Xerostomia Therapy Due to Ionized Radiation Using Preconditioned Bone Marrow-Derived Mesenchymal Stem Cells

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Objectives The aim of this study was to describe the process of regeneration of damaged salivary glands due to ionizing radiations by bone marrow mesenchymal stem cells (BM-MSCs) transplantation that have been given hypoxic preconditioning with 1% O₂ concentration.

Materials and Methods Stem cell culture was performed under normoxic (O₂: 21%) and hypoxic conditions by incubating the cells for 48 hours in a low oxygen tension chamber consisting of 95% N₂, 5% CO₂, and 1% O₂. Thirty male Wistar rats were divided into four groups: two groups of control and two groups of treatment. A single dose of 15 Gy radiation was provided to the ventral region of the neck in all treatment groups, damaging the salivary glands. BM-MSCs transplantation was performed in the treatment groups for normoxia and hypoxia 24-hour postradiation.

Statistical Analysis Statistical analysis was done using normality test, followed by MANOVA test (p < 0.05).

Results There was a significant difference in the expression of binding SDF1-CXCR4, Bcl-2 (p < 0.05) and also the activity of the enzyme α-amylase in all groups of hypoxia.

Conclusion BM-MSCs transplantation with hypoxic precondition increases the expression of binding SDF1-CXCR4, Bcl-2 that contributes to cell migration, cell survival, and cell differentiation.

Keywords ► bone marrow mesenchymal stem cells  ► hypoxic precondition  ► salivary gland defect  ► SDF1-CXCR4  ► Bcl-2  ► α-amylase

Introduction Salivary gland is one of the normal tissues frequently affected by the head and neck radiation therapy. Irreversible salivary gland defect frequently occurs due to the radiation exposure. The salivary gland damage results in the decrease of saliva production and in a very severe condition called xerostomia.1 Following irradiation-induced, irreversible hyposalivation often occur because the stem cell sterilization of the primitive salivary glands.2 It is required to apply an alternative approach to treat the severe damage of glands and the left tissue. One of the alternative approaches for this purpose is the stem cell therapy.

Some factors that affect the success of stem cell therapy include the following: stem cells strongly attach and survive in the defect area and they can integrate with the surrounding microenvironment.3 However, considering the low amount of survived cells, which shows the low viability of the transplanted stem cells, the effectiveness of stem cell therapy decreases. The low rate of viability is probably due to the conventional method of culture, which was done under normoxic condition with 21% O₂. It is contrary to the in vitro environment, which is a hypoxic condition with 1 to 7% O₂ depending on the stem cell location and type. Therefore, it is assumed that the same condition as the micro environment is necessary, to get a desirable result regarding the viability.
of the transplanted stem cells in the injured tissue and is expected to allow the stem cells proliferation and differentiation into the origin-like cells. In addition to the appropriate microenvironment, there are others factors that also play a role in the success of stem cell therapy, the factors that can induce stem cells to migrate into the defect area. One of the mediators that plays an important role in the migration process into the defect area is stromal derived-cell factor 1 (SDF1) through binding with CXCR4 receptor.

The purpose of this study was to explain the mechanism of regeneration of salivary gland defect due to ionizing radiations by BM-MSCs transplantation that has been given hypoxic preconditioning with 1% of O₂ concentration.

Materials and Methods

Ethical Approval
All animal studies were performed via a protocol approved by the Institutional Animal Care and Use Committee of Faculty of Veterinary Medicine, Universitas Airlangga, and complied with the National Research Council’s guidelines (366-KE) through ethical seminar.

Salivary Gland Damage in Animal Model
Damage to salivary glands in a healthy male Wistar rat was induced by single dose of 15 Gy radiation in the ventral region of the neck. Rats used in this study were 3 to 4 months old and each with 250 to 300 g weight. Rats were kept in an individual plastic cage in the laboratory for Experimental Animal of Institute of Tropical Disease, Universitas Airlangga with adequate ventilation.

Treatment
This research was a true experimental posttest control group design. The bone marrow mesenchymal stem cells (BM-MSCs) were isolated from the femur of Wistar male rats. Stem cell culture was divided into two conditions: normoxic (O₂: 21%) and hypoxic conditions for 48 hours of incubation in a low oxygen tension chamber consisting of 95% N₂, 5% CO₂, and 1% O₂. Forty male Wistar rats were transversely incised from paraffin blocks. Rats were divided into groups: control (T0–), negative control (T0+), positive control (T1+), and treatment (T1–). The mean of SDF-1 and CXCR4 expression significant differences of that of the normal tissue.

Statistical Analysis
Data were analyzed statistically with normality test and MANOVA test using Statistical Package for the Social Sciences (SPSS) 17.0 (IBMTM, Chicago, Illinois, United States).

Immunohistochemical Methods for Observation of SDF1-CXCR4 and Bcl-2
Immunohistochemistry (IHC) was performed to determine the expression of SDF1-CXCR4 and Bcl 2. First, salivary glands were transversely incised from paraffin blocks. Monoclonal antibodies, namely anti-SDF1-CXCR4 and anti-Bcl-2 were used in this technique. Samples were observed under a light microscope at a magnification of 200× to check SDF1-CXCR4 and Bcl-2 expression. The expression of each variable was described by the number of cells with brown discoloration due to diaminobenzidine (DAB-chromogen) in each incision.

Histological Observation of Salivary Glands
Light microscopy examination was performed to study salivary gland histology and regenerate acinar cells. After that, samples were prepared for histology in the following steps: submandibular gland of rats was fixed in 10% formalin buffer, followed by dehydration using a series of alcohol, from 70, 80, 90, to 96% (absolute). Gland tissues of rats were cleared in a xylene solution. The tissues were infiltrated with an embedding agent, the liquid paraffin. Micrómetro was set with a distance at 4 to 6 µ for sectioning, and the sections were placed on a slide. The embedding process must be reversed to get the paraffin wax out of the tissue and allow water soluble dyes to penetrate the sections. Therefore, the slides are “deparaffinized” by washing them through xylenes to alcohols to water before any staining can be done. The routine H&E staining was performed. After that, Canada balsam was used to mount the stained section and covered with a coverslip. Submandibular gland and acinar cell regenerations are observed and identified based on the histological measures of that of the normal tissue.

Results
The IHC examination results showed that in the hypoxic group the expression of binding SDF1-CXCR4 and Bcl 2, and α-amylase enzyme increased significantly than in the normoxia group. The mean of SDF-1 and CXCR4 expression significantly expressed in hypoxic preconditioning group than in the normoxic group. A significant difference (p < 0.05) between normoxia and hypoxia group with SDF1-CXCR4 binding also showed in MANOVA test (Table 1).
mainly CXCR4 expression (that was stronger than the expression in the normoxia group (►Fig. 1). It also can be seen from the green color of the microscopic image of salivary gland tissue that occupies most of the ductal basal membrane. The distribution of BM-MSCs, as depicted by labeled PKH 26, shows that green coloration is stronger in hypoxic groups than in the normoxic group (►Fig. 2). The mean of Bcl-2 expression was significantly expressed in hypoxic preconditioning group than in the normoxic group; the MANOVA test results indicated a significant difference (p < 0.05; ►Table 2). It can be seen from a comparison of the expression of Bcl-2 (brown chromogen) between treatments that groups with acute hypoxia showed stronger Bcl-2 expression compared with the acute normoxia group (►Fig. 3). Furthermore, there were significant differences in MANOVA test (p < 0.05) of α-amylase expression between the treatment groups. The acute hypoxia group showed a significant increase in α-amylase enzyme activity compared with that in the acute normoxia group (►Table 3).

**Discussion**

Stem cell transplantation is one of the promising therapies that is predicted to restore the function of the salivary gland damage by regenerating acinar cells. Stem cell therapy can be successful in addition to the need for adaptive stem cells and when transplanted stem cells are attached and integrated

**Table 1** Average value and standard deviation (SD) expression of SDF1-CXCR4

<table>
<thead>
<tr>
<th>Groups</th>
<th>SDF1-CXCR4</th>
<th>p-Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Negative control group (T0–)</td>
<td>1.640</td>
<td>0.555</td>
</tr>
<tr>
<td>Positive control group (T0+)</td>
<td>1.160</td>
<td>0.219</td>
</tr>
<tr>
<td>Treatment group 1 (T1)</td>
<td>2.920</td>
<td>0.540</td>
</tr>
<tr>
<td>Treatment group 2 (T2)</td>
<td>3.720</td>
<td>1.825</td>
</tr>
</tbody>
</table>

Abbreviation: SD, standard deviation.

**Table 2** Average values and standard deviations of expression of Bcl-2

<table>
<thead>
<tr>
<th>Groups</th>
<th>Bcl-2</th>
<th>p-Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Negative control group (T0–)</td>
<td>23.600</td>
<td>5.176</td>
</tr>
<tr>
<td>Positive control group (T0+)</td>
<td>4.000</td>
<td>1.870</td>
</tr>
<tr>
<td>Treatment group 1 (T1)</td>
<td>5.400</td>
<td>1.140</td>
</tr>
<tr>
<td>Treatment group 2 (T2)</td>
<td>11.800</td>
<td>1.303</td>
</tr>
</tbody>
</table>

Abbreviation: SD, standard deviation.

Note: p < 0.05 is significant.
The results of this study indicate that BM-MSCs cells that have been given hypoxic preconditioning have better therapeutic ability than those under normoxic conditions so that they can induce cell repair processes. It can be seen from the green color of the microscopic image of salivary gland tissue that occupies most of the ductal basal membrane. This shows the migration of BM-MSCs in the basal membrane of the ducts and acinar cells which are heavily damaged by radiation exposure. The distribution of PKH 26-labeled BM-MSCs shows that green coloration is stronger in hypoxic group than in the normoxic group. The IHC examination results in the hypoxic group showed that the expression of binding SDF1-CXCR4 and Bcl-2 increased significantly than in the normoxic group.

A recent study states that the activation of SDF-1–CXCR4 bonds in tissues plays a role in the transduction of various signals that can regulate several biological functions, such as cell survival, proliferation, chemotaxis, and cell differentiation, which is in line with result of this study. One of the main functions of SDF1–CXCR4 bond is regulating the trafficking of BM-MSCs cells in during homing in the injured area. Moreover, the MANOVA test results showed that there were significant differences in the expression of α-amylase between the treatment groups. The hypoxia group showed a significant increase in the activity of α-amylase enzyme compared with that in the acute normoxic group after transplantation. The results of this study indicate the regeneration of the damaged salivary glands.

All these have shown the influential effect of low O2 concentration on MSCs biology and raised serious concern over its therapeutic efficiency and biosafety. Environmental stress to the in vitro cultured MSCs can be caused by higher O2 concentration. Moreover, in recent years after many transplantation studies, early senescence, longer population doubling time, DNA damage, and poor engraftment have presented clear evidence regarding the negative influence of ambient O2 concentration on MSCs. A large number of number of studies suggest that hypoxia activates several transcription factors in the nucleus, such as HIF-1α and NFκβ, and these factors interact with paracrine factors such as MEK and PI3K/Akt. All these interactions increase the secretion of several growth factors, such as stromal-derived factor 1 (SDF-1), hepatocyte growth factor (HGF), and vascular endothelial growth factor.

### Table 3  Average values and standard deviations of α-amylase enzyme expression

<table>
<thead>
<tr>
<th>Groups</th>
<th>α-amylase</th>
<th>p-Value</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Mean</td>
<td>SD</td>
</tr>
<tr>
<td>Negative control group (T0–)</td>
<td>289,259.000</td>
<td>18,645.313</td>
</tr>
<tr>
<td>Positive control group (T0+)</td>
<td>95,330.400</td>
<td>31,503.973</td>
</tr>
<tr>
<td>Treatment group 1 (T1)</td>
<td>152,088.200</td>
<td>6,434.510</td>
</tr>
<tr>
<td>Treatment group 2 (T2)</td>
<td>186,118.400</td>
<td>5,971.156</td>
</tr>
</tbody>
</table>

### Abbreviation: SD, standard deviation.
Note: p < 0.05 is significant.
(VEGF) with increased expression of each receptor such as CXCR4, increased secretion of some anti apoptotic proteins, such as Bcl-2 and Bcl-xL as a survival factor.16

**Conclusion**

This study concluded that (1) BM-MSCs transplantation with hypoxic preconditioning of 1% O₂ increases the expression of SDF1-CXCR4 and Bcl-2 that contribute to cell migration and cell survival. (2) BM-MSCs can improve regeneration process of the damaged salivary glands by increasing the activity of the α-amylase as a marker of regeneration process in the salivary gland tissue.

**Conflict of Interest**

None declared.

**Acknowledgments**

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