

Pediatric Sepsis Markers: Interleukins and Others

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Abstract

Cytokines are a very important part in the regulation of immune response in sepsis. They are cellular signaling proteins whose production is a result of activation of specific receptors. Variations in the genes encoding cytokines have a significant role in interindividual susceptibility to sepsis and its severity, and their function may be modulated by age, gender, and some environmental factors. Cytokines are classified into the subsets of proinflammatory, anti-inflammatory, and multiple function cytokines. Regulated balance between proinflammatory cytokines, anti-inflammatory cytokines, and soluble inhibitors of proinflammatory cytokines is important for eliminating pathogens and reducing inflammation. High levels of some cytokines, such as tumor necrosis factor- α , interleukin (IL)-1, and IL-6, determine the course of disease and outcome in sepsis. Likewise, numerous other biomarkers may be potential indicators of sepsis, but none has been routinely used. The best approach to the diagnosis of sepsis is the combination of different biomarkers.

Keywords

- ▶ sepsis
- ▶ immune response
- ▶ cytokines
- ▶ other biomarkers

Introduction

Bacterial sepsis is a complex and dynamic clinical syndrome that is a result of a systemic inflammatory response to bacteria and their products and has a high mortality rate.¹ Markers that can identify high-risk patients are crucial for early detection of sepsis as well as for monitoring the course of disease. Cytokines are a very important part of the development of sepsis, particularly for their role in the regulation of immune response. They are cellular signaling proteins whose production is a result of activation of receptors such as Toll-like receptors (TLRs), retinoid acid inducible protein 1-like receptors and nucleotide-binding oligomerization domainlike receptors that are present on the cell surface and transfer specificity in the innate immune response. Activation of these receptors by pathogens results in production of cytokines, as well as coagulation proteins and complement. Various researches suggest that not only cytokines

by themselves but also the variations in the genes encoding cytokines are important and have a significant role in interindividual susceptibility and severity to sepsis.^{2,3} Also, during the progression of sepsis, the inflammatory cytokines may be modulated by age, gender, and some environmental factors.⁴

Cytokines are small protein mediators with low molecular weights (<40 kDa), which activate and differentiate the immune response. Cytokines are classified into the subsets of proinflammatory, anti-inflammatory, and multiple function cytokines. Tumor necrosis factor- α (TNF- α), inducible protein-10 (IP-10), interleukin (IL)-2, IL-6, IL-8, IL-12, and IL-17 are proinflammatory cytokines. Proinflammatory response includes activation of many immunological pathways where release of specific cytokines represents “cytokine cascade.” Anti-inflammatory cytokines are IL-4, IL-10, IL-1 receptor α , TNF soluble receptor, and transforming growth factor- β 2 (TGF- β 2). These cytokines strive to restore immunological

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balance. Multiple function inflammatory cytokines include IL-1 β , IL-3, monocyte chemoattractant protein-1, soluble CD40 ligand, and growth factors such as granulocyte-colony stimulating factor. Their secondary mediators are thromboxanes, nitric oxide, leukotrienes, prostaglandins, platelet-activating factor, and complements.^{5,6} The inflammatory response, apart from cytokines, includes many biomarkers such as acute-phase proteins release, coagulation factors, vascular endothelium products, leukocytes, histocytes, and platelets.⁷ As it is known, what is important in sepsis is innate immune response, which is triggered by bacterial infection. In the past, sepsis was explained as an exacerbated release of proinflammatory cytokines, such as TNF- α , IL-1, IL-6, IL-12, macrophage migration inhibitory factor (MIF), and interferon- γ (IFN- γ), but recently it has been shown that compensatory anti-inflammatory response that occurs after hyperinflammatory period is also important. It is known that the proinflammatory response is antagonized by anti-inflammatory cytokines, including IL-10, IL-4, and TGF- β , which attempt to restore immunological balance.^{8,9} Most adverse effects of sepsis such as systemic inflammatory response syndrome (SIRS), disseminated intravascular coagulation (DIC), septic shock, complement activated response syndrome, and multiple organ dysfunction syndrome are associated with an imbalance in the production of proinflammatory mediators as well as counterweight synthesis of anti-inflammatory cytokines. Regulated balance between proinflammatory cytokines, anti-inflammatory cytokines, and soluble inhibitors of proinflammatory cytokines, such as soluble TNF receptors (sTNFRs), IL-1 receptor antagonist (IL-1Ra), and IL-1 receptor type II (IL-1R2), is important for eliminating pathogens and reducing inflammation.¹⁰ It is also known that high levels of some cytokines, such as TNF- α , IL-1, and IL-6, determine the course of disease and outcome in sepsis.¹¹

Interleukins

TNF- α is a 17-kDa protein derived predominantly from macrophages but also partly from nonimmune cells such as fibroblasts.¹² The release of TNF- α begins within 30 minutes after the stimulus, following gene transcription and RNA (ribonucleic acid) translation. TNF- α acts through specific transmembrane receptors, named TNF receptor (TNFR) 1 and TNFR2,¹³ and activates immune cells as well as the release of other immunoregulatory mediators. In experimental endotoxemia, the circulating levels of TNF- α reach the peak in one and half hour after stimulus.¹⁴ The release of TNF- α causes vasodilatation and increases vascular permeability, which may lead to systemic edema. Decreased blood volume and hypoproteinemia can lead to progression of shock. Leukocyte and platelet adhesion with depletion of coagulation factors may lead to DIC with multiple-organ failure and death.¹⁵ In experimental animals, the injection of TNF- α causes a syndrome that looks like septic shock,¹⁶ and infusion of recombinant TNF- α into humans results in SIRS.¹⁷ TNF- α was one of the first mediators identified in inflammation¹⁸ and has been suggested as a marker for the prediction of early sepsis in children, especially when used with IL-6.¹⁹ But due to the short half-life (~70 minutes) and its interaction with soluble receptor, its detection is exceptionally difficult.²⁰

IL-1 is a proinflammatory cytokine that is released from activated macrophages, as TNF- α , and signaled through two receptors, IL-1 receptor type I and IL-1R2, which has comparable effects on immune cells.²¹ IL-1 family includes two agonists (IL-1 α and IL-1 β) and one antagonist (IL-1Ra) as mediators of immune response to sepsis.³ IL-1Ra levels reach peak 2 to 4 hours after application of endotoxin and remain elevated for more than 24 hours. It has a longer half-life than IL-6 and is potentially a better biomarker in the diagnosis of sepsis.²² IL-1 induces coagulation and extravasation of inflammatory cells, and persistently elevated levels can be correlated with multiple-organ failure and worse prognosis in adults.²³

TNF- α and IL-1 synergistically induce a shocklike state, which is characterized by vascular permeability, severe pulmonary edema, and hemorrhage.²⁴ Also, they are responsible for fever and belong to a group of pyrogenic cytokines.²⁵ TNF- α and IL-1 act on various cells such as macrophages, endothelial cells, and neutrophils. TNF- α promotes the activation and differentiation and prolongs survival of macrophages.²⁶ In endothelial cells, TNF- α augment the expression of intercellular adhesion molecule-1, vascular cell adhesion molecule-1, and chemokines.²⁷ Also, TNF- α promotes neutrophils extravasation into tissues and upregulates action on endothelial expression of procoagulant.¹⁸ TNF- α and IL-1 activate macrophages to secrete other proinflammatory cytokines (IL-6, IL-8, and MIF) and lipid mediators.⁹

Soluble cytokine receptors and receptor antagonists, such as sTNFRs, IL-1R2, and IL-1Ra, modulate the actions of TNF- α and IL-1. Plasma concentrations of sTNFRs correlate not only with disease severity but also with mortality.²⁸ As opposed to that, administration of IL-1Ra increased survival and in that way suggests positive therapeutic effect for IL-1Ra.²⁹

On top, TNF- α and IL-1 are involved in many other inflammatory diseases such as rheumatoid arthritis, osteoarthritis, and atherosclerosis.³⁰⁻³²

IL-6 is a glycoprotein produced by macrophages, lymphocytes, fibroblasts, endothelial dendritic, and smooth muscle cells.^{33,34} During the acute phase of an infection, B and T lymphocytes are stimulated to produce IL-6. IL-6 activates the coagulation system, and modulates hematopoiesis as well as stimulates the release of TNF- α and IL-1. IL-6 is an early marker of sepsis in neonates and children and is more potent than C-reactive protein (CRP).³⁵ IL-6 has high sensitivity and specificity in predicting positive cord blood culture in newborns with premature rupture of membranes,³⁶ and plasma levels of IL-6 of 160 pg/mL are 100% sensitive for the diagnosis of early onset sepsis in neonates. The combination of IL-6, TNF- α , and CRP leads to sensitivity and negative predictive values that increase close to 90% in diagnosis early onset neonatal sepsis.³⁷ IL-6 mediates a systemic reaction to an inflammatory stimulus, that is, fever, leukocytosis, and the release of hepatic acute-phase proteins such as CRP, complement components, fibrinogen, and ferritin.³⁸

IL-8 is a proinflammatory cytokine and a potent neutrophil activating agent that is released from monocytes, endothelial cells, and neutrophils in response to IL-1 and TNF- α . IL-8 also regulates leukocyte migration. It is a frequently studied cytokine as a marker of neonatal sepsis. IL-8 peaks 2.5 to 3 hours

after stimulus. In one study, elevated levels of IL-8 predict organ failure in adults with septic shock.³⁹ Contrary to this, another study indicates that IL-8 is not a powerful biomarker in adults,⁴⁰ whereas some studies in children have shown that IL-8 can serve as an indicator of outcome in children with septic shock.⁴¹ IL-8 was identified as one of 34 genes that were increased in nonsurvivors relative to survivors and that was confirmed with involving genome-wide expression profiling in pediatric septic shock using RNA from blood samples supply within 24 hours of admission.⁴² A serum IL-8 level of 220 pg/mL or less measured within first 24 hours in children with septic shock may predict survival with probability of 95%. Also, IL-8 was increased in nonsurvivors relative to survivors, based on 28-day mortality in sepsis. This indicates that IL-8 elevation correlates with a more severe condition.⁴³ IL-8 can also be produced in placental or fetal cells in infectious started in the uterus⁴⁴ and, as IL-6, is a useful biomarker of early onset neonatal sepsis with sensitivity of 90% and specificity between 75 and 100%. The fact that serum concentration of IL-8 rises rapidly after an infection (within 2–4 hours) makes it useful as an early marker of sepsis.⁴⁵ In recent investigations, high value of IL-8 in a very low birth weight premature infants with clinical signs of early infection may be associated with the development of retinopathy.⁴⁶

IL-10 is an anti-inflammatory cytokine and is produced by monocytes, macrophages, natural killer (NK) cells, and B and T lymphocytes.⁴⁷ IL-10 suppresses the production of proinflammatory mediators, and in an experimental model, administration of recombinant murine IL-10 protects the mice from lethal endotoxemia.⁴⁸ IL-10 is structurally related to the IL-6 cytokine family, although its function is opposite. High IL-6:IL-10 ratio in sepsis was found in patients with a worse prognosis.⁴⁹ Recently, it was investigated that IL-10 might regulate the transition from early reversible sepsis to late irreversible septic shock and that polymorphisms in the IL-10 gene promoter affect sepsis susceptibility.⁵⁰

IL-12, initially called NK cell stimulatory factor or cytotoxic lymphocyte maturation factor, is a protein composed of two polypeptide subunits, p39 and p24. It plays a key role in the differentiation of Th1 and induces the production of IFN- γ . Recently, IL-12 was characterized as a major cytokine in the pathogenesis of gram-negative endotoxemia in mice.⁵¹ Also, IL-12 stimulates the differentiation of naive CD4⁺ T-cells and protects them from antigen-induced apoptotic death.⁵² In humans, a selective defect in preoperative monocyte IL-12 production disrupts the host defense against postoperative infections and increases the risk of lethal sepsis.⁵³ IL-12 was measured in newborns at the time when sepsis was first suspected clinically and was higher in patients with sepsis than in those without sepsis.⁵⁴

IL-18 is a proinflammatory cytokine produced by activated macrophages that are included in the induction of cell-mediated immunity. Elevated serum levels of IL-18 are associated with poor clinical outcome in severe sepsis. IL-18 may play an important role in the pathogenesis of idiopathic thrombocytopenic purpura. Several studies demonstrated that IL-18, as novel prognostic cytokine, is involved in severe sepsis with thrombocytopenia.⁵⁵ Also, a group of authors

reported that IL-18, as well as seven other serum markers, was elevated in preterm infants with infection,⁵⁶ whereas the other authors demonstrated that IL-18 had no diagnostic ability in neonates.⁵⁷

TGF- β is an important anti-inflammatory cytokine that plays a role in sepsis-induced immunosuppression, tissue repair, and fibrosis.⁵⁸ Except suppression of the release of proinflammatory mediators and stimulation production of immunosuppressive factors such as sTNFRs and IL-1Ra, TGF- β also inhibits IL-2 secretion and T-lymphocyte proliferation.⁵⁹ Recent data suggests that TGF- β may have cardioprotective effects with reverse the depression of myocardial contraction induced by proinflammatory cytokines in patients with septic shock.⁶⁰ Animal models showed that treatment with TGF- β blocked endotoxin improved survival in *Salmonella typhosa* endotoxin-induced septic shock.⁶¹ Notwithstanding, TGF- β levels were shown to peak early in disease, and they did not correlate strongly with severity of disease nor with the prognosis.⁶²

IL-4 is a cytokine with many immunoregulatory functions, and the most important is regulation of T lymphocyte differentiation.⁶³ IL-4 is a cytokine produced by lymphocytes and its important role is suppressing the secretion of monocyte-derived proinflammatory cytokines.⁶⁴ Although, studies suggest that IL-4 plays an important role in the pathogenesis of sepsis, its precise role is unknown. It was reported that in humans, the messenger RNA expression of IL-4 was associated with survival of patients with severe sepsis, but the plasma levels of IL-4 on the day of admission did not differ between survivors and nonsurvivors.⁶⁵

Recent studies investigated IL-3, a novel mediator that can potentiate inflammation in sepsis. In a mouse model of abdominal sepsis, it has been shown that innate response activator B cells produce IL-3, which induces myelopoiesis of Ly-6C (high) monocytes and neutrophils and originated a cytokine storm. High plasma IL-3 levels were associated with high mortality. Therefore, IL-3 is identified as an orchestrator of emergency myelopoiesis and can be a new therapeutic target for treating sepsis.⁶⁶

Other Biomarkers

CD64 is a neutrophil cell surface marker, known as Fc γ R1. It is a receptor on the neutrophil and its function is to bind the Fc portion of IgG (hence γ) antibodies. Those antibodies facilitate bacterial opsonization and phagocytosis. The level of CD64 is measured by the flow cytometric analysis of blood samples, and in pediatrics, CD64 has been investigated primarily to identify premature and term neonates with sepsis. Some authors showed that CD64 was elevated in children with documented infections, but was unable to distinguish between viral and bacterial infections. They also found that procalcitonin (PCT) was more specific and CRP was more sensitive than CD64.^{67,68} In a small group of older children, it has been shown that CD64 was able to distinguish between sepsis and SIRS better than CRP and PCT, especially when combined with the lipopolysaccharide-binding protein (LBS),⁶⁹ but much more investigation needs to be done to determine the clinical utility of this biomarker.⁷⁰ One of recent

studies confirmed CD64 utility as a good marker of bacterial sepsis since it was found to have mean sensitivity of 71% and a mean specificity of 87%.⁷¹

The expression of CD11b on neutrophils in sepsis may predict the development of organ failure and poor prognosis in patients with septic shock.⁷² One study demonstrated that CD11b expression has increased in some infants from the infection group up to 3 days before the onset of symptoms. This suggested that CD11b level may enable an early diagnosis of infection.⁷³ Some authors reported that neutrophil and monocyte CD11b expressions were significantly elevated in infected neonates. The neutrophil CD11b had a sensitivity of 66% and specificity of 71%, whereas monocyte CD11b had a sensitivity of 70% and specificity of 62% for detecting neonatal infection.⁷⁴ Also, CD15s is a potentially valuable biomarker of severe bacterial infection in infants.⁷¹

Lactate level is an important biomarker that can distinguish sepsis from septic shock and can predict the prognosis. Initially, serum lactate was recognized and used as an indicator of tissue hypoxia. Serum lactate levels rise when lactate production outstrips the body's ability to metabolize it. Also, an increase in lactate occurs when there is a decrease in its metabolic capacity, which is often seen in SIRS. The majority of researches, which found increased serum lactate in patients with sepsis, have been conducted on adults.^{75,76} It has been found that the patients with increased serum lactate in sepsis were sicker and had increased mortality, and it was also observed that patients whose lactate levels decreased with proper therapy had better outcomes.^{77,78} Related to this, lactate level was used as a diagnostic, monitoring, and prognostic biomarker. The serum level of lactate in septic children may identify a population at higher risk for severe outcomes. In children with SIRS, a single elevated lactate level increases risk of organ dysfunction several times. Since elevated lactate levels correspond with organ dysfunction and the need for resuscitative therapies, it may be an early indicator of resuscitation requirement in children.⁷⁹

CCL3 (also known as macrophage inflammatory protein-1 α) and CCL4 (also known as MIP-1 β) are small purification molecules that lead to increased infiltration of inflammatory cells. CCL3 belongs to the C-C chemokine family that is secreted by monocytes and macrophages and can be secreted by T cells. It is chemotactic and activates macrophages to induce secretion of TNF, IL-6, and IL-1 and it enhances killing as well. CCL3 can also be chemotactic for eosinophils, B and CD8 T cells. CCL4 (MIP-1 β) is primarily chemotactic for lymphocytes and monocytes. A single dose of lipopolysaccharide (LPS) given to healthy persons has been shown to cause an increase in both CCL3 and CCL4, which peaks approximately 2 hours after LPS is administered. Patients with bacterial meningitis have an increase of CCL4 in the cerebral spinal fluid.⁸⁰

LBP is mainly synthesized in the liver, but it can also be synthesized by epithelial and muscle cells as an acute-phase protein. It binds LPS of gram-negative bacteria to CD14 as well as TLRs, and modulates the microbial-induced activation of the inflammatory host response. Levels of LPB peak within 6 to 8 hours after an acute infection. It has better sensitivity and specificity for detecting sepsis than LPS-soluble, PCT, and CD14

complexes in early onset sepsis. But it is as equally effective as CRP in detecting sepsis in neonates older than 48 hours. Although LPB has a promising potential, further research is required for neonatal sepsis.⁸¹

Serum amyloid A (SAA) is an acute-phase protein regulated by the proinflammatory cytokines (IL-1, IL-6, TNF α). It is an apolipoprotein produced in the liver as well as derived from a variety of other tissues such as monocytes, endothelial cells, and smooth muscle cells. There is a significant increase in SAA levels from 8 to 24 hours after the onset of sepsis, and it was shown that SAA had better diagnostic accuracy than CRP in septic evaluation in neonatal early onset sepsis. Especially the des-arginine variant of SAA holds promise as a marker of acute and chronic inflammation.⁸²

IFN- γ -IP-10 is induced by IFN- γ in many types of cells including monocytes and lung epithelial cells. IP-10, also named CXCL10, is a potent chemokine for activated T lymphocytes and regulates cell proliferation, apoptosis, and adhesion molecule expression.⁸³ IP-10 level was higher in neonates with sepsis and necrotizing enterocolitis than in neonates who had only necrotizing enterocolitis.⁸⁴

High mobility group box 1 (HMGB1) is a chromatin protein localized in the nucleus and the cytoplasm. Cytokines TNF- α and IFN- γ stimulate macrophages to release HMGB1, while endotoxin induces late release of HMGB1. In animal models, HMGB1 can be detected in serum 8 hours after endotoxemia, and plateau levels are archived from 16 to 32 hours.⁸⁵ Antibodies (anti-HMGB1) can reduce endotoxin-induced acute lung injury and increase survival. In humans, HMGB1 were elevated in patients with surgical sepsis and in those with DIC. Also, it was significantly higher in septic nonsurvivors versus survivors.^{86,87}

Triggering receptor expressed on myeloid cells-1 (TREM-1) belongs to the immunoglobulin superfamily and stimulates the release of cytokines such as TNF- α and IL-1 β . Because it is not easily identified, its soluble form (sTREM) is a better option for a biomarker in sepsis. sTREM is higher in patients with septic shock and reaches peak levels at approximately 2 hours after infectious exposure. Its sustained elevation appears to predict a poor outcome. All this makes it an accurate marker of sepsis.^{88,89}

Several other reactants show potential and have been studied as biomarkers of sepsis. Some of these biomarkers are α -1 antitrypsin, fibronectin, lactoferrin, neopterin, and haptoglobin. Very few potential biomarkers reach the true biomarker stage. Although PCT is currently the most promising diagnostic biomarker for sepsis, recent evidence suggested that IL-8 can be used to stratify children with septic shock. Recently, great efforts have been made to develop a multibiomarker-based sepsis risk model for predicting illness severity and outcome for children with septic shock.⁹⁰ IL-8 and CCL4 are clinically appealing because of their relative simplicity, but both stratification biomarkers have insufficient positive predictive values, sensitivities, and specificities to develop a comprehensive pediatric septic shock stratification tool.^{41,91} The biological response during septic shock is exceedingly complex, and it is possible that a multibiomarker stratification strategy can more comprehensively meet these needs.⁹² Numerous acute-phase

proteins potentially may be biomarkers for sepsis, but none has been routinely used. The goal of the final model will be to predict illness severity and outcome of pediatric patients with septic shock. It will provide a decision-making and stratification tool for the care of children with septic shock.

Conclusion

Numerous studies have explained many different pathophysiological processes involved in sepsis and have revealed an important regulatory role of pro- and anti-inflammatory cytokines. But despite a great number of clinical studies, cytokines pathophysiology is still incompletely understood, and specific anticytokine treatments have not been successful in clinical trials. This is due to the fact that sepsis is a complex and dynamic process that involves excessive inflammatory and immune response. A small number of researches have been conducted in pediatric sepsis using multiple markers. The best approach to the diagnosis of sepsis is the combination of different biomarkers because the multiple indicators of infection can improve specificity and sensitivity of the whole assay.

References

- Rittirsch D, Flierl MA, Ward PA. Harmful molecular mechanisms in sepsis. *Nat Rev Immunol* 2008;8(10):776–787
- Wen AQ, Gu W, Wang J, et al. Clinical relevance of IL-1beta promoter polymorphisms (-1470, -511, and -31) in patients with major trauma. *Shock* 2010;33(06):576–582
- Zhang AQ, Pan W, Gao JW, et al. Associations between interleukin-1 gene polymorphisms and sepsis risk: a meta-analysis. *BMC Med Genet* 2014;15:8
- Watanabe E, Buchman TG, Hirasawa H, Zehnbauser BA. Association between lymphotoxin-alpha (tumor necrosis factor-beta) intron polymorphism and predisposition to severe sepsis is modified by gender and age. *Crit Care Med* 2010;38(01):181–193
- Schulte W, Bernhagen J, Bucala R. Cytokines in sepsis: potent immunoregulators and potential therapeutic targets—an updated view. *Mediators Inflamm* 2013;2013:165974
- Blackwell TS, Christman JW. Sepsis and cytokines: current status. *Br J Anaesth* 1996;77(01):110–117
- de Jong HK, van der Poll T, Wiersinga WJ. The systemic pro-inflammatory response in sepsis. *J Innate Immun* 2010;2(05):422–430
- Balk R, Roger C. Bone, MD and the evolving paradigms of sepsis. *Contrib Microbiol* 2011;17:1–11
- Cohen J. The immunopathogenesis of sepsis. *Nature* 2002;420(6917):885–891
- van der Poll T, van Deventer SJ. Cytokines and anticytokines in the pathogenesis of sepsis. *Infect Dis Clin North Am* 1999;13(02):413–426, ix
- Berner R, Niemeyer CM, Leititis JU, et al. Plasma levels and gene expression of granulocyte colony-stimulating factor, tumor necrosis factor-alpha, interleukin (IL)-1beta, IL-6, IL-8, and soluble intercellular adhesion molecule-1 in neonatal early onset sepsis. *Pediatr Res* 1998;44(04):469–477
- Parameswaran N, Patial S. Tumor necrosis factor- α signaling in macrophages. *Crit Rev Eukaryot Gene Expr* 2010;20(02):87–103
- Lewis M, Tartaglia LA, Lee A, et al. Cloning and expression of cDNAs for two distinct murine tumor necrosis factor receptors demonstrate one receptor is species specific. *Proc Natl Acad Sci U S A* 1991;88(07):2830–2834
- Shannon E, Noveck R, Sandoval F, Kamath B, Kearney M. Thalidomide suppressed interleukin-6 but not tumor necrosis factor-alpha in volunteers with experimental endotoxemia. *Transl Res* 2007;150(05):275–280
- Prashant A, Vishwanath P, Kulkarni P, et al. Comparative assessment of cytokines and other inflammatory markers for the early diagnosis of neonatal sepsis—a case control study. *PLoS One* 2013;8(07):e68426
- Bhatia M, Mochhala S. Role of inflammatory mediators in the pathophysiology of acute respiratory distress syndrome. *J Pathol* 2004;202(02):145–156
- van der Poll T, Büller HR, ten Cate H, et al. Activation of coagulation after administration of tumor necrosis factor to normal subjects. *N Engl J Med* 1990;322(23):1622–1627
- Schouten M, Wiersinga WJ, Levi M, van der Poll T. Inflammation, endothelium, and coagulation in sepsis. *J Leukoc Biol* 2008;83(03):536–545
- Silveira RC, Procianny RS. Evaluation of interleukin-6, tumor necrosis factor- α and interleukin-1 β for early diagnosis of neonatal sepsis. *Acta Paediatr* 1999;88(06):647–650
- Duncombe G, Veldhuizen RA, Gratton RJ, Han VK, Richardson BS. IL-6 and TNFalpha across the umbilical circulation in term pregnancies: relationship with labour events. *Early Hum Dev* 2010;86(02):113–117
- O'Neill LA. The interleukin-1 receptor/Toll-like receptor superfamily: 10 years of progress. *Immunol Rev* 2008;226(01):10–18
- Smith K, Bigham MT. Biomarkers in pediatric sepsis. *Open Inflamm J* 2011;4:24–30
- Loisa P, Rinne T, Laine S, Hurme M, Kaukinen S. Anti-inflammatory cytokine response and the development of multiple organ failure in severe sepsis. *Acta Anaesthesiol Scand* 2003;47(03):319–325
- Okusawa S, Gelfand JA, Ikejima T, Connolly RJ, Dinarello CA. Interleukin 1 induces a shock-like state in rabbits. Synergism with tumor necrosis factor and the effect of cyclooxygenase inhibition. *J Clin Invest* 1988;81(04):1162–1172
- Dinarello CA. Infection, fever, and exogenous and endogenous pyrogens: some concepts have changed. *J Endotoxin Res* 2004;10(04):201–222
- Conte D, Holcik M, Lefebvre CA, et al. Inhibitor of apoptosis protein cIAP2 is essential for lipopolysaccharide-induced macrophage survival. *Mol Cell Biol* 2006;26(02):699–708
- Shimaoka M, Park EJ. Advances in understanding sepsis. *Eur J Anaesthesiol Suppl* 2008;42(42):146–153
- Gårdlund B, Sjölin J, Nilsson A, Roll M, Wickerts CJ, Wretling B. Plasma levels of cytokines in primary septic shock in humans: correlation with disease severity. *J Infect Dis* 1995;172(01):296–301
- Ohlsson K, Björk P, Bergenfeldt M, Hageman R, Thompson RC. Interleukin-1 receptor antagonist reduces mortality from endotoxin shock. *Nature* 1990;348(6301):550–552
- Choy E. Understanding the dynamics: pathways involved in the pathogenesis of rheumatoid arthritis. *Rheumatology (Oxford)* 2012;51(05, Suppl 5):v3–v11
- Kapoor M, Martel-Pelletier J, Lajeunesse D, Pelletier JP, Fahmi H. Role of proinflammatory cytokines in the pathophysiology of osteoarthritis. *Nat Rev Rheumatol* 2011;7(01):33–42
- Jawień J. New insights into immunological aspects of atherosclerosis. *Pol Arch Med Wewn* 2008;118(03):127–131
- Park JY, Pillinger MH. Interleukin-6 in the pathogenesis of rheumatoid arthritis. *Bull NYU Hosp Jt Dis* 2007;65(01, Suppl 1):S4–S10
- Scheller J, Rose-John S. Interleukin-6 and its receptor: from bench to bedside. *Med Microbiol Immunol (Berl)* 2006;195(04):173–183
- Kishimoto T. The biology of interleukin-6. *Blood* 1989;74(01):1–10
- Tasci Y, Dilbaz B, Uzmez Onal B, et al. The value of cord blood interleukin-6 levels for predicting chorioamnionitis, funisitis and neonatal infection in term premature rupture of membranes. *Eur J Obstet Gynecol Reprod Biol* 2006;128(1-2):34–39

- 37 Remick DG, Bolgos G, Copeland S, Siddiqui J. Role of interleukin-6 in mortality from and physiologic response to sepsis. *Infect Immun* 2005;73(05):2751–2757
- 38 Bloos F, Reinhart K. Rapid diagnosis of sepsis. *Virulence* 2014; 5(01):154–160
- 39 Takala A, Jousela I, Jansson SE, et al. Markers of systemic inflammation predicting organ failure in community-acquired septic shock. *Clin Sci (Lond)* 1999;97(05):529–538
- 40 Calfee CS, Thompson BT, Parsons PE, Ware LB, Matthay MA, Wong HR. Plasma interleukin-8 is not an effective risk stratification tool for adults with vasopressor-dependent septic shock. *Crit Care Med* 2010;38(06):1436–1441
- 41 Wong HR, Cvijanovich N, Wheeler DS, et al. Interleukin-8 as a stratification tool for interventional trials involving pediatric septic shock. *Am J Respir Crit Care Med* 2008;178(03):276–282
- 42 Wong HR, Shanley TP, Sakthivel B, et al. ; Genomics of Pediatric SIRS/Septic Shock Investigators. Genome-level expression profiles in pediatric septic shock indicate a role for altered zinc homeostasis in poor outcome. *Physiol Genomics* 2007;30(02): 146–155
- 43 Standage SW, Wong HR. Biomarkers for pediatric sepsis and septic shock. *Expert Rev Anti Infect Ther* 2011;9(01):71–79
- 44 Fan Y, Yu JL. Umbilical blood biomarkers for predicting early-onset neonatal sepsis. *World J Pediatr* 2012;8(02):101–108
- 45 Bhandari V. Effective biomarkers for diagnosis of neonatal sepsis. *J Pediatric Infect Dis Soc* 2014;3(03):234–245
- 46 Silveira RC, Fortes Filho JB, Procianny RS. Assessment of the contribution of cytokine plasma levels to detect retinopathy of prematurity in very low birth weight infants. *Invest Ophthalmol Vis Sci* 2011;52(03):1297–1301
- 47 Jones LL, Vignali DA. Molecular interactions within the IL-6/IL-12 cytokine/receptor superfamily. *Immunol Res* 2011;51(01):5–14
- 48 Howard M, Muchamuel T, Andrade S, Menon S. Interleukin 10 protects mice from lethal endotoxemia. *J Exp Med* 1993;177(04): 1205–1208
- 49 Ng PC, Lam HS. Biomarkers for late-onset neonatal sepsis: cytokines and beyond. *Clin Perinatol* 2010;37(03):599–610
- 50 Zeng L, Gu W, Chen K, et al. Clinical relevance of the interleukin 10 promoter polymorphisms in Chinese Han patients with major trauma: genetic association studies. *Crit Care* 2009;13(06):R188
- 51 Hazelzet JA, Kornelisse RF, van der Pouw Kraan TC, et al. Interleukin 12 levels during the initial phase of septic shock with purpura in children: relation to severity of disease. *Cytokine* 1997;9(09):711–716
- 52 Estaquier J, Idziorek T, Zou W, et al. T helper type 1/T helper type 2 cytokines and T cell death: preventive effect of interleukin 12 on activation-induced and CD95 (FAS/APO-1)-mediated apoptosis of CD4+ T cells from human immunodeficiency virus-infected persons. *J Exp Med* 1995;182(06):1759–1767
- 53 Weighardt H, Heidecke CD, Westerholt A, et al. Impaired monocyte IL-12 production before surgery as a predictive factor for the lethal outcome of postoperative sepsis. *Ann Surg* 2002;235(04): 560–567
- 54 Sherwin C, Broadbent R, Young S, et al. Utility of interleukin-12 and interleukin-10 in comparison with other cytokines and acute-phase reactants in the diagnosis of neonatal sepsis. *Am J Perinatol* 2008;25(10):629–636
- 55 Cui YL, Wang B, Gao HM, et al. Interleukin-18 and miR-130a in severe sepsis patients with thrombocytopenia. *Patient Prefer Adherence* 2016;10:313–319
- 56 Kingsmore SF, Kennedy N, Halliday HL, et al. Identification of diagnostic biomarkers for infection in premature neonates. *Mol Cell Proteomics* 2008;7(10):1863–1875
- 57 Bender L, Thaarup J, Varming K, Krarup H, Ellermann-Eriksen S, Ebbesen F. Early and late markers for the detection of early-onset neonatal sepsis. *Dan Med Bull* 2008;55(04):219–223
- 58 Globe GC, Schiemann WP, Lodish HF. Role of transforming growth factor β in human disease. *N Engl J Med* 2000;342(18):1350–1358
- 59 Wan YY, Flavell RA. TGF- β and regulatory T cell in immunity and autoimmunity. *J Clin Immunol* 2008;28(06):647–659
- 60 Kumar A, Kumar A, Paladugu B, Mensing J, Parrillo JE. Transforming growth factor- β 1 blocks in vitro cardiac myocyte depression induced by tumor necrosis factor- α , interleukin-1 β , and human septic shock serum. *Crit Care Med* 2007;35(02):358–364
- 61 Perrella MA, Hsieh CM, Lee WS, et al. Arrest of endotoxin-induced hypotension by transforming growth factor beta1. *Proc Natl Acad Sci U S A* 1996;93(05):2054–2059
- 62 Knapp S, Thalhammer F, Locker GJ, et al. Prognostic value of MIP-1 α , TGF- β 2, sELAM-1, and sVCAM-1 in patients with gram-positive sepsis. *Clin Immunol Immunopathol* 1998;87(02):139–144
- 63 Seder RA, Paul WE, Davis MM, Fazekas de St Groth B. The presence of interleukin 4 during in vitro priming determines the lymphokine-producing potential of CD4+ T cells from T cell receptor transgenic mice. *J Exp Med* 1992;176(04):1091–1098
- 64 Opal SM, DePalo VA. Anti-inflammatory cytokines. *Chest* 2000; 117(04):1162–1172
- 65 Wu HP, Wu CL, Chen CK, et al. The interleukin-4 expression in patients with severe sepsis. *J Crit Care* 2008;23(04):519–524
- 66 Weber GF, Chousterman BG, He S, et al. Interleukin-3 amplifies acute inflammation and is a potential therapeutic target in sepsis. *Science* 2015;347(6227):1260–1265
- 67 Lam HS, Wong SP, Cheung HM, et al. Early diagnosis of intra-abdominal inflammation and sepsis by neutrophil CD64 expression in newborns. *Neonatology* 2011;99(02):118–124
- 68 Rudensky B, Sirota G, Erlichman M, Yinnon AM, Schlesinger Y. Neutrophil CD64 expression as a diagnostic marker of bacterial infection in febrile children presenting to a hospital emergency department. *Pediatr Emerg Care* 2008;24(11):745–748
- 69 Groselj-Grenc M, Ihan A, Pavcnik-Arnol M, Kopitar AN, Gmeiner-Stopar T, Derganc M. Neutrophil and monocyte CD64 indexes, lipopolysaccharide-binding protein, procalcitonin and C-reactive protein in sepsis of critically ill neonates and children. *Intensive Care Med* 2009;35(11):1950–1958
- 70 Cid J, Aguinaco R, Sánchez R, García-Pardo G, Llorente A. Neutrophil CD64 expression as marker of bacterial infection: a systematic review and meta-analysis. *J Infect* 2010;60(05): 313–319
- 71 Markic J, Jeroncic A, Polancec D, et al. CD15s is a potential biomarker of serious bacterial infection in infants admitted to hospital. *Eur J Pediatr* 2013;172(10):1363–1369
- 72 Muller Kobold AC, Tulleken JE, Zijlstra JG, et al. Leukocyte activation in sepsis; correlations with disease state and mortality. *Intensive Care Med* 2000;26(07):883–892
- 73 Turunen R, Andersson S, Nupponen I, Kautiainen H, Siitonen S, Repo H. Increased CD11b-density on circulating phagocytes as an early sign of late-onset sepsis in extremely low-birth-weight infants. *Pediatr Res* 2005;57(02):270–275
- 74 Genel F, Atlihan F, Gulez N, et al. Evaluation of adhesion molecules CD64, CD11b and CD62L in neutrophils and monocytes of peripheral blood for early diagnosis of neonatal infection. *World J Pediatr* 2012;8(01):72–75
- 75 Mikkelsen ME, Miltiades AN, Gaieski DF, et al. Serum lactate is associated with mortality in severe sepsis independent of organ failure and shock. *Crit Care Med* 2009;37(05):1670–1677
- 76 Jansen TC, van Bommel J, Bakker J. Blood lactate monitoring in critically ill patients: a systematic health technology assessment. *Crit Care Med* 2009;37(10):2827–2839
- 77 Arnold RC, Shapiro NI, Jones AE, et al; Emergency Medicine Shock Research Network (EMShockNet) Investigators. Multicenter study of early lactate clearance as a determinant of survival in patients with presumed sepsis. *Shock* 2009;32(01):35–39
- 78 Jones AE, Shapiro NI, Trzeciak S, Arnold RC, Claremont HA, Kline JA; Emergency Medicine Shock Research Network (EMShockNet) Investigators. Lactate clearance vs central venous oxygen saturation as goals of early sepsis therapy: a randomized clinical trial. *JAMA* 2010;303(08):739–746

- 79 Scott HF, Donoghue AJ, Gieski DF, Marchese RF, Mistry RD. The utility of early lactate testing in undifferentiated pediatric systemic inflammatory response syndrome. *Acad Emerg Med* 2012; 19(11):1276–1280
- 80 Alder MN, Lindsell CJ, Wong HR. The pediatric sepsis biomarker risk model: potential implications for sepsis therapy and biology. *Expert Rev Anti Infect Ther* 2014;12(07):809–816
- 81 Pavcnik-Arnol M, Hojker S, Derganc M. Lipopolysaccharide-binding protein, lipopolysaccharide, and soluble CD14 in sepsis of critically ill neonates and children. *Intensive Care Med* 2007;33(06):1025–1032
- 82 Arnon S, Litmanovitz I, Regev RH, Bauer S, Shainkin-Kestenbaum R, Dolfin T. Serum amyloid A: an early and accurate marker of neonatal early-onset sepsis. *J Perinatol* 2007;27(05):297–302
- 83 Neville LF, Mathiak G, Bagasra O. The immunobiology of interferon-gamma inducible protein 10 kD (IP-10): a novel, pleiotropic member of the C-X-C chemokine superfamily. *Cytokine Growth Factor Rev* 1997;8(03):207–219
- 84 Ng PC, Li K, Chui KM, et al. IP-10 is an early diagnostic marker for identification of late-onset bacterial infection in preterm infants. *Pediatr Res* 2007;61(01):93–98
- 85 Sama AE, D'Amore J, Ward MF, Chen G, Wang H. Bench to bedside: HMGB1—a novel proinflammatory cytokine and potential therapeutic target for septic patients in the emergency department. *Acad Emerg Med* 2004;11(08):867–873
- 86 Yang H, Ochani M, Li J, et al. Reversing established sepsis with antagonists of endogenous high-mobility group box 1. *Proc Natl Acad Sci U S A* 2004;101(01):296–301
- 87 Hatada T, Wada H, Nobori T, et al. Plasma concentrations and importance of High Mobility Group Box protein in the prognosis of organ failure in patients with disseminated intravascular coagulation. *Thromb Haemost* 2005;94(05):975–979
- 88 Routsis C, Giamarellos-Bourboulis EJ, Antonopoulou A, et al. Does soluble triggering receptor expressed on myeloid cells-1 play any role in the pathogenesis of septic shock? *Clin Exp Immunol* 2005; 142(01):62–67
- 89 Gibot S, Cravoisy A, Kolopp-Sarda MN, et al. Time-course of sTREM (soluble triggering receptor expressed on myeloid cells)-1, procalcitonin, and C-reactive protein plasma concentrations during sepsis. *Crit Care Med* 2005;33(04):792–796
- 90 Kaplan JM, Wong HR. Biomarker discovery and development in pediatric critical care medicine. *Pediatr Crit Care Med* 2011; 12(02):165–173
- 91 Nowak JE, Wheeler DS, Harmon KK, Wong HR. Admission chemokine (C-C motif) ligand 4 levels predict survival in pediatric septic shock. *Pediatr Crit Care Med* 2010;11(02): 213–216
- 92 Wong HR, Cvijanovich N, Lin R, et al. Identification of pediatric septic shock subclasses based on genome-wide expression profiling. *BMC Med* 2009;7:34