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PROPERTIES OF PHOSPHATIDYLINOSITOL KINASE IN HUMAN PLATELETS.  
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We have reported the specific <sup>32</sup>P-labelling in phosphatidyl-  
inositol-4-monophosphate (PIP) of intact platelets upon addition  
of the agents which elevate intracellular cAMP (Thrombos. Res. 44,  
155, 1986). This event may be catalyzed by the action of PI-kinase,  
the properties of which has not been elucidated yet. Thereby,  
attempts were made to assay and to characterize PI-kinase of  
human platelets. Fresh lysed platelets prelabelled with <sup>32</sup>P in  
cold Tris-HCl buffer containing 2mM EGTA were incubated at 37 C  
in the presence of MgCl<sub>2</sub> for designated times and the phospho-  
lipids were extracted and analyzed by thin layer chromatography.  
<sup>32</sup>P-labelling in PIP was gradually increased in consort with the  
decreased labelling in PI-4,5-bisphosphate. As the changes in the  
labelling was not affected by the presence of apyrase and as the  
radioactive inositol trisphosphate was not detected, it was  
suggested that the changes is due to the action of phosphomono-  
esterase rather than PI-kinase or phospholipase C. When <sup>32</sup>P-ATP  
was added to non-labelled lysed platelets upon incubation, <sup>32</sup>P was  
labelled only into PIP and the amount was markedly increased  
until 5min. after incubation. Since the labelling was strongly  
inhibited by apyrase, it likely reflects the activity of PI-kinase.  
The activity of PI-kinase thus measured required Mg<sup>2+</sup> strictly  
for the activity and the maximal activity was obtained in the  
presence of 30mM Mg<sup>2+</sup>. In contrast, it was markedly inhibited in  
the presence of Ca<sup>2+</sup> (as low as 2mM Ca<sup>2+</sup> in the presence of 2mM  
EGTA), which was compatible with our previous findings with intact  
platelets. The activity of A-kinase was not inhibited by a low  
concentration of Ca<sup>2+</sup>. Furthermore, the activity was inhibited by  
cAMP or dbcAMP in a dose related manner and no enhancement of the  
activity was obtained by the addition of catalytic subunit of  
A-kinase, though a significant reduction in the activity was  
observed in the presence of inhibitor protein to A-kinase. From  
these observations, the following conclusions were obtained; 1) The  
activity of PI-kinase in lysed platelets may be determined by  
pulse labelling with <sup>32</sup>P-ATP. 2) It requires Mg<sup>2+</sup> absolutely and  
is inhibited by a very low concentration of Ca<sup>2+</sup>. 3) PI-kinase is  
activated by A-kinase but the activated enzyme is inhibited by  
cAMP, suggesting the presence of feedback mechanism.

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PLATELET-FIBRIN CLOTS FORMED BY THROMBIN SELECTIVELY RETAIN  
PHOSPHATIDYLINOSITOL 4,5-BISPHOSPHATE (PIP<sub>2</sub>). John D. Vickers,  
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It is established that stimulation of human platelets with  
thrombin for 60 s in the absence of fibrinogen increases the  
amount of PIP<sub>2</sub> compared with unstimulated controls (4.7 ± 0.24  
nmol/10<sup>9</sup> plat. vs 3.83 ± 0.14 nmol/10<sup>9</sup> plat., p<0.01, n=8). How-  
ever, stimulation with thrombin for 60 s in the presence of  
fibrinogen causes a large decrease in the amount of PIP<sub>2</sub> that  
can be extracted with acidified chloroform/methanol compared  
with unstimulated controls (1.62 ± 0.39 nmol/10<sup>9</sup> plat. vs 3.84 ±  
0.44 nmol/10<sup>9</sup> plat., p<0.001, n=6). Stimulation of rabbit plate-  
lets with thrombin in the presence of fibrinogen also decreases  
the amount of extractable PIP<sub>2</sub> (60% at 60 s, p<0.001, n=8).  
Similar decreases in amount can not be demonstrated for phospha-  
tidylinositol 4-phosphate, phosphatidylinositol, phosphatidic  
acid or phosphatidylcholine under the same conditions, indicat-  
ing that the decrease is specific for PIP<sub>2</sub>. With rabbit plate-  
lets, polymerized fibrin formed by reptilase, which does not  
stimulate platelets or induce clot retraction, does not cause  
the decrease in extractable PIP<sub>2</sub> (3.06 ± 0.05 nmol/10<sup>9</sup> plat.  
were extracted compared with 3.18 ± 0.07 nmol/10<sup>9</sup> plat. without  
reptilase). However, stimulation of rabbit platelets with ADP in  
the presence of polymerizing fibrin formed by reptilase causes a  
larger decrease in extractable PIP<sub>2</sub> (to 2.54 ± 0.19 nmol/10<sup>9</sup>  
plat., p<0.05, n=4) than is caused by ADP and fibrinogen alone  
(to 2.87 ± 0.06 nmol/10<sup>9</sup> plat., p<0.05, n=4). Inhibition by  
glycyl-L-prolyl-L-arginyl-L-proline of polymerization of fibrin  
formed by the action of thrombin prevents the large decrease in  
the amount of extractable PIP<sub>2</sub> (4.37 ± 0.30 nmol/10<sup>9</sup> plat. were  
extracted) from human platelets. These results indicate that the  
interaction of polymerizing fibrin with stimulated platelets is  
required for the decrease in PIP<sub>2</sub>. The decrease in extractable  
PIP<sub>2</sub> seen with polymerizing fibrin can not be explained by in-  
creased degradation of PIP<sub>2</sub> to IP<sub>3</sub> or PIP. Thus, when human or  
rabbit platelets are stimulated with thrombin in the presence of  
fibrinogen, an association of polymerizing fibrin with the stimu-  
lated platelets occurs that leads to decreased extractability  
of PIP<sub>2</sub>. This may mean that PIP<sub>2</sub> forms a specific association  
with platelet proteins that are involved in clot retraction.

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TURNOVER OF THE PHOSPHOMONOESTER GROUPS OF POLYPHOSPHO-  
INOSITIDES IN UNSTIMULATED HUMAN PLATELETS.  
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The metabolic activity of the polyphosphoinositides in  
unstimulated human platelets was studied by 1) short-term  
labelling with <sup>32</sup>P-P<sub>1</sub>, 2) by replacement of <sup>32</sup>P-  
P<sub>1</sub> from pre-labelled platelets with unlabelled phosphate  
and 3) by depriving the cells of metabolic ATP. Under  
short-term labelling conditions, the 4- and 5-phosphates of  
phosphatidylinositol-4-phosphate (PIP) and phosphatidylinositol  
4,5-bisphosphate (PIP<sub>2</sub>) had the same specific radioactivity  
as the γ-phosphate of metabolic ATP. The specific radioactivity  
of the 1-phosphates of phosphatidylinositol, PIP and PIP<sub>2</sub>,  
was similar, but only 4-13 % as compared to the γ-phosphate of  
ATP. When <sup>32</sup>P-P<sub>1</sub> pre-labelled platelets were incubated  
with up to 25 mM of unlabelled phosphate, the displacement of  
the <sup>32</sup>P-label from PIP, PIP<sub>2</sub> and metabolic ATP followed  
similar kinetics. Inhibition of ATP regeneration in  
<sup>32</sup>P-P<sub>1</sub> pre-labelled platelets resulted in a rapid fall  
in <sup>32</sup>P-metabolic ATP with much slower fall in <sup>32</sup>P-PIP<sub>2</sub>.  
<sup>32</sup>P-PIP increased initially and decreased thereafter in  
parallel with PIP<sub>2</sub>. However, ATP turnover was not  
abolished, as indicated by the marked (25 % of the control)  
incorporation of extracellular <sup>32</sup>P-P<sub>1</sub> into PIP and  
PIP<sub>2</sub> in metabolically inhibited platelets. This low  
phosphate turnover may explain the relative resistance of PIP  
and PIP<sub>2</sub> to metabolic inhibition.  
We conclude that PIP and PIP<sub>2</sub> are present as a single  
metabolic pool in human platelets. Turnover of the 4- and  
5-phosphates of PIP and PIP<sub>2</sub> in unstimulated platelets is  
as rapid as that of the γ-phosphate of metabolic ATP, and  
accounts for about 7 % of basal ATP consumption.

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EVIDENCE THAT ABNORMAL PLATELET AGGREGATION IN SPONTANEOUSLY  
HYPERTENSIVE RATS IS LINKED WITH PHOSPHOINOSITIDES TURNOVER AND  
PHOSPHORYLATION OF 47,000 DALTON PROTEIN. Huzoor-Akbar and  
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We have shown earlier that abnormal platelet aggregation in  
spontaneously hypertensive rats (SHR) is not caused by  
prostaglandins (Thromb. Res. 41, 555-566, 1986). In this study  
platelets from SHR and normotensive (Wistar Kyoto, WKY) rats  
were used to examine the role of phosphoinositides (PIs) and  
protein phosphorylation in increased platelet activation in  
hypertension. Thrombin (0.05 U/ml) induced rapid hydrolysis of  
phosphatidylinositol-4,5-bis-phosphate (PIP<sub>2</sub>), phosphatidyl-  
inositol-4-phosphate (PIP), and phosphatidylinositol (PI) in  
(<sup>32</sup>P)-PO<sub>4</sub> labeled platelets. However, significantly greater  
hydrolysis of PIP<sub>2</sub> and PI was seen in SHR platelets than in WKY  
platelets (see Table). Thrombin also caused two- to three-fold  
increased accumulation of phosphatidic acid (PA) in SHR  
platelets than in WKY platelets (see Table).

	% CHANGE IN	TIME (SECONDS)				
		3	5	15	30	240
PIP <sub>2</sub>	WKY	-15±3	-13±5	-15±4	-4±5	+7±7
	SHR	-23±4	-32±2	-25±4	-8±5	+15±7
PIP	WKY	-14±3	-8±3	-4±6	+3±4	+7±4
	SHR	-18±3	+7±5	+5±3	+20±4	+22±5
PI	WKY	-4±4	-13±4	-8±4	-1±3	+9±5
	SHR	-7±4	-12±3	-24±7	-5±4	+0±4
PA	WKY		+8±2	+28±12	+42±6	+106±35
	SHR		+19±10	+89±9	+134±25	+219±39

Thrombin caused phosphorylation of 18,000 Dalton (P18) and 47,000  
Dalton (P47) proteins in SHR and WKY Platelets. Significantly  
increased phosphorylation of P47 was seen at 5, 15, 60 and 240  
seconds of incubation with thrombin in SHR platelets (60%, 68%,  
98% and 91%) than in WKY platelets (13%, 37%, 44% and 47%). The  
extent of P18 phosphorylation was same in both SHR and WKY  
platelets. Aspirin (500 μM) did not affect phosphorylation of  
P47 or P18 in SHR or WKY Platelets. These data lead us to  
suggest that increased turnover of PIs and increased  
phosphorylation of P47 are involved in abnormal platelet  
aggregation in SHR (Supported in part by the COHC grant #86-01-A  
and the Ohio University College of Osteopathic Medicine).