: acute pancreatitis < 2 %, clinically significant bleeding < 1 %) [29, 48, 66]
- Frequency of algorithmic EUS-guided sampling of both suspected metastatic disease and the primary tumour in cases, in which results of EUS staging would impact further clinical management (performance target: > 98 %) [29]
- The yield of malignant diagnoses in EUS-guided sampling of solid pancreatic masses and the sensitivity for diagnosis of pancreatic cancer (performance targets: ≥ 70 and 85 %, respectively) [29, 48, 57, 66, 106]
- The percentage of adequate samples (performance target: ≥ 85 %) [29, 48, 66] and
- The frequency of inconclusive cytopathological diagnoses (atypical, suspicious: ≤ 10 %) [48, 66, 107].

Particular quality parameters for EUS-guided therapeutic interventions have not yet been established, but should be derived from those for EUS-guided sampling and advanced endoscopic and interventional procedures [29, 108].

EUS-guided sampling: Indications and clinical impact

General indications of EUS-guided sampling

EUS has high accuracy for the diagnosis and staging of benign and malignant conditions within and outside the gastrointestinal tract (Table 2) [61, 62]. EUS-guided sampling relies on its ability to obtain specimens with either fine-needle aspiration (FNA) or fine-needle biopsy (FNB) [57, 66].

Several prospective and retrospective studies and meta-analyses have substantiated the key importance of EUS-guided sampling particularly in suspicious nodal disease for staging, guiding treatment, and predicting outcome in non-small cell lung cancer (NSCLC) [109 – 114], various extrathoracic malignancies [115, 116], upper gastrointestinal cancer [117, 118], rectal cancer [23], as well as proving recurrence of malignancy [20, 119 – 122]. Speci-
cificity of cytological diagnosis of malignant PCL is adequate, but the sensitivity is low (88–97 % and 51–65 %, respectively) [150 – 153]. The combination of molecular analysis and CEA or cytology has a better performance for the diagnosis of neoplastic mucinous PCL than either of the individual tests [154 – 156].

**Recommendation 14**

Biochemical, cytological, and molecular analysis of EUS aspirates from pancreatic cystic lesions may facilitate differentiation between mucinous and non-mucinous cysts and evaluation of the malignancy risk (LoE 2a, GoR B). Strong consensus (100 %).

**Biliary cancer**

Several studies report successful EUS-guided sampling of gallbladder wall lesions [157 – 161], extrahepatic cholangiocarcinoma and indeterminate biliary strictures [157, 162 – 173], as well as tumours of the papilla of Vater [174], but with wide variation in sensitivity (43 – 100 %) [175]. EUS-guided sampling proved to be successful following failed or negative ERCP-guided brushing or biopsy [163, 165 – 169, 175]. Comparison of EUS-guided sampling with that of ERCP-guided techniques revealed a better outcome for EUS in gallbladder cancer, biliary strictures produced by pancreatic cancer rather than cholangiocarcinoma, and in EUS-visible masses, with a combined approach considered clinically appropriate [159, 161, 164, 173, 175]. EUS-guided sampling of hilar lymph nodes may be used to diagnose and stage suspected biliary cancer [159, 160, 162, 168, 172, 176].

**Recommendation 15**

Complementary to ERCP-guided brushing and biopsy, EUS-guided sampling may be used for diagnosis of indeterminate biliary strictures and gallbladder masses (LoE 2a, GoR B). Strong consensus (100 %).

**Subepithelial gastrointestinal tumours**

Differential diagnosis of gastrointestinal subepithelial tumours (SET) includes leiomyoma, schwannoma, gastrointestinal stromal tumour (GIST), lipoma, ectopic pancreas, neuroendocrine tumour, cyst and others [177 – 179]. Whilst a reliable differential diagnosis is possible on EUS features alone in some SETs, so, with the frequently occurring hypoechoic SET originating from the 4th layer (muscularis propria), i.e. GIST, leiomyoma and schwannoma, diagnosis may be problematic [177 – 179]. EUS-guided sampling combines smear cytology (SC) with immunohistochemistry for maximal diagnostic accuracy. The diagnostic yield remains suboptimal using standard EUS needles of various diameters, ranging from 20 [180] to 93 % [181], as retrieved tissue is not always sufficient to allow immunohistochemical analysis [181 – 195]. The European Society of Gastrointestinal Endoscopy (ESGE) guideline on EUS-guided sampling in gastroenterology states that in hypoechoic SET of the stomach ≤ 20 mm the usefulness of EUS-guided sampling is limited due to the moderate diagnostic yield and lacking capability to determine the mitotic index [59].

**Recommendation 16**

EUS-guided sampling may be used for differential diagnosis of subepithelial gastrointestinal tumours ≥ 20 mm in cases with high surgical risk or suspected non-resectability (LoE 2b, GoR C). Strong consensus (100 %).

**Lung cancer**

In suspected lung cancer conventional techniques (bronchoscopic or percutaneous-transthoracic biopsy) fail to establish a histological diagnosis in up to ¼ of patients. EUS allows effective transoesophageal or transbronchial guidance for sampling of centrally located lung masses or lymph nodes adjacent to the oesophagus or the respiratory tract [196 – 203]. Subtyping of non-small cell lung cancer is the basis of personalized oncological treatment and is feasible using EUS-guided sampling in 77 – 90.6 % [204 – 206]. Genotyping was possible using EUS-/EBUS fine-needle aspirates in 77 – 98.4 % [124, 204 – 209].

**Recommendation 17**

In suspected lung cancer EUS-guided sampling (EUS-FNA, EBUS-TBNA) of centrally-located mass lesions or suspicious lymph nodes is recommended to establish a definite tissue diagnosis complementing other diagnostic techniques (LoE 2b, GoR B). Strong consensus (100 %).

**Mediastinal and abdominal lymphadenopathy of unknown origin**

For lymph node classification as malignant or benign, EUS-FNA has a higher accuracy than EUS features alone [210, 211]. A meta-analysis (n = 9310) showed significant improvement in evaluating mediastinal lymphadenopathy with EUS-guided sampling over EUS features of lymph nodes (sensitivity 88 vs. 84.7 %; specificity 96.4 vs. 84.6 %) [212]. Several studies and meta-analyses have demonstrated a high diagnostic yield and accuracy of EUS-guided sampling (EUS-FNA and EBUS-TBNA) for the evaluation of indeterminate mediastinal and abdominal lymphadenopathy, with or without a malignant disease background [114, 116, 120, 212 – 234].

**Recommendation 18**

EUS-guided sampling (EUS-FNA, EBUS-TBNA) is recommended as the primary diagnostic technique for tissue diagnosis of mediastinal or abdominal lymphadenopathy of unknown aetiology (LoE 3a, GoR B). Strong consensus (100 %).

**EUS-guided sampling for tumour staging**

**Algorithmic approach**

For tumour staging an algorithmic approach of EUS-guided sampling following the TNM classification of the respective malignant tumour is sensible. Potential sites of metastases within reach of EUS-FNA should be examined and sampled first, followed by regional lymph node stations, in which demonstration of metastatic involvement would alter management [76, 235 – 237]. This systematic approach in an inverse TNM order meets concerns that needle contamination with tumour cells from the first sampling target, from the instrumental channel or from gas-
Guidelines

Mediastinal lymph nodes

In the absence of metastases, mediastinal lymph node involvement is the most important factor affecting the management and outcome. EUS-guided sampling may play a pivotal role in the detection and diagnosis of disease recurrence and distant spread of gastrointestinal and pancreatic malignancies. After acquiring clinical information, EUS with EUS-guided sampling can be used to perform lymph node staging, allowing targeted biopsy of all relevant mediastinal lymph node locations. However, EUS sampling alone is insufficient for nodal staging of non-small cell lung cancer (NSCLC). EUS-FNA of mediastinal lymph nodes is more sensitive and accurate than EUS alone for the assessment of tumour spread to mediastinal lymph nodes compared with the oesophageal approach alone, and a 13% increase in the proportion of EBUS-TBNA patients with lymph node involvement is shown when combining mediastinoendoscopy with EUS-FNA. A meta-analysis (n = 533 patients) showed a sensitivity of 85% and a specificity of 99% for EBUS-TBNA compared with EUS-FNA of mediastinal lymph nodes in NSCLC, with EUS-guided sampling only being useful in selected cases. EUS-FNA of mediastinal lymph nodes for the N staging of oesophageal cancer is more sensitive and accurate than EUS alone and significantly impacts therapeutic decisions. EUS-FNA of mediastinal lymph nodes in a variety of extra-thoracic malignant disease has been shown to be more accurate than CT in defining the disease stage [244, 246, 247]. EUS-FNA of mediastinal lymph nodes in patients with suspected lung cancer requires a sensitivity and specificity of 83-84% and 94-100%, respectively. EUS and EUS-TBNA in combination allow targeting of nearly all relevant mediastinal lymph node locations. In patients with suspected lung cancer, EUS-FNA has a sensitivity of 99% and a specificity of 98%, with a substantial effect on clinical management [248].

Adrenal glands and liver masses

Previous studies have shown high accuracy and clinical impact of EUS-guided sampling of adrenal masses in oesophageal cancer staging. Malignant involvement of adrenal masses can provide valuable information for the staging of lung cancer and other malignancies. EUS-guided sampling of adrenal masses is viable and has been shown to be more accurate than CT in defining the disease stage [245]. EUS-FNA of adrenal masses has shown a sensitivity of 95% and a specificity of 96% [246]. EUS-guided sampling of adrenal masses is indicated in adrenal masses of uncertain nature, to rule out malignancy. EUS-guided sampling of adrenal masses is indicated in adrenal masses of uncertain nature, to rule out malignancy. EUS-guided sampling of adrenal masses is indicated in adrenal masses of uncertain nature, to rule out malignancy. EUS-guided sampling of adrenal masses is indicated in adrenal masses of uncertain nature, to rule out malignancy. EUS-guided sampling of adrenal masses is indicated in adrenal masses of uncertain nature, to rule out malignancy. EUS-guided sampling of adrenal masses is indicated in adrenal masses of uncertain nature, to rule out malignancy. EUS-guided sampling of adrenal masses is indicated in adrenal masses of uncertain nature, to rule out malignancy. EUS-guided sampling of adrenal masses is indicated in adrenal masses of uncertain nature, to rule out malignancy. EUS-guided sampling of adrenal masses is indicated in adrenal masses of uncertain nature, to rule out malignancy. EUS-guided sampling of adrenal masses is indicated in adrenal masses of uncertain nature, to rule out malignancy. EUS-guided sampling of adrenal masses is indicated in adrenal masses of uncertain nature, to rule out malignancy. EUS-guided sampling of adrenal masses is indicated in adrenal masses of uncertain nature, to rule out malignancy. EUS-guided sampling of adrenal masses is indicated in adrenal masses of uncertain nature, to rule out malignancy. EUS-guided sampling of adrenal masses is indicated in adrenal masses of uncertain nature, to rule out malignancy. EUS-guided sampling of adrenal masses is indicated in adrenal masses of uncertain nature, to rule out malignancy. EUS-guided sampling of adrenal masses is indicated in adrenal masses of uncertain nature, to rule out malignancy. EUS-guided sampling of adrenal masses is indicated in adrenal masses of uncertain nature, to rule out malignancy. EUS-guided sampling of adrenal masses is indicated in adrenal masses of uncertain nature, to rule out malignancy. EUS-guided sampling of adrenal masses is indicated in adrenal masses of uncertain nature, to rule out malignancy. EUS-guided sampling of adrenal masses is indicated in adrenal masses of uncertain nature, to rule out malignancy. EUS-guided sampling of adrenal masses is indicated in adrenal masses of uncertain nature, to rule out malignancy. EUS-guided sampling of adrenal masses is indicated in adrenal masses of uncertain nature, to rule out malignancy.
for solid pancreatic lesions and metastatic lymph nodes, satisfactory for biliary mass lesions, but only moderate for gastrointestinal SET and PCL [59, 66, 325, 326]. Currently, the 22 G aspiration needle is most widely used, with increased utilization of 25 G aspiration needles [327–329].

**22 G vs. 25 G aspiration needles**

Meta-analyses demonstrate slight superiority of the 25 G over the 22 G aspiration needle for EUS-FNA of solid pancreatic lesions [330, 331].

**19 G vs. 22 G and 25 G aspiration needles**

There was no difference in diagnostic accuracy between the 19 G and the 22 G aspiration needle by intention-to-treat analysis, but the 19 G was superior to the 22 G needle in terms of the amount of cellular material and diagnostic accuracy by per-protocol analysis (following exclusion of 5 technical failures, all with pancreatic head lesions). Further analysis favoured the 19 G aspiration needle for the sampling of lesions within the pancreatic body and tail [332]. A flexible 19 G aspiration needle has been shown to be effective for procuring aspirates from pancreatic head lesions even by the transduodenal route [333–335].

**21 G vs. 22 G aspiration needles for EBUS-TBNA**

A meta-analysis of 5 studies (n = 1720 patients) did not reveal significant differences in the diagnostic yield, sample adequacy, or the mean number of needle passes between the 21 and 22 G needles during EBUS-TBNA [336].

**EUS-guided tissue acquisition for histopathological processing**

With increasing requirements of histopathological, immunohistochemical and molecular biological diagnosis [337, 338], three types of “histology needles” have been developed to facilitate acquisition of core cylinders suitable for histopathological examination: trucut needles [50, 51, 339, 340], aspiration needles with a side fenestration (core trap) and reverse bevel technology [53, 55, 341, 342] and aspiration needles with a shark mouth design. Acquisition of tissue suitable for histopathological processing is also possible with standard aspiration needles, not just with “histology needles” [54, 59, 66, 323, 324, 343, 344]. The diagnostic yield for histological material is dependent on the needle diameter.

**Recommendation 24**

In a referral EUS centre it is advisable to have needles of different size available, and the choice among different needles should be made considering the anatomical location and the type of the target lesion and the preferred mode of processing of the material (LoE 2b, GoR C). Strong consensus (100%).

**Recommendation 25**

22 G aspiration needles have a high diagnostic yield, low risk of adverse events, and good technical performance, and therefore should be regarded as the current standard for EUS-guided sampling (LoE 2a, GoR B). Strong consensus (100%).

**Recommendation 26**

25 G aspiration needles are not inferior to 22 G aspiration needles with respect to cytological yield (LoE 1a, GoR A). The use of 25 G needles should be considered in particular for EUS-guided transduodenal sampling of pancreatic head lesions (LoE 2a, GoR C). Strong consensus (100%).

**Recommendation 27**

Due to a high rate of technical failure, the use of nonflexible 19 G needles is not recommended for transduodenal sampling of pancreatic head masses (LoE 2b, GoR B). Strong consensus (100%).

**Factors influencing the yield of EUS-guided sampling**

**Features of the target lesion**

The hypocellular character of the PCL aspirate renders the diagnostic yield and accuracy of EUS-guided sampling of PCL considerably lower than in the case of solid pancreatic lesions. For gastrointestinal SET the diagnostic yield of EUS-guided sampling (specific diagnosis) is approximately 34% [187] to 82% [184], with most studies reporting a yield of 52–63% [179]. Data on the influence of the size and site of solid pancreatic lesions on EUS-FNA accuracy are conflicting [138, 345–348]. The diagnostic accuracy of EUS-FNA for pancreatic cancer is considerably hampered in patients with chronic pancreatitis [349–353]. There are inconsistent results in studies evaluating the influence of biliary stents on EUS-FNA performance characteristics [354, 355].

**Sampling technique**

**Fanning technique**

Sampling of all parts of solid lesions and lymph nodes (“fanning technique”) is recommended by the ESGE guidelines on EUS-guided sampling in gastroenterology [54]. According to the results of an RCT, significantly fewer passes are required to establish a diagnosis using this particular technique [356].

**Recommendation 28**

Fanning technique should be applied in EUS-guided sampling to increase sample adequacy (LoE 2b, GoR C). Strong consensus (100%).

**Suction**

Traditionally negative suction pressure using a 5 ml or 10 ml syringe is used to facilitate EUS-guided tissue acquisition. Alternatively, the fine-needle capillary sampling technique [357] or slow removal of the stylet (“slow-pull technique”) [358] may be used to draw cells up into the needle lumen. Applying suction to the needle may increase the cellular yield of the aspirate but, at the same time, potentially increases artefacts and contamination by blood. Applying high negative pressure suction has been suggested to facilitate procurement of core tissue in EUS-FNA and EBUS-TBNA [184, 359–363]. An RCT demonstrated that EUS-guided lymph node sampling with suction compared with the fine-needle capillary technique did not improve the likelihood of correct diagnosis, but, due to excessive blood contamination, provided smears of significantly poorer quality [364].

For sampling of pancreatic masses using 22 and 25 G aspiration needles, one RCT favoured EUS-guided fine-needle sampling with 10 ml suction over no suction [365]. In a multicentre RCT the adequacy and accuracy of 22 G EUS-FNA from solid pancreatic lesions were significantly better with 20 ml suction compared with 10 ml suction and no suction [366].

**Recommendation 29**

For aspiration of cysts and fluid collections, negative pressure suction is recommended (LOE 5, GoR D). Strong consensus (100%).

**Recommendation 30**

For sampling of solid pancreatic lesions, negative pressure suction should be considered (LOE 1b, GoR A). Strong consensus (100%).

**Recommendation 31**

In lymph nodes and other highly vascularized solid lesions, sampling without applying suction should be considered (LOE 2b, GoR C). Strong consensus (100%).

**Use of a stylet**

No advantage of using a stylet in terms of specimen quality and diagnostic yield has been demonstrated [367 – 371]. An RCT suggested a lower proportion of adequate samples and a higher probability for blood contamination in needle passes using a stylet compared with needle passes performed without a stylet [367]. A multicentre RCT reported non-inferiority of performing 22 G EUS-FNA of various solid lesions without a stylet compared with using a stylet with respect to the acquisition rate of histological specimens [372].

**Recommendation 32**

Aspiration needles may be used with or without a stylet with the same diagnostic yield (LoE 1b; GoR A). Strong consensus (100%).

**Number of needle passes**

The number of passes is determined based on the results of on-site cytopathological evaluation, should this be available (ROSE) [54, 373, 374]. As an alternative to ROSE-guided sampling, a high number (≥7) of needle passes is thought to be a predictor of a high diagnostic sensitivity of EUS-FNA [327].

**Pancreatic lesions**

ROSE study results indicate that diagnosis of a solid pancreatic lesion using standard aspiration needles may require 5 – 7 passes [375 – 377]. When using a 25 G aspiration needle for sampling of a solid pancreatic lesion, 2 – 4 needle passes were adequate for a good diagnostic accuracy [378 – 381]. Using a standard 22 G aspiration needle, 1 – 2 needle passes were sufficient to achieve a high diagnostic yield and accuracy in 92% of cases with solid pancreatic lesions [382].

In the case of PCL with no solid component, a single pass and complete cyst fluid aspiration should be performed to prevent cyst infection [54]. In the case of PCL with a solid component, the diagnostic yield is significantly higher for ≥2 needle passes (78%) compared to one needle pass (44%) [383].

**Non-pancreatic lesions**

In the case of lymph nodes, liver metastases and adrenal lesions, a lesser number of needle passes is sufficient to provide a high diagnostic yield. Approximately 3 needle passes were sufficient for EUS-FNA and EBUS-TBNA of lymph nodes [364, 375, 377, 379, 384].

**Recommendation 33**

If ROSE is not available, the number of needle passes should be based on gross visual inspection of the obtained material and the type of target lesion (LoE 2b, GoR C). Strong consensus (100%).

**Targeted sampling**

EFSUMB guidelines recommend the use of contrast-enhanced endoscopic ultrasound (CE-EUS) and of EUS-elastography to facilitate differential diagnosis of solid pancreatic lesions, PCL and lymph nodes [385, 386].

**Lymph nodes**

EUS-elastography, currently strain elastography, has been shown to increase the discriminatory ability of normal EUS criteria, and to improve the specificity of lymph node staging in gastrointestinal malignancy [386, 387]. A meta-analysis calculated a sensitivity of 88% and specificity of 85% with EUS-elastography for differentiating between benign and malignant lymph nodes [388]. EFSUMB guidelines suggest using EUS-elastography for identifying suspicious lymph node and/or harder lymph node regions as targets of EUS-guided sampling [386].

**Solid pancreatic lesions**

The selection of patients for EUS-guided sampling with a high suspicion of diagnoses other than PDAC is of pivotal importance to avoid unnecessary pancreatic head resection with inherent morbidity and mortality [343]. Contrast-enhanced ultrasound (CEUS) and CE-EUS are able to discriminate hypovascular PDAC from other iso- and hypervascular solid pancreatic lesions (non-PDAC) with an accuracy > 85% [385, 389]. A meta-analysis reported the sensitivity and specificity of CE-EUS to diagnose PDAC at 94 and 89%, respectively [390]. When there is a high suspicion of non-PDAC, patients with solid pancreatic mass lesions which are hypervascular or isovascular in comparison with the surrounding pancreatic parenchyma should be referred to pre-therapeutic EUS-guided sampling. Negative results of EUS-guided sampling of solid pancreatic lesions which are hypovascular on CE-EUS should be considered false-negative, initiating repeat sampling or proceeding directly to surgery [391, 392]. An RCT demonstrated that CE-EUS-guided FNA of solid pancreatic lesions is more efficient than conventional EUS-FNA with fewer needle passes required to obtain a diagnostic sample [393].

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10 The terms “number of passes” and ROSE are explained in the addendum on terminology (online version only).
EUS-elastography has an high sensitivity, but only a moderate specificity for differentiating malignant from benign solid pancreatic lesions [387]. Meta-analyses report a sensitivity of 95–97% and a specificity of 67–76% [394–398]. EFSUMB guidelines recommend that in the case of persisting strong clinical suspicion of pancreatic cancer with inconclusive or negative EUS-guided sampling, a hard focal lesion on elastography and/or a hypovascular lesion on CEUS and or CE-EUS indicates the need for repeat EUS-FNA or referral to surgery [386].

Cystic pancreatic lesions
The presence of epithelial mural nodules, thick walls, and septa has been shown to be highly predictive for malignancy in BD-IPMN [399, 400]. International and European guidelines recommend surgical treatment of BD-IPMN in particular if enhancing nodules or “positive cytology” has been verified [144, 145]. Targeted biopsy of the cyst wall and solid components has a significant incremental diagnostic value over cyst-fluid aspiration alone for the diagnosis of mucinous and malignant cystic neoplasms [383, 401, 402] and is recommended by the ESGE guideline on EUS-guided sampling in gastroenterology [54]. EUS and CE-EUS significantly improve the discrimination and characterization of mural nodules in IPMN in comparison with B-mode EUS and CT [403 – 406], suggesting that CE-EUS may facilitate EUS-guided sampling of solid components in PCL.

Subepithelial gastrointestinal tumours
Differential diagnosis of hypoechoic gastric SETs by EUS and other imaging techniques is difficult [66, 178, 179, 191, 407]. Preliminary findings suggest that CE-EUS can discriminate GIST (hypervascular) from benign lesions (leiomyoma, lipoma: hypovascular) [408].

Recommendation 34
Contrast-enhanced EUS and/or EUS-elastography may be used to guide and target EUS-guided sampling of lymph nodes (LoE 2a), solid pancreatic masses (LoE 2a), pancreatic cystic lesions (LoE 2b) and gastrointestinal subepithelial tumours (LoE 3b) (GoR C). Strong consensus (100%).

Recommendation 35
In addition to aspiration of cyst fluid, the cyst wall and solid components, if present, should be sampled in pancreatic cystic lesions (LoE 2b, GoR B). Strong consensus (100%).

EUS-guided sampling: Processing and evaluation of specimen

Techniques for expelling a specimen from the needle
The techniques for expelling a specimen are related to the needle system used. There are three techniques for expelling a specimen from aspiration needles: 1. Air flushing; 2. Saline flushing; 3. Extrusion with the stylet. No differences between air flushing and stylet reinsertion with regard to the number of diagnostic specimens, overall accuracy and specimen quality were found in an RCT, but smears were bloodier following stylet use for the expelling of the sample [365].

Recommendation 36
Material can be expelled from EUS-aspiration needles using the stylet, by spraying with air on slides, or by flushing the needle with saline (LoE 2b, GoR B). Strong consensus (100%).

Recommendation 37
Controlled release of material onto slides using the stylet is the preferred method despite more blood contamination (LoE 5, GoR D). Strong consensus (100%).

Methods for specimen preparation and processing
Optimizing specimen handling and processing may reduce the frequency of non-diagnostic samples [409, 410]. The handling of the specimen depends on the diagnostic methods used and the final cytopathological evaluation.

Cytopathology: conventional smears and liquid-based preparations
For cytological analysis the specimen should be smeared onto slides (smear cytology, SC) or incorporated into special preservation solutions (liquid-based cytology, LBC) for further handling [411, 412]. Prospective comparative trials demonstrated that LBC was less sensitive with material obtained by EUS-guided sampling from lymph nodes and pancreatic lesions compared with traditional SC [413–415]. A further RCT suggested that LBC may serve as a complementary preparation technique if blood contamination of smears is abundant [414].

Histopathology: cell block and core tissue preparation
In general, histopathological processing including paraffin-embedding, microtome-sectioning, and staining is possible with small coherent tissue cores embedded in a formalin fixative or with CB. A wide variety of ancillary diagnostic techniques (immunohistochemistry, molecular analysis) may be performed on histopathologically processed material [66, 186, 382, 409, 411, 412, 416–420].

CB may be prepared from any cell suspension (e.g. liquid aspirates, needle rinses, or specimen in preservative solutions for LB). Pellets of artificially aggregated cells are obtained by using repeated centrifugation and/or special cell capturing gels [410, 412, 417, 421]. Small core particles obtained with aspiration needles or “histology needles” can be directly delivered into formalin solution for fixation followed by standard histopathological processing [53, 217, 409, 410, 422, 423]. Alternatively, core particles may be isolated from samples expelled on glass slides or preservation solutions by the stylet method before further cytopathological processing (“split-up” method) [186, 382, 424].

Cytopathological evaluation of a specimen
Cytopathological vs. histopathological evaluation
In most cases of PDAC and lymph node metastases, cytopathological evaluation alone is adequate to establish a diagnosis of malignancy. The diagnostic sensitivity of EUS-FNA cytology is highly dependent on the proficiency and experience of the cytopathologist evaluating the specimen [107, 425] and to a lesser extent on the availability of ROSE. In a number of clinical settings and target lesions, it is tissue architecture, immunohistochemistry and molecular analysis rather
than cellular features that are essential for accurate pathological assessment [419, 420]. For benign diseases, e.g. autoimmune pancreatitis [426, 427], differential diagnosis of SET (GIST vs. leiomyoma or schwannoma) [52, 181, 189, 191, 428, 429], subtyping of NSCLC and malignant lymphoma [217, 266, 430 – 432], diagnosis of rare tumours [137, 226, 382], grading of neuroendocrine tumours [433 – 435], molecular profiling of solid tumours [124 – 128, 130, 204 – 209, 409, 436, 437] and differential diagnosis of mediastinal lymph node metastases [52, 226, 438], a core sample is preferred to a cytological aspirate [66].

A complementary approach: combining cytopathological and histopathological assessment

There are two reasonable strategies to achieve high diagnostic accuracy in EUS-guided sampling, without the attendance of a cytopathologist: combining cytopathological and histopathological evaluation [52, 138, 192, 231, 382, 415, 424, 439, 440], or relying on core tissue or CB material for histopathological assessment [53, 55, 217, 333, 409, 422, 423, 427, 430, 441 – 444]. Several studies have shown an incremental yield of combining SC with CB technique or histological core assessment in terms of diagnostic yield and accuracy [138, 192, 231, 382, 415, 439, 440, 445, 446].

Specimen handling for diagnosis of infectious diseases

Suspected infectious disease (e.g. abscesses, tuberculosis or atypical mycobacteriosis) requires specific handling of the aspirate. Microbiological cultures are possible if samples are placed in saline, whereas a polymerase chain reaction (PCR) amplifying mycobacterial DNA requires the specimen placed in formalin [447, 448]. A moderate to high accuracy of SC including Ziehl-Neelsen stain for acid-fast bacteria alone for the diagnosis of tuberculosis and its differentiation from sarcoidosis or histoplasmosis has been reported [447, 449 – 454]. Further studies showed significantly higher sensitivity of PCR vs. microbiological culture and/or SC, and of microbiological culture vs. SC [448, 455 – 457]. In a meta-analysis the overall diagnostic yield of EBUS-TBNA for the diagnosis of mediastinal tuberculous lymphadenitis was 80% with the culture positive rate (54%) being significantly higher than the SC positive rate (33%) [458].

Recommendation 38

The processing of specimen obtained by EUS-guided sampling varies according to the cytopathological or histopathological methods used for diagnosis (LoE 2b, GoR C). Strong consensus (100%).

Recommendation 39

Tissue fragments can be effectively isolated from the whole sample after gross visual examination without impairing the cytopathological result (LoE 2b, GoR B). Strong consensus (100%).

Recommendation 40

Without rapid on-site cytopathological evaluation, the combination of cytopathological and histopathological processing seems to provide the most reliable results (LoE 2b, GoR B). Strong consensus (100%).

On-site assessment of the adequacy of specimens

Rapid on-site cytopathological evaluation

ROSE is reported to be used in the majority of centres in the United States [106, 329], but on-site cytopathological service is not routinely available in Europe [327, 328]. Positive assessments of the role of ROSE for EUS-FNA of solid pancreatic lesions and for EUS-FNA and EBUS-TBNA of lymph nodes have been documented [138, 375, 413, 459 – 471]. A high concordance of preliminary on-site interpretation and final cytopathological diagnoses has been reported [460, 466, 472, 473]. Other studies have suggested that ROSE is not essential for high diagnostic yield and accuracy of EUS-FNA [54, 66, 474 – 476]. Only five studies with EUS-FNA of pancreatic lesions used a 2-cohort design with head-to-head comparison of adequacy and accuracy of ROSE vs. non-ROSE. These studies have been included in a meta-analysis, showing that ROSE was associated with a significant 10% improvement of the per-case adequacy rate only in studies with a low adequacy rate (<90%). No significant impact of ROSE on diagnostic yield, accuracy or number of needle passes was observed [477]. In a second meta-analysis of 68 single-cohort and 2 two-cohort studies, ROSE was associated with a low, statistically significant advantage of adequacy rates (2.3%), [478]. A further meta-analysis showed a non-significantly higher per-case adequacy in ROSE studies compared with studies without ROSE [136]. The largest meta-analysis assessing the diagnostic performance of EUS-FNA of solid pancreatic lesions demonstrated significantly lower heterogeneity among studies with a cytopathologist present on-site, but advantages over studies without ROSE in terms of sensitivity (88% vs. 80%) and diagnostic odds ratio did not reach statistical significance [132]. Two RCTs showed no benefit of ROSE in EBUS-TBNA for diagnosing lung cancer and in EUS-FNA of solid pancreatic lesions with regard to diagnostic accuracy, procedure time and safety. However, in both trials ROSE-assisted sampling required fewer needle passes to obtain a diagnosis [467, 467a]. The relatively small benefit of ROSE has to be balanced against the cost and time expenditure of a permanent on-site cytopathology service [377, 479 – 481].
Gross visual inspection and ROSE performed by the endosonographer
In the absence of ROSE, gross visual assessment of a sample to assess the adequacy of EUS-guided sampling is possible [482]. Studies show conflicting levels of the ability of endosonographers and trained EUS and cytology assistants to assess slide adequacy and to differentiate malignant and benign specimens [483]. In a prospective evaluation of the visual assessment of core particles obtained with a 19 G ProCore needle, a core length of ≥ 4 mm was an indicator for an adequate histological sample and for a high overall, histological and cytological diagnostic yield [443].

Studies have demonstrated the improved ability of endosonographers, following completion of structured training, to smear and stain slides, to operate a microscope [484], to judge adequacy of a cytological specimen confidently [485], and to differentiate benign and malignant smears with an accuracy of 89% [486]. ROSE performed by an experienced endosonographer may be reliable, but requires completion of an intensive structured training program.

Recommendation 44
Rapid on-site cytopathological assessment does not substantially improve the diagnostic yield of malignancy or generally reduce the number of needle passes. In centres with an adequacy rate < 90%, ROSE may improve the adequacy of EUS-guided sampling (LoE 2a, GoR C). Strong consensus (100%).

Recommendation 45
Rapid on-site cytopathological assessment may be used to guide the number of needle passes, provide a reliable immediate diagnosis of malignancy, and assess the need for ancillary studies (LoE 2b, GoR C). Strong consensus (100%).

Reporting and reliability of cytopathology in EUS-guided sampling
Cytopathological reporting
In most cytopathological laboratories, the diagnostic categories for EUS-cytology follow a Bethesda-like system: unsatisfactory, benign, atypical, suspicious, and positive for malignancy [487, 488]. Based on a standard nomenclature for FNA [489], the Papanicolaou Society of Cytopathology has developed guidelines on standardized terminology and nomenclature for pancreatobiliary cytopathology, recommending a six-tiered system of diagnostic categories: non-diagnostic, negative, atypical, neoplastic, suspicious, and positive [12].

The category “neoplastic” is unique for pancreatic cytopathology, including benign neoplasms (serous cystadenoma) as well as pre-malignant mucinous cysts, neuroendocrine tumours and solid pseudopapillary neoplasms [490]. The inconclusive and ill-defined diagnostic categories “atypical” and “suspicious for malignancy” [13] are inconsistently conveyed [491 – 493]. Insufficient adherence to standardized nomenclature in cytopathological reporting hampers individual patient management and is a substantial obstacle for the comparison of results between centres and studies [412, 490].

Reliability of negative (“benign”) results and findings positive for malignancy/neoplasia
A few studies have analysed the reproducibility of cytopathological diagnoses of specimens obtained by EUS-guided sampling. In most studies a good-to-excellent diagnostic agreement between experienced cytopathologists was documented [425, 494, 495]. For clinical decision-making it is important to understand that there is still a significant risk of malignancy with negative findings in EUS-guided sampling, which is considerably high for PCL, solid pancreatic lesions in patients with underlying chronic pancreatitis, indeterminate biliary strictures, and gastrointestinal SET. The major cause for false-negative diagnoses of EUS-guided sampling is sampling error, whereas interpretative error is rare [66, 165, 325, 349, 350, 491, 492, 496 – 501].

In studies with good reference standards and long follow-up, false-positive findings have been reported for EUS-guided sampling in 1.1 – 5.3% when only cases with a positive cytopathological result were considered and in 7.8% if suspicious cytopathology results were included. False-positive rates seem to be higher in luminal compared with extra-luminal (e.g. pancreatic) primary cancer [238, 502, 503]. False-positive findings may result from procedure-related factors, e.g. traversal of a neoplastic area, needle contamination with tumour cells [240, 241], inadvertent aspiration of cancer-cell contaminated luminal fluid [239], or from cytopathological misinterpretation [66, 325, 491, 496 – 498, 501, 502, 504, 505].

Malignancy risk associated with inconclusive findings
Studies have evaluated the malignancy risk related with inconclusive diagnostic cytopathological categories for EUS-guided sampling of pancreatic lesions. The suspicious category is associated with an 80 – 96% risk of neoplastic or malignant final diagnosis, whereas the atypical category carries a markedly variable and significantly lower malignancy risk [492, 493, 506]. Clinical predictors associated with a high likelihood of a neoplastic diagnosis following an atypical cytopathological diagnosis in pancreatic EUS-FNA can be used to adapt management of patients with inconclusive cytopathological diagnoses: ancillary testing, watchful waiting, or initiation of a specific treatment [501, 507 – 510].

Recommendation 46
A standardized and validated classification system should be used for cytopathology reporting in EUS-guided sampling (LoE 5, GoR D). Strong consensus (100%).

Recommendation 47
After inconclusive cytopathological results, the use of ancillary studies and/or repeat EUS-guided sampling should be weighed against the clinical background of patient history, clinical and laboratory data, EUS morphology, results of other imaging tests and follow-up (LoE 2b, GoR B). Strong consensus (100%).

11 Terms are explained in the addendum on terminology (long version only).
12 Terms are explained in the addendum on terminology (long version only).
13 Terms are explained in the addendum on terminology (long version only).
Adverse events of EUS-guided interventions and their prevention

General considerations
EUS-guided sampling (including EBUS-TBNA) is a safe diagnostic technique. Severe adverse events are rare. There is no evidence that needle size or needle type affects morbidity of EUS-guided sampling [54, 70, 330, 513]. A systematic review of complications of 51 EUS-FNA studies including 10 941 patients describes an overall complication rate of 0.98%. The procedure-related mortality was estimated at 0.02% [514]. Morbidity of EUS-guided therapeutic interventions is considerably higher than EUS-guided sampling, with risk and type of complications depending on the type of procedure [71, 515, 516].

Adverse events
Perforation
The majority of perforations associated with EUS-guided sampling are caused by the passage of the gastrointestinal echoendoscope and not the sampling procedure. Multicentre surveys and studies report perforation rates of 0.03 – 0.15% [516 – 520]. The risk of duodenal perforation is higher than oesophageal perforation [70, 516].

Acute pancreatitis
The frequency of acute pancreatitis following EUS-FNA of pancreatic lesions is between 0.19 – 2.35% [70]. Pancreatitis risk is higher in patients with PCL than in patients with solid pancreatic lesions [70, 514].

Intra- and extraluminal haemorrhage
Severe bleeding following EUS-FNA is a rare event (overall 0.13%) [514]. There are three reports in the literature of fatal bleeding following pancreatic EUS-FNA [70]. There is indirect evidence that EUS-FNA of PCL may cause a higher risk of bleeding compared with EUS-FNA of solid pancreatic lesions [514, 515, 521 – 523]. Guidelines suggest discontinuing oral anticoagulants, heparin and LMWHs in therapeutic dosages as well as ADP-antagonists before EUS-guided sampling, whereas withdrawal of ASA is recommended only before EUS-FNA of PCL [54, 513, 524 – 526]. Despite limited evidence for distinct cut-off values, a platelet count < 50 000/ml and an international normalized ratio (INR) > 1.5 are regarded as contraindications to EUS-guided sampling. There is no data regarding needle size, needle type, number of needle passes or other technical factors related to bleeding risk following EUS-guided sampling.

Recommendation 48
In patients on antiplatelets and/or anticoagulants, a risk assessment balancing thromboembolic events versus bleeding should be performed prior to EUS-/EBUS-guided diagnostic and therapeutic interventions (LoE 5, GoR D). Strong consensus (100%).

Recommendation 49
Decision on suspension of antiplatelet drugs and/or anticoagulants or delay of the procedure should be made based on an individual risk assessment (LoE 5, GoR D). Strong consensus (100%).

Fever and infection
Bacteremia following upper as well as lower gastrointestinal tract EUS-FNA is a rare event [527 – 530]. In large series 0.4 – 1.0% of patients experienced febrile episodes following EUS-FNA for solid pancreatic masses [359, 531, 532]. For endosonographic sampling of mediastinal lesions, studies report an overall frequency of major infectious complications in 0.07% of patients [533]. Serious infectious complications have been reported following EUS-FNA of mediastinal cysts [70, 533], ascites [313], perirectal cysts [26], as well as of PCL and (peri-) pancreatic fluid collections [534]. Despite a lack of convincing evidence, several guidelines recommend peri-interventional antibiotic treatment for EUS-FNA of PCL [54, 513, 524]. Due to potentially catastrophic consequences of mediastinal infection, guidelines and reviews regard EUS-FNA of mediastinal cysts to be contraindicated [54, 70, 513, 524, 533].

Recommendation 50
Transoesophageal EUS-guided sampling of cystic mediastinal lesions should be avoided due to the high risk of infection (LoE 4, GoR C). Strong consensus (100%).

Recommendation 51
Peri-interventional antibiotic treatment is recommended for EUS-guided sampling of cystic lesions and fluid collections as well as in EUS-guided drainage procedures (LoE 5, GoR D). Strong consensus (100%).

Tumour cell seeding and needle track metastasis
Several studies agree that EUS-guided sampling for malignant solid and cystic pancreatic tumours and for cholangiocarcinoma is not a risk factor for the development of peritoneal seeding, tumour recurrence, or decreased survival [535 – 541]. The risk of peritoneal seeding following biopsy of pancreatic cancer appears to be significantly lower with EUS-FNA compared to percutaneous FNA [542, 543]. However, 11 individual cases of tumour cell seeding caused by EUS-guided sampling of malignant pancreatic neoplasia (n=8, in 4 cases cystic tumours) [544 – 551] and of lymph node metastases (n=3) [552 – 554] suggest cautious use of EUS-guided sampling.

Recommendation 52
Tumour cell dissemination along the needle track following EUS-guided sampling is an exceptionally rare event. Pre-operative EUS-guided sampling of pancreatobiliary malignancies is not associated with increased risk of postoperative recurrence, decreased overall survival or decreased cancer-specific survival (LoE 2c, GoR B). Strong consensus (100%).

Miscellaneous complications
There are reports of bile peritonitis following EUS-FNA of the gallbladder or of the liver in patients with biliary obstruction [293, 555, 556], of pneumothorax following transoesophageal EUS-FNA of mediastinal lymph nodes [202], pneumoperitoneum following EUS-FNA of a pancreatic tumour [557] and of pancreatic duct leak caused by EUS-FNA of pancreatic mass lesions [558, 559]. One case of phlegmonous gastritis following EUS-FNA has
been reported [560]. Two cases of a hypertensive crisis induced by EUS-FNA of retroperitoneal paraganglioma or adrenal phaeochromocytoma have been documented [561, 562].

**Risk factors for complications**

There are limited data linking complication rates of EUS-FNA with specific target lesions. Data from a systematic review confirmed that EUS-FNA is exceptionally safe for mediastinal lesions (n = 1310; complication rate: 0.38 %), abdominal masses (n = 381; 0.26 %) and adrenal glands (n = 81; 0 %). Procedure-related morbidity seems to be higher for pancreatic lesions (n = 8246; 1.03 %), liver lesions (n = 344; 2.33 %) and perirectal lesions (n = 193; 2.07 %) [514]. Several studies have shown that performing EUS with/without EUS-guided sampling and ERCP in a single session is safe [563–570]. Observations from high-volume centres suggest that complications are more frequent in the learning phase of the examiners [100, 422, 534].

**Recommendation 53**

EUS-guided sampling of solid lesions is a low-risk procedure. EUS-guided sampling of pancreatic cystic lesions has a higher frequency of adverse events compared to solid pancreatic lesions (LoE 2a, GoR B). Strong consensus (100 %).

**Recommendation 54**

EUS-guided sampling and ERCP may be safely performed in a single session (LoE 2b, GoR B). Strong consensus (100 %).

**Recommendation 55**

Any complications of EUS-guided sampling and therapeutic interventions should be documented in a standardized format in every centre. The incidence of adverse events should be monitored and used as a quality indicator (LoE 5, GoR D). Strong consensus (100 %).

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