A. H. Drummond and J. L. Gordon (University Department of Pathology, Cambridge, CB2 1QP, England): Receptors for 5-Hydroxytryptamine on Rat Blood Platelets. (287)

When 5-hydroxytryptamine (5HT) or its analogue, 5-methoxy- α -methyltryptamine (5MO α MT) are added to rat citrated platelet-rich plasma (PRP), the platelets change in shape but do not aggregate. The response to both of these agents is inhibited by the 5HT antagonist, cinanserin (IC₅₀ = 3×10^{-9} M). Cinanserin is at least 10,000 times less potent against the active uptake of 5HT (IC₅₀ > 5×10^{-5} M). 5MO α MT is not actively transported by the platelet, although some instantaneous binding can be measured which is independent of temperature (4°-37°). 5MO α MT does not inhibit 5HT uptake over the concentration range at which it induces the shape change ($10^{-8}-10^{-5}$ M). Binding of (³H)-5HT to rat platelets at 4° indicates the presence of three binding sites, one of which is specifically blocked by cinanserin (IC₅₀ = 2.8×10^{-9} M). Close correlation between the inhibitory potency of various drugs against (³H)-5HT binding and 5HT-induced shape change suggests that this site is the 5HT receptor on the platelet which initiates the shape change. Our results indicate that 5HT induces the platelet shape change by combination with a specific cinanserinsensitive 5HT receptor, which is unconnected with the uptake site.

A. Hawiger, J. Hawiger, S. Timmons, S. Steckley and C. Cheng (Vanderbilt University School of Medicine and VA Hospital, Nashville TN., U.S.A.): The Role of Membrane Receptors in Pathways of Human Platelet Activation by Endotoxin and other Microbial Products. (288)

Endotoxin-producing gram-negative rods cause thrombocytopenia and contribute to shock and death of 25% of bacteremic patients. Cellular pathways of clot-promoting activity of endotoxin compared with other microbial products were studied. Membrane receptor for endotoxin was demonstrated using human platelets as target cells separated from plasma proteins by albumin gradient-Sepharose 2B gel filtration. Endotoxin receptor was distinct from thrombin-sensitive protein and from IgG Fc receptor. It was resistant to neuraminidase and trypsin. It was related to membrane hydrophobic region(s) as demonstrated with fluorescent probe. Endotoxin-receptor interaction triggered unmasking of platelet procoagulant (Platelet Factor 3) and selective release of ^aH-serotonin. A similar receptor was found for histoplasmin (yeast antigen). In contrast, staphylococcal protein A (cell wall antigen) required IgG Fc fragment and complement for its interaction with platelet membrane.

Thus in addition to the well known indirect pathway of activation of human platelets by microbial antigens through receptors for IgG Fc fragment and complement there is a direct pathway of activation due to the existence of specific antigen receptors on human platelets.

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N. O. Solum (Institute for Thrombosis Research, Rikshospitalet, Oslo, Norway): Effects of Bovine Factor VIII-Related Protein on Human Platelets and Isolated Human Platelet Membranes. (289)

The first phase of aggregation of human platelets induced by bovine factor VIIIrelated protein does not require an intact energy metabolism and probably represents a passive agglutination phenomenon due to receptors for the protein on the outside of the platelet cytoplasmic membrane. However, washed human platelets lost their ability to be aggregated by bovine factor VIII-related protein after freezing and thawing of the platelets. Furthermore, isolated human platelet membranes were not flocculated by the bovine protein in the absence of added calcium ions. Additions of calcium chloride (2.1 mM) alone flocculated the isolated membranes. The membranes used represented the material sedimenting between 10,000 and 100,00 g from a homogenate obtained by homogenizing washed platelets suspended in 0.27 M sucrose in an Aminco-French pressure cell (1.361 atm. 2×1 min). Subsequent sucrose density gradient centrifugation of the preparations did not reveal bands of particulate matter at a higher sucrose density than 1.12. In

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contrast, freezing and thawing of formalin-treated human platelets (4 mg/ml formaldehyde) did not destroy their ability to be aggregated by bovine factor VIII-related protein. Application of the method described above to the formalin-treated platelets resulted in a low yield of 10,000–100,000 g sedimenting material and to more particulate bands on subsequent sucrose density gradient centrifugation. The stabilizing effect of the formalin treatment (through protein polymerization?) may also be seen from the fact that such platelets are aggregated by bovine factor VIII-related protein even after their ability to aggregate with thrombin has been lost.

H. D. Kaulen and R. Gross (Medizinische Universitätsklinik, 5 Köln 41, Deutschland): Platelet Phospholipid Synthesis During Thrombin-Induced Platelet Aggregation. (290)

To study the effect of aggregation and release on platelet membrane metabolism human platelet phospholipids were labeled with either glycerol-C 14 or P 32. After washing off the residual label in order to avoid interference of aggregation with uptake, the changes in the distribution of the label in the various phospholipid classes were followed during the first four minutes of thrombin-induced aggregation. Glycerol-C 14 was incorporated into all glycerophospholipid synthesis, and this incorporation was not changed by aggregation. P 32, labeling preferentially the phosphoinositides, was rapidly incorporated over non-aggregated controls in the course of aggregation from water-soluble precursors into phosphatidic acid, di- and triphosphoinositide. As no new de-novo synthesis is taking place it is concluded that phospholipid phosphokinase reactions at the platelet membrane are activated with thrombin aggregation. Accordingly, phosphoinositide kinases have been demonstrated in isolated platelet membranes.

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A.-B. Otnæss (Institute of Medical Biology, University of Tromsø, Tromsø, Norway): Effect of Phospholipase C on Human Platelets. (291)

The effect on human platelets of phospholipase C (*Bacillus cereus*) has been studied. Platelets prepared by gel filtration lost 20-30% of their phospholipids when incubated with phospholipase C for 20 min. Phosphatidylethanolamine (PE) was reduced by about 50%, whereas phosphatidylcholine and phosphatidylserine were reduced each by about 20%. Sphingomyelin was not reduced.

These data suggest an asymmetrical distribution of phospholipids in the platelet membrane, PE being more accessible and therefore probably mainly located in the outer part of the membrane.

The loss of phospholipids was not accompanied by aggregation, nor did the platelets lose their ability to aggregate with thrombin or ADP. Data on release of serotonin, platelet factor 3 and 4 and scanning electron micrographs of treated platelets will be given.

P. C. French, J. J. Sixma and H. Holmsen (Rikshospitalet, Oslo 1, Norway): The Uptake of Adenine by Isolated Human Platelet Membranes. (292)

Adenine uptake into isolated platelet membranes had about the same Km $(181 \pm 21.9 \text{ nM})$ as uptake into intact platelets $(151 \pm 21 \text{ nM})$ and was also competitively inhibited by papaverine and hypoxanthine. No uptake occured at 0° and accumulated adenine was converted to AMP. AMP was not bound to protein as judged by chromatography of triton X-100 solubilized membranes chromatographed on Sephades G-25. The pH optimum for adenine uptake was at pH 5.5. Exogenous 5-phosphoribosyl-1-pyrophosphate stimulated uptake.

These data may be explained by uptake of adenine by facilitated diffusion followed by conversion to AMP by adenine phosphoribosyltransferase (E. C. 2.4.2.7.) but group translocation cannot be entirely excluded.