

Small-Molecule Amyloid Beta-Aggregation Inhibitors in Alzheimer's Disease Drug Development

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Abstract

Alzheimer's disease (AD) is still an incurable neurodegenerative disease that causes dementia. AD changes the brain function that, over time, impairs memory and diminishes judgment and reasoning ability. Pathophysiology of AD is complex. Till now the cause of AD remains unknown, but risk factors include family history and genetic predisposition. The drugs previously approved for AD treatment do not modify the disease process and only provide symptomatic improvement. Over the past few decades, research has led to significant progress in the understanding of the disease, leading to several novel strategies that may modify the disease process. One of the major developments in this direction is the amyloid β ($A\beta$) aggregation. Small molecules could block the initial stages of $A\beta$ aggregation, which could be the starting point for the design and development of new AD drugs in the near future. In this review we summarize the most promising small-molecule $A\beta$ -aggregation inhibitors including natural compounds, novel small molecules, and also those are in clinical trials. Moreover, we briefly summarized some reported docking studies of small-molecule $A\beta$ aggregation inhibitors. These will give us an idea about the chemical features required to design novel small molecules with anti- $A\beta$ aggregation properties.

Keywords

- ▶ Alzheimer's disease
- ▶ small molecule amyloid β -aggregation inhibitors
- ▶ drug design

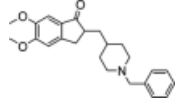
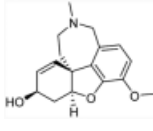

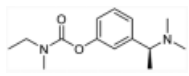
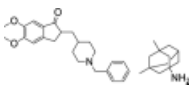
Introduction

Alzheimer's disease (AD) is the most common form of dementia and is still an incurable, progressive neurodegenerative disorder.¹ AD is a multifactorial disease in which a complex of proteins, enzymes, or receptors is involved. The pathogenesis of AD is not completely clear. However, the typical pathological hallmarks are amyloid β ($A\beta$) deposits, tau (τ) protein aggregation, oxidative stress, and decreased levels of acetylcholine (ACh) in the brain.² The etiological mechanisms underlying the neuropathological changes in AD remain unclear but are probably affected by both environmental and genetic factors.³ AD is characterized by relatively slow, chronic but progressive neurodegeneration and impairment in cognition accompanied by abnormal behavior and personality changes, ultimately leading to full dementia. Incidence increases with age, affect-

ing an estimated 35 million patients worldwide.⁴ However, since dementia primarily affects those aged over 60, increased longevity has led to increased rates of AD.⁵

Currently there are no effective treatments or interventions to mitigate AD progression, and the incidence rates for AD doubled every 5 years from age 65. The global burden of AD patients is therefore expected to be 106.8 million by 2050.⁶ Current treatment of the disease, essentially symptomatic, is based on three cholinesterase inhibitors (donepezil, galantamine, and rivastigmine) and memantine, affecting the glutamatergic system (–Table 1). Unfortunately, none of these drugs stop the progressive loss of neurons and there is no treatment that can halt the progressive deterioration of cognitive faculties in AD patients.⁷ Despite enormous information gained, the prevailing hypotheses regarding AD pathogenesis have failed to deliver strategies for mitigation of symptoms.⁸

Table 1 Medication for Alzheimer's disease

Drug name	Brand name	Structure	FDA approved in
Donepezil	Aricept		1996
Galantamine	Razadyne		2001
Memantine	Namenda		2003
Rivastigmine	Exelon		2000
Donepezil + Memantine	Namzaric		2014

Abbreviation: FDA, U.S. Food and Drug Administration.

Amyloid Cascade Hypothesis of Alzheimer's Disease

The amyloid hypothesis proposes A β as the main cause of the disease. It suggests that misfolding of the extracellular A β protein accumulated in senile plaques (SPs) and the intracellular deposition of misfolded tau protein in neurofibrillary tangles (NFTs) cause memory loss and confusion (also cause personality and cognitive decline over time). Accumulated A β peptide is the main component of SPs and is derived from the proteolytic cleavage of a larger glycoprotein named amyloid precursor protein (APP). APP is a transmembrane protein that plays an important role in a range of biological activities, including neuronal development, signaling, intracellular transport, and other aspects of neuronal homeostasis. APP is the precursor molecule cut by β - and γ -secretases to produce a 37 to 49 amino acid residue peptide. A β lies at the heart of the amyloid cascade hypothesis and its amyloid fibrillar form is the primary component of amyloid plaques found in the brains of AD patients.^{9–15} A β peptides are mainly observed in the region of the hippocampus and the neocortex as well as in the cerebrovasculature.^{9–13} Human

APP can be processed via two alternative pathways: amyloidogenic and nonamyloidogenic. APP is first cleaved by α -secretase (nonamyloidogenic pathway) or β -secretase (amyloidogenic pathway), generating membrane-tethered α - or β -C terminal fragments.⁹

Since the first description of presenile dementia by Alois Alzheimer in 1907, SPs and NFTs are considered as the key pathological hallmarks of AD. The identification of A β in SPs and genetic studies that identified mutations in the APP, presenilin 1 (PSEN1), and presenilin 2 (PSEN2) genes leading to the accumulation of A β and early-onset familial dementia resulted in the formulation of the “amyloid cascade hypothesis.”^{15,16}

The enzymatic processes responsible for the metabolism of APP to A β are now reasonably well understood. APP is sequentially cleaved by β - and γ -secretases. β -Secretase first cleaves APP to release a large secreted derivative, sAPP β . A fragment of 99 amino acids remains membrane bound, and is rapidly cleaved by γ -secretase to generate A β . Hence, numerous different A β species exist, but those ending at position 40 (A β 40) are the most abundant ones (~80–90%), followed by A β 42 (~5–10%). The longer forms of A β , particularly A β 42, are more hydrophobic, fibrillogenic, and are the principal species deposited in the brain.⁹

The primary amino acid sequence of A β (**Fig. 1**) was first discovered from extracellular deposits and amyloid plaques in 1984. A β monomers aggregate into various types of assemblies, including oligomers, protofibrils, and amyloid fibrils (**Fig. 2**). Amyloid fibrils are larger and insoluble, and they can further assemble into amyloid plaques. While amyloid oligomers are soluble and may spread throughout the brain. Amyloid plaques with A β as the main component are most commonly found in the neocortex in the brain of AD patients.⁹ Amyloid cascade hypothesis (**Fig. 3**) proposes that the deposition of A β is the initial pathological event in AD leading to the formation of SPs and then to NFTs, neuronal cell death, and ultimately dementia. While there is substantial evidence supporting the hypothesis, there are also limitations, such as SPs and NFTs may develop independently, and SPs and NFTs may be the products rather than the causes of neurodegeneration in AD.¹⁵ On the other hand, tau (τ) proteins are also found in several less common neurodegenerative diseases, notably in the absence of neurotic plaques. The NFTs in the different diseases have some distinctive morphological features and may exhibit a distinct composition of tau isoforms that differ from AD.^{17,18}

In contrast, the amount of oligomeric A β is increased in AD brain extracts. A β oligomers trigger synapse failure and memory impairment, resulting in impaired brain function in the final stages of the disease. Further studies reported that cognitive deficits appeared before plaque deposition or the detection of insoluble amyloid fibrils. These all evidence proved that A β oligomers trigger neuronal death rather than insoluble fibrils or plaques.⁹

Current Trend of AD Research

AD has been studied over a century, but acetylcholinesterase inhibitors and memantine are the only drugs currently approved for its management (**Table 1**). These drugs

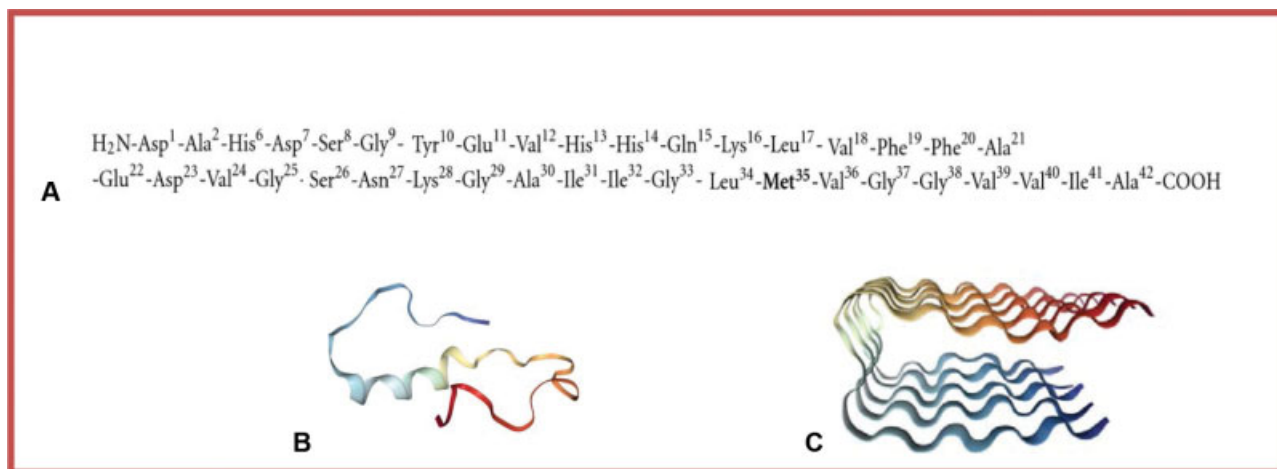


Fig. 1 Structure of Aβ monomer and fibrils. (A) The primary amino acid sequence of the 42-amino acid Aβ, (B) 3D structure of Aβ 1–40 monomer (PDB code: 2LFM), and (C) Aβ 1–42 fibrils, homopentamer structure (PDB code: 2BEG).

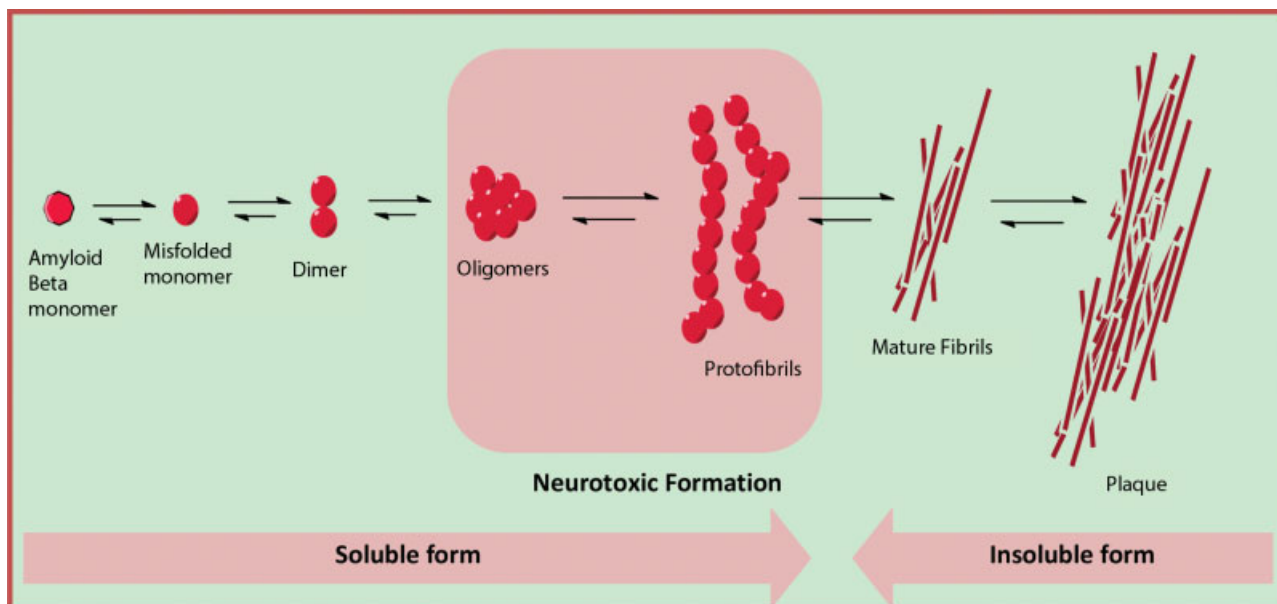


Fig. 2 Conversion of amyloid β (Aβ) monomers to oligomers, protofibrils and fibrils.

provide symptomatic improvement alone but do less to modify the disease process. The extensive insight into the molecular and cellular pathomechanism in AD over the past few decades has provided us significant progress in the understanding of the disease. Several novel strategies that seek to modify the disease process have been developed.¹

Current directions in the search for novel, potentially effective agents for the treatment of AD include agents acting upon Aβ (early-stage intervention), such as vaccines, antibodies, and small-molecule inhibitors or modulators of β- and γ-secretases; agents directed against the tau protein as well as compounds acting as antagonists of neurotransmitter (mild-stage intervention), and agents targeting microglia or some anti-inflammation (late-stage intervention) (→ Fig. 4). The major developments in the amyloid and tau-based therapeutics could hold the key to treatment of AD in the near future.¹ Development of new effective drugs acting

upon the central nervous system is usually a difficult and time-consuming process, and in the case of AD to date clinical trials have had a very high failure rate. Most of phase II clinical trials end with a positive outcome, but do not succeed in phase III, often due to serious adverse effects or lack of therapeutic efficacy. Now it becomes one of the greatest challenges in modern medicine to develop novel drugs with strong disease-modifying properties.^{19,20}

Several large clinical trials are actively studying individuals to discover potential therapies by 2025.²¹ Preclinical studies performed in academic as well as industrial settings focus on many potential molecular targets involved in the pathogenesis of AD. Because of a huge attrition rate, only selected candidates are accepted into clinical trials as potential anti-AD agents. Due to the importance of these studies, several comprehensive reviews on anti-AD drug development prospects have been published in recent years.^{1,22–27} In

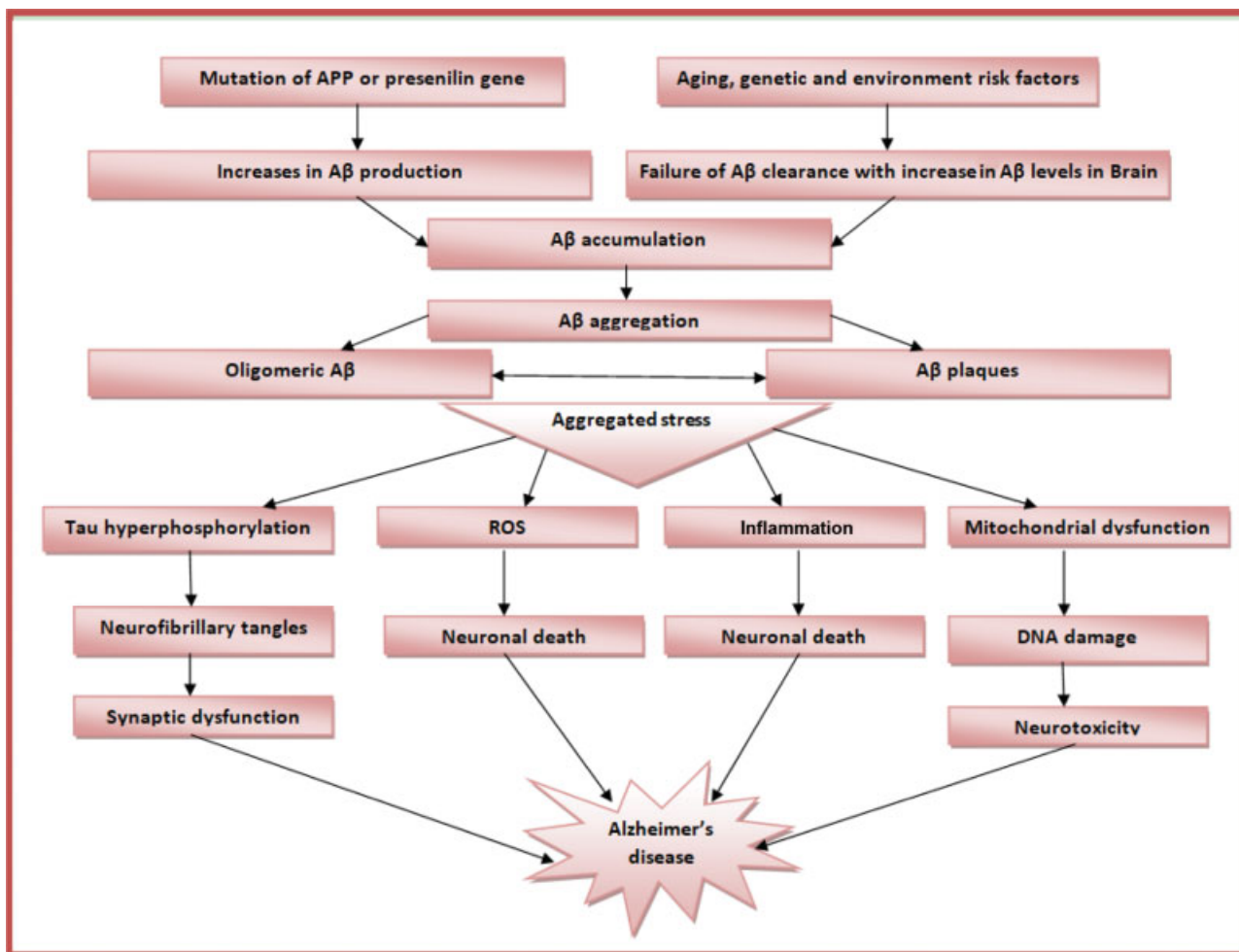


Fig. 3 Proposed amyloid cascade hypothesis.

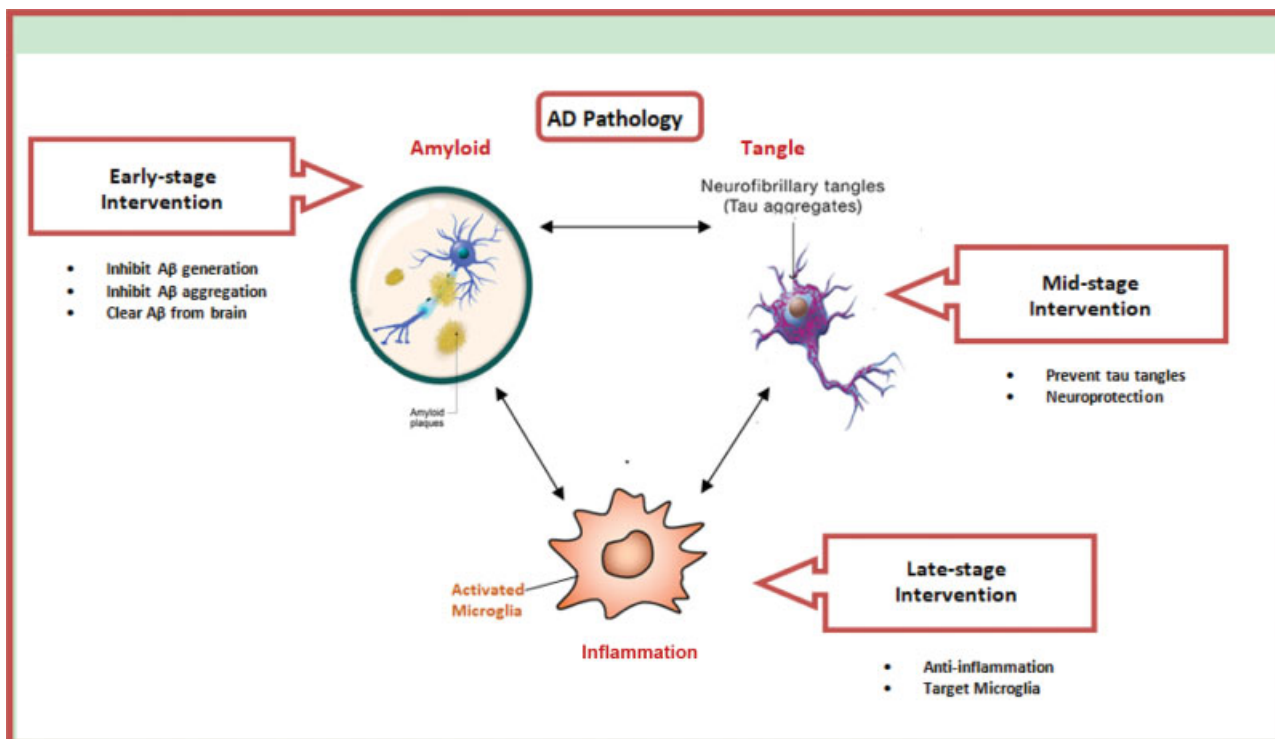


Fig. 4 Current trend of Alzheimer's disease research.

this review we present the latest advances in the scope of the most promising small-molecule A β -aggregation inhibitors that are currently undergoing AD drug development and clinical trials.

Small-Molecule Amyloid β -Aggregation Inhibitors

The progressive production and subsequent accumulation of A β play a central role in AD. A β is released in a soluble form that may be responsible for cognitive dysfunction in the early stages of the disease, then progressively forms oligomeric, multimeric, and fibrillar aggregates, triggering neurodegeneration. Eventually, the aggregation and accumulation of A β culminates with the formation of extracellular plaques, one of the morphological hallmarks of AD. Based on the conformation/oligomerization hypothesis, molecules able to stabilize the soluble A β conformation destabilize the altered amyloidogenic conformer or prevent the required conformational transition, which could be effective inhibitors of amyloid plaque formation. These could be very potent drug candidates for AD treatment.²⁸ Indeed, recent and ongoing clinical trials in AD continue to experiment with very early drug interventions in attempts to develop a disease-modifying agent but unfortunately, so far, no such approved agents are currently available.²⁹ Some small molecules are able to inhibit the formation and extension of A β fibrils, and also destabilize A β fibrils in vitro. These include natural compounds or commercially available bioactive compounds, drugs, surfactants, Cu/Zn chelators, phenothiazines, and sulfonated dyes such as Congo red (CR) and thioflavin T (ThT).^{30–32} As the strategy of inhibiting A β aggregation has increasingly gained acceptance, greater numbers of inhibitors have been developed and the structure–activity relationships of potent inhibitors have been systematically explored.^{28,33,34} CR was the first small molecule reported to bind to amyloid in tissue sections.³⁵ Later ThT and thioflavin S were also shown to stain amyloid deposits. CR and ThT, the two dyes (\rightarrow Fig. 5), are the classical reagents to detect characteristic A β sheet-mediated fibrillization. CR and ThT have also been shown to inhibit A β fibril formation at higher concentrations.^{36,37}

Anti-A β Natural Compounds

Several plant-derived natural compounds are known to exhibit anti-amyloid aggregation activity which makes them attractive as potential therapies to treat AD. These natural compounds are known to exhibit direct binding to various amyloid species

including oligomers and fibrils, which in turn can lead to conformational change in the β -sheet assembly to form non-toxic aggregates.⁴⁰ Several polyphenols (\rightarrow Fig. 6), including curcumin, resveratrol, and epigallocatechin-3-gallate (EGCG), have progressed to clinical trials for AD treatment. Moreover, based on their multiple functions, including antioxidant, anti-inflammatory, and metal chelating capacities, polyphenols are a rich source for a variety of different structural backbones that can be utilized in rational drug design efforts to find multi-functional anti-amyloid agents.^{38–41} Several recent reviews summarized the progress in natural product-based amyloid inhibitors and also analyzed their mechanisms of inhibition in vitro.^{38,39}

Small Molecules in Clinical Trials

Compounds such as small molecules that target specific A β subregions represent the first generation of amyloid-based therapeutics with the potential to demonstrate disease-modifying activity. Although the results of ongoing clinical trials are inconclusive, these compounds hold the promise of a new day in the development of disease-modifying therapies for AD.⁴²

Phase III clinical trials are currently underway for agent **ALZT-OP1** developed by AZ Therapeutics⁴³ for the prevention and treatment of early AD. It is a combination drug therapy consisting of the administration of two previously approved drugs, which have been shown to inhibit A β aggregation and neuroinflammation. **GV-971** (Shanghai Green Valley) is a sodium oligo-mannurate developed for the oral treatment of mild-to-moderate AD and its phase III clinical trials also finished on October 25, 2018 as a safe, well-tolerated oral drug. It showed the ability to reduce the toxicity of A β peptide in vitro.^{44,45} **KHK6640** (Kyowa Hakko Kirin Pharma), also an A β aggregation inhibitor, is in phase III clinical trial.⁴⁶

Phase II clinical trials include several low-molecular-weight compounds that could be attributed to the general group of “anti-amyloidogenic” drugs.⁴⁷ **Scyllo-Inositol (ELND005)**⁴⁸ is a small-molecule inhibitor of A β aggregation in phase II/III clinical development at Transition Therapeutics for the oral treatment of AD. **Posiphen**,⁴⁹ discovered by the U.S. National Institute on Aging, is a small, orally active anti-amyloidogenic agent which is in phase II clinical trials for the treatment of AD and Parkinson's disease. **Bexarotene** (Cleveland Clinic) is a U.S. Food and Drug Administration approved anticancer agent but is not approved for the use of AD. It reduced A β in the brain in experimental models of AD, currently in phase II clinical study.

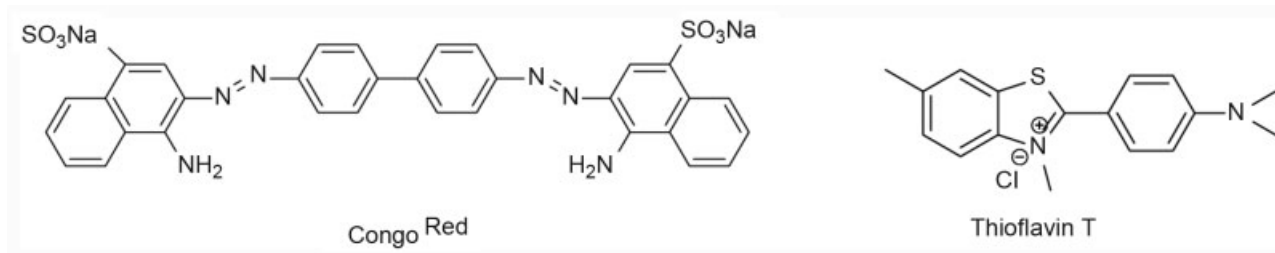


Fig. 5 Chemical structures of Congo red and thioflavin T.

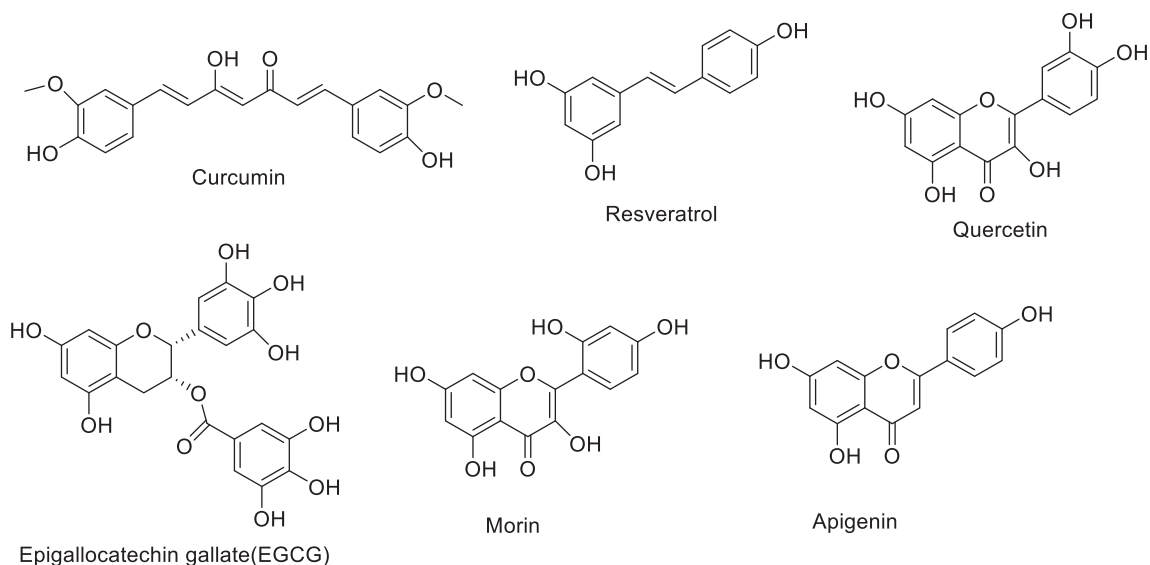


Fig. 6 Chemical structures of natural products identified as inhibitors of amyloid β aggregation.

AZD-3293 (AstraZeneca) is in phase I clinical study. Successfully it finished single and multiple ascending dose studies on Japanese volunteers. **PPI1019** (PRAECIS Pharmaceuticals Inc.) is in phase I trial, completed single-dose escalation safety study of PPI-1019 in patients with mild–moderate AD. **AD-04** (German Center for Neurodegenerative Diseases) is in phase I clinical trial (www.clinicaltrials.gov). **ALZ-801** (Alzheon)⁵⁰ and **Ro-63-8695** (GlaxoSmithKline)⁵¹ target amyloid aggregation and are currently in phase I trials. Phase I clinical trials are initiated for **SAN-61** (Diamedica company) and **Exebryl-1** (Proteo Tech) also as a

potential dual-action small molecule. Both simultaneously work as an inhibitor on amyloid plaques and neurofibrillary aggregates.^{52,53} Available structures of some small-molecule $A\beta$ -aggregation inhibitors are summarized in **► Fig. 7**.^{42,47}

Molecular Docking Studies of Small-Molecule Amyloid β -aggregation Inhibitors

Studies suggest that monomeric $A\beta$ is nontoxic and prevents neuronal cell death caused by oxidative stress.⁵⁴ The toxic

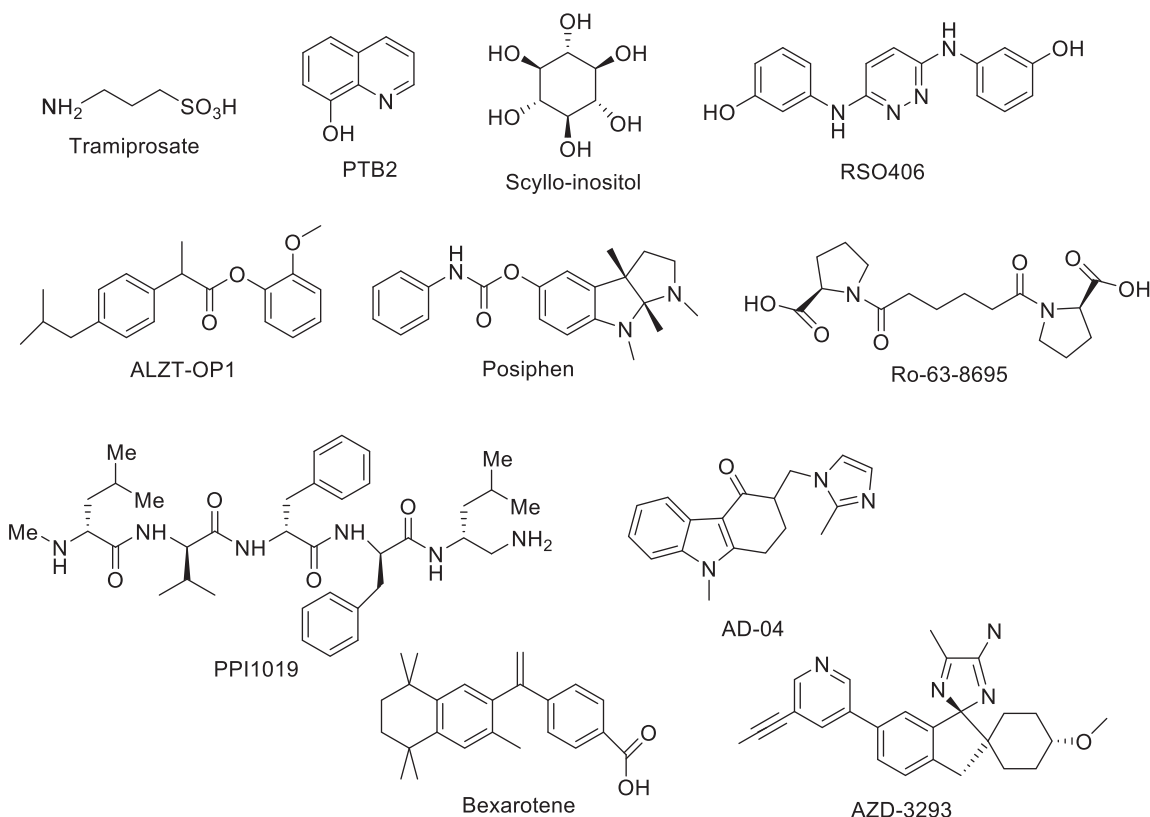


Fig. 7 Available structures of some small-molecule amyloid β -aggregation inhibitors.

effects of aggregation-prone A β result from conformational transition of A β monomers from predominantly α -helical to β -sheet structures that result in monomeric A β aggregation and fibrillization. Therefore, preventing the conformational transition of the A β monomer from an initial random coil or α -helix into a β -sheet is the primary goal of blocking A β toxicity by small-molecule inhibitors.³⁸

The amino acid sequence of human A β 42 is: NH₂-¹Asp-²Ala-³Glu-⁴Phe-⁵Arg-⁶His-⁷Asp-⁸Ser-⁹Gly-¹⁰Tyr-¹¹Glu-¹²Val-¹³His-¹⁴His-¹⁵Gln-¹⁶Lys-¹⁷Leu-¹⁸Val-¹⁹Phe-²⁰Phe-²¹Ala-²²Glu-²³Asp-²⁴Val-²⁵Gly-²⁶Ser-²⁷Asn-²⁸Lys-²⁹Gly-³⁰Ala-³¹Ile-³²Ile-³³Gly-³⁴Leu-³⁵Met-³⁶Val-³⁷Gly-³⁸Gly-³⁹Val-⁴⁰Val-⁴¹Ile-⁴²Ala-COOH. Residues ¹Asp-²⁸Lys and ²⁹Gly-⁴²Ala respectively represent a hydrophilic domain and a hydrophobic domain. Unfolded and soluble monomeric A β primarily have a random-coil structure when they are released into extracellular matrix.⁵⁵ Conformational transition of A β from an unfolded monomeric native state to a β -sheet happens rapidly and initiates the aggregation process to form toxic A β oligomers.⁵⁶ A β 42 oligomers are the most toxic form and play important causal roles in AD.⁹ Numerous studies indicate that some of the amino acid residues are responsible for the aggregation, these are (1) the fibril-forming A β fragment: amino acid residues ¹²Val-²⁴Val and ³⁰Ala-⁴⁰Val, responsible for their native states, self-assembling into a β -sheet structure;⁵⁷ (2) amino acid residues ²⁴Val-²⁸Lys in a β -sheet conformation promote A β oligomerization;⁵⁸ (3) the nucleation site of aggregation: hexapeptide sequence ¹⁶KLVFFA¹⁹ (¹⁶Lys-²¹Ala) acting as a steric zipper, which leads to dimer formation and eventually larger aggregates;^{38,59-61} (4) aromatic residues in the A β monomer, phenylalanine (Phe⁴, Phe¹⁸ and Phe¹), and tryptophan (Tyr¹⁰) play a significant role in the A β self-assembly process by enhanced fibril assembly kinetics;^{62,63} (5) the three specific sites, Arg-to-Gly, Tyr-to-Phe, and His-to-Arg, are believed to be important in A β aggregation or A β -induced neurotoxicity;^{64,65} and (6) the amino acid residues His¹³ and Met³³ also promote toxic conformations in A β oligomers.⁶⁶⁻⁶⁸

A wide range of natural and synthetic molecules (both large and small molecules) has been investigated for their ability to counteract A β aggregation and toxicity.^{38,69} In this review, we will summarize the molecular modeling studies of some important natural and synthetic inhibitors of A β aggregation.

Curcumin

The antiaggregating activity of curcumin (**Fig. 6**) against A β has been extensively investigated including its structural features that contribute to the antioligomerization activity. Curcumin can interact with A β oligomers and A β fibrils. Modeling studies have shown that curcumin interacts with the ¹²V and ¹⁶KLVFFA¹⁹ residues of the A β .⁷⁰⁻⁷² Also, it interacts with ³⁷Gly in A β 42 in the C-terminus of the A β fibrils⁷³ and the aromatic residues (Tyr, Phe, and His) in the A β dimer.⁷⁴ Another computational investigation suggested that amino acids ¹⁶K, ¹⁷L, ¹⁸V, and ²⁰F in ¹⁶KLVFFA¹⁹ residues were involved in stabilizing the curcumin-A β octamer assembly. These investigations show that curcumin binding to A β -aggregates leads to significant conformational change and shifts the equilibrium toward

the formation of nontoxic A β aggregates including dimers, oligomers, protofibrils, and fibrils in the A β -aggregation pathways which prevent neurotoxicity associated with various forms of A β -aggregates.⁷⁵

Resveratrol

Several studies indicate that resveratrol (**Fig. 6**) directly interacts with A β monomers and fibrils. It inhibits A β fibrillization and converts toxic oligomers into nontoxic species. Interaction between resveratrol and aromatic side residues (⁴Phe, ¹⁰Tyr, ¹⁹Phe, and ²⁰Phe) of A β selectively remodeled A β conformers that possess β -sheet structures into nontoxic species. Researchers found that its polyphenol aglycones and glycosides were responsible for remodeling toxic A β oligomers into nontoxic species.⁷⁶⁻⁷⁹

Epigallocatechin Gallate

EGCG (**Fig. 6**) inhibits conformational change from a random-coil to a β -sheet structure and A β oligomers formed in the presence of EGCG.⁸⁰ There are 12 important residues (⁴Phe, ⁵Arg, ¹⁹Phe, ²⁰Phe, ²²Glu, ²⁸Lys, ²⁹Gly, ³⁴Leu to ³⁷Gly, and ⁴¹Ile of A β) that strongly interact with EGCG. Simultaneously, the side chains of some hydrophobic residues (Phe, Met, and Ile) and the main chains of ²⁸Lys and ²⁹Gly provide nonpolar interactions with A β . These causes remodeling of mature A β fibrils and toxic oligomers into smaller nontoxic aggregates with the loss of β -sheet content.^{38,81} Modeling studies reveal that EGCG displays an antiaggregation effect from mainly two pathways: first, EGCG binds to the native form of A β through interactions with the side chains of specific residues, and second, EGCG binds to the misfolded A β species with noncovalent interactions involving the A β backbone and subsequently remodels toxic aggregates into small nontoxic, off-pathway oligomers.³⁸

Tramiprosate

3-Aminopropanesulfonic acid (**Fig. 7**) targets the ¹³HHQK¹⁶ subregion (¹³His-¹⁶Lys) at the N-terminus of A β . ¹³His-¹⁶Lys amino acids are important for oligomerization, fibril propagation, and neurotoxicity.⁴² Due to its structural simplicity, tramiprosate is highly specific to A β . But unfortunately, it failed in the late stages of a phase III clinical trial.²⁸ Despite its clinical failure, the data obtained from the studies will assist us in the design and development of novel small molecules with anti-amyloid aggregation properties.⁸²

Scyllo-Inositol

Scyllo-Inositol (**Fig. 7**), a potential therapeutic compound for AD, has been shown to inhibit A β (1-42) fibrillogenesis in vitro. It stabilizes cell-derived small molecular weight oligomers and reduces amyloid plaque load. Also, it has shown promise in current phase II/III clinical trials.^{48,83,84} Modeling studies revealed that scyllo-Inositol interacts with the C-terminus of A β .⁴²

RS-0406

N,N'-Bis(3-hydroxyphenyl)pyridazine-3,6-diamine (**Fig. 7**) is a small molecule which inhibits A β 1-42 fibrillogenesis and

a novel β -sheet breaker.⁸⁵ RS-0406 interacts with the ¹⁶KLVFFA¹⁹ residues and inhibits A β fibrillation.^{85,86}

New Small Molecules

The last few years have seen a surge in the discovery of small molecules as disease-modifying therapies for AD.⁸⁷ Here, we will highlight some recent developments in the design of the small-molecule A β -aggregation inhibitors as potential disease-modifying agents and briefly summarized some previously reported docking studies of these recently developed small molecules.

In a recent study, Sancho and coworkers identified compounds **I–III** (**► Fig. 8**) as A β aggregation inhibitors based on a high-throughput study by screening a chemically diverse compound library.⁸⁸ Molecular modeling study of compound **I** (2-methyl-5,6,7,8-tetrahydro-4*H*-[1*b*]benzothieno[2,3-*d*] [1,3]oxazin-4-one) with an A β dimer assembly showed that the tetrahydrobenzenethieno ring was in van der Waal's contact with ¹⁶KLVFFA¹⁹ residues, whereas benzenethieno and oxazinone rings were in contact with side chains of isoleucine and leucine at the C-terminal via nonpolar contact. Modeling compound **II** (2,5-dichloro-*N*-(4-piperidinophenyl)-3-thiophenesulfonamide) in the A β -dimer showed that the phenylpiperidine substituent was oriented toward the *N*-terminal in the LVFF region. The phenyl ring underwent T-shaped π - π interaction with ¹⁶KLVFFA¹⁹ residues. The 2,5-dichlorothiophenesulfonamide was oriented toward the C-terminal, where it was in van der Waal's contact with amino acid side chains of isoleucine, glycine, and leucine. In contrast, the modeling study of compound **III** (*N*-(4-chloro-2-nitrophenyl)-*N'*-phenylurea) in the A β -dimer model showed that

this compound only interacted at the *N*-terminal region. The two phenyl rings linked to the urea moiety were in close proximity to the ¹⁶KLVFFA¹⁹ region. Compound **IV** (6-(4-chlorophenyl)sulfonyl)-2-phenylpyrazolo[1,5-*a*]pyrimidin-7-amine) exhibits a V-shaped conformation in the A β dimer assembly. The bicyclic pyrazolopyrimidine underwent π -alkyl interactions with amino acid side chains of isoleucine and leucine at the C-terminal, whereas the chlorophenyl ring interacts with the phenylalanine ring at the *N*-terminal. Compared with compounds **I–IV**, compound **IV** is a larger molecule, longer along its axis, which helps in making additional contacts in the steric-zipper assembly and provides better interaction.^{87,88}

Luo and coworkers reported the design of novel tacrine-alkoxybenzene hybrids, compound **V** (**► Fig. 8**), as dual inhibitors of cholinesterases and A β aggregation. They reported that these compounds were able to prevent self-induced A β -aggregation.⁸⁹ The tetrahydroacridine ring was oriented toward the LVFF region, where it underwent π -alkyl interactions with leucine and valine side chains of ¹⁶KLVFFA¹⁹ residues and the NH of tetrahydroacridine ring formed a hydrogen bond with the backbone C = O of valine.^{87,89}

Kanai and coworkers reported compound **VI** (4-benzyl-*N*-isoneopentyl-6-phenoxycolinamide, **► Fig. 8**), which exhibited dose-dependent inhibition of A β aggregation.⁹⁰

Modeling study of compound **VI** in the steric-zipper model showed that it exhibited a Y-shaped conformation. Benzyl, phenoxy, and pyridine aromatic rings undergo π - π stacked interactions with phenylalanine rings on either side of steric-zipper interface. Significantly, the isopentyl side chain undergoes several nonpolar contacts with side chains of

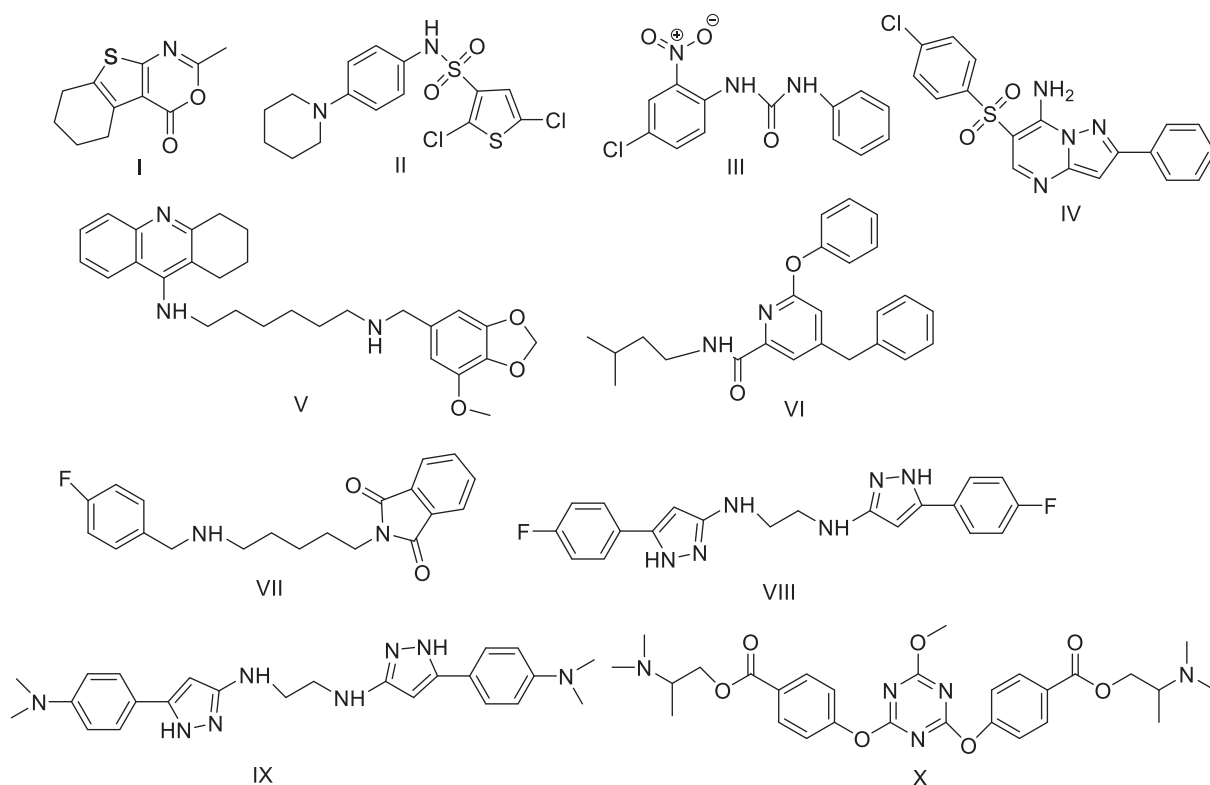


Fig. 8 Recently developed some small-molecule amyloid β -aggregation inhibitors.

phenylalanine and valine, respectively, and the pyridine nitrogen forms hydrogen bonds with lysine side chains.^{87,90}

Inspired by the chemical structure of the cholinesterase inhibitor donepezil (►Table 1), Malawska and coworkers developed heterodimeric isoindoline-1,3-dione derivatives of AChE inhibitors with anti-A β aggregation activity.⁹¹ Molecular modeling of the most potent isoindoline-1,3-dione derivative VII (2-(5-(4-fluorobenzylamino)pentyl)isoindoline-1,3-dione) (►Fig. 8) showed that it exhibited a linear binding mode with the steric-zipper assembly. The isoindole ring undergoes multiple π -alkyl and π - π interactions with side chains of valine and phenylalanine, respectively, with <5Å distance. The fluorobenzene substituent undergoes π -alkyl interactions with valine side chains on either side and more interestingly, the fluorine atom forms hydrogen bonds with lysine side chains. These multiple polar and nonpolar contacts are able to stabilize the steric-zipper assembly, which can lead to conformational changes and reduce the cytotoxicity of A β aggregates.^{87,91}

In a recent study, Muhs and coworkers used a rational design approach to develop small molecules based on a 3-aminopyrazole scaffold.⁹² The idea was to design small molecules that can undergo complimentary interactions with the donor-acceptor-donor hydrogen bonding pattern seen in the β -sheet assembly of A β . This approach led to the identification of dimeric 3-aminopyrazole derivatives VIII and IX (►Fig. 8) that were able to prevent A β oligomerization, fibril formation, and reduce cytotoxicity.^{87,92} The most potent compound IX formed a more stable complex in the steric-zipper octamer assembly, exhibited a linear conformation, and is able to fit nicely at the steric-zipper interface. The phenylpyrazole moieties underwent π - π , π -alkyl, and π -cation interactions with phenylalanine, valine, and lysine amino acid residues respectively on either side of the steric-zipper interface. These results support anti-A β aggregation properties of compound IX.^{87,93}

A novel small library of triazine-based small molecules was reported as multitargeting agents with dual cholinesterase and amyloid inhibition.⁹³ The most potent triazine compound X (►Fig. 8) undergoes favorable interactions with several amino acids located at both the C- and N-terminals the A β -dimer model due to its larger size of the compound. The central triazine ring with two dimethylaminopropyl benzoate units was oriented closer to the ¹⁶KLVFFAED²¹ region with the triazine and benzoate aromatic rings undergoing π - π stacking contacts with phenylalanine and alanine side chains. However, the presence of three ionizable groups (tertiary amines) will diminish its blood-brain barrier permeability.^{87,93}

Future Perspectives

Currently there were 132 agents in 156 trials of anti-AD therapies.⁹⁴ According to the Cummings et al's review, in 2018 a total of 26 agents were in phase III trial. Among them 54% are anti-amyloid (three are antiaggregation).⁹⁵ Recent reviews show that lessons are learned from all trials; even negative and futile outcomes are highly informative and provide guidance for future trials.⁹⁴ Although the results of

ongoing clinical trials are inconclusive, these compounds hold the promise of a new day in the development of disease-modifying therapies for AD. As our knowledge of their molecular structures and the molecular interactions responsible for activity of small molecules advances, more new generation of small-molecule A β -aggregation inhibitors will be developed in the near future. Effective drugs will have a dramatic impact on the number of persons affected in the future and also the quality of life of the AD patients. Advanced research in the field of AD will lead us to a better future for aging populations.

Conclusion

AD is still an incurable neurodegenerative disorder that has proved challenging to manage and treat with current therapies. Disease modification is the ultimate goal for AD drug development but has, so far, remained elusive. In this circumstance, small-molecule A β -aggregation inhibitors could be the key to treat AD.

Conflict of Interest

The authors declare no conflict of interest.

Reference

- Godyń J, Jończyk J, Panek D, Malawska B. Therapeutic strategies for Alzheimer's disease in clinical trials. *Pharmacol Rep* 2016;68(01):127-138
- Baharloo F, Moslem MH, Nadri H, et al. Benzofuran-derived benzylpyridinium bromides as potent acetylcholinesterase inhibitors. *Eur J Med Chem* 2015;93:196-201
- Reitz C, Brayne C, Mayeux R. Epidemiology of Alzheimer disease. *Nat Rev Neurol* 2011;7(03):137-152
- Wimo A, Jönsson L, Bond J, Prince M, Winblad B; Alzheimer Disease International. The worldwide economic impact of dementia 2010. *Alzheimers Dement* 2013;9(01):1.e3-11.e3
- Prince MJ, Acosta D, Castro-Costa E, Jackson J, Shaji KS. Packages of care for dementia in low- and middle-income countries. *PLoS Med* 2009;6(11):e1000176
- Brookmeyer R, Johnson E, Ziegler-Graham K, Arrighi HM. Forecasting the global burden of Alzheimer's disease. *Alzheimers Dement* 2007;3(03):186-191
- Ahmad SS, Akhtar S, Jamal QM, et al. Multiple targets for the management of Alzheimer's disease. *CNS Neurol Disord Drug Targets* 2016;15(10):1279-1289
- Meyer PF, Savard M, Poirier J, Morgan D, Breitner J; Alzheimer's Disease Neuroimaging Initiative. Hypothesis: cerebrospinal fluid protein markers suggest a pathway toward symptomatic resilience to AD pathology. *Alzheimers Dement* 2019;•••:1-12
- Chen GF, Xu TH, Yan Y, et al. Amyloid beta: structure, biology and structure-based therapeutic development. *Acta Pharmacol Sin* 2017;38(09):1205-1235
- Haass C, Selkoe DJ. Cellular processing of beta-amyloid precursor protein and the genesis of amyloid beta-peptide. *Cell* 1993;75(06):1039-1042
- Glennner GG, Wong CW. Alzheimer's disease: initial report of the purification and characterization of a novel cerebrovascular amyloid protein. 1984. *Biochem Biophys Res Commun* 2012;425(03):534-539
- Selkoe DJ, Hardy J. The amyloid hypothesis of Alzheimer's disease at 25 years. *EMBO Mol Med* 2016;8(06):595-608
- Viswanathan A, Greenberg SM. Cerebral amyloid angiopathy in the elderly. *Ann Neurol* 2011;70(06):871-880

- 14 Nunan J, Small DH. Regulation of APP cleavage by alpha-, beta- and gamma-secretases. *FEBS Lett* 2000;483(01):6–10
- 15 Reitz C. Alzheimer's disease and the amyloid cascade hypothesis: a critical review. *Int J Alzheimers Dis* 2012;2012:369808
- 16 About a peculiar disease of the cerebral cortex. By Alois Alzheimer, 1907 (Translated by L. Jarvik and H. Greenson). *Alzheimer Dis Assoc Disord* 1987;1(01):3–8
- 17 Michaelis ML. Drugs targeting Alzheimer's disease: some things old and some things new. *J Pharmacol Exp Ther* 2003;304(03):897–904
- 18 Murphy MP, LeVine H III. Alzheimer's disease and the amyloid- β peptide. *J Alzheimers Dis* 2010;19(01):311–323
- 19 Robinson DM, Keating GM. Memantine: a review of its use in Alzheimer's disease. *Drugs* 2006;66(11):1515–1534
- 20 Ghezzi L, Scarpini E, Galimberti D. Disease-modifying drugs in Alzheimer's disease. *Drug Des Devel Ther* 2013;7:1471–1478
- 21 Stites SD, Rubright JD, Karlawish J. What features of stigma do the public most commonly attribute to Alzheimer's disease dementia? Results of a survey of the U.S. general public. *Alzheimers Dement* 2018;14(07):925–932
- 22 Lundkvist J, Halldin MM, Sandin J, et al. The battle of Alzheimer's disease - the beginning of the future unleashing the potential of academic discoveries. *Front Pharmacol* 2014;5:102
- 23 Schneider LS, Mangialasche F, Andreasen N, et al. Clinical trials and late-stage drug development for Alzheimer's disease: an appraisal from 1984 to 2014. *J Intern Med* 2014;275(03):251–283
- 24 Zheng H, Fridkin M, Youdim M. New approaches to treating Alzheimer's disease. *Perspect Medicin Chem* 2015;7:1–8
- 25 Wisniewski T, Goñi F. Immunotherapy for Alzheimer's disease. *Biochem Pharmacol* 2014;88(04):499–507
- 26 Jia Q, Deng Y, Qing H. Potential therapeutic strategies for Alzheimer's disease targeting or beyond β -amyloid: insights from clinical trials. *BioMed Res Int* 2014;2014:837157
- 27 Kumar A, Singh A, Ekavali. A review on Alzheimer's disease pathophysiology and its management: an update. *Pharmacol Rep* 2015;67(02):195–203
- 28 Re F, Airoidi C, Zona C, et al. Beta amyloid aggregation inhibitors: small molecules as candidate drugs for therapy of Alzheimer's disease. *Curr Med Chem* 2010;17(27):2990–3006
- 29 Judge D, Roberts J, Khandker R, Ambegaonkar B, Black CM. Physician perceptions about the barriers to prompt diagnosis of mild cognitive impairment and Alzheimer's disease. *Int J Alzheimers Dis* 2019;2019:3637954
- 30 Bush AI. Metal complexing agents as therapies for Alzheimer's disease. *Neurobiol Aging* 2002;23(06):1031–1038
- 31 Taniguchi S, Suzuki N, Masuda M, et al. Inhibition of heparin-induced tau filament formation by phenothiazines, polyphenols, and porphyrins. *J Biol Chem* 2005;280(09):7614–7623
- 32 Pollack SJ, Sadler IJ, Hawtin SR, Tailor VJ, Shearman MS. Sulfonated dyes attenuate the toxic effects of β -amyloid in a structure-specific fashion. *Neurosci Lett* 1995;197(03):211–214
- 33 Porat Y, Abramowitz A, Gazit E. Inhibition of amyloid fibril formation by polyphenols: structural similarity and aromatic interactions as a common inhibition mechanism. *Chem Biol Drug Des* 2006;67(01):27–37
- 34 Reinke AA, Gestwicki JE. Structure-activity relationships of amyloid beta-aggregation inhibitors based on curcumin: influence of linker length and flexibility. *Chem Biol Drug Des* 2007;70(03):206–215
- 35 Wu C, Scott J, Shea JE. Binding of Congo red to amyloid protofibrils of the Alzheimer A β (9–40) peptide probed by molecular dynamics simulations. *Biophys J* 2012;103(03):550–557
- 36 Lorenzo A, Yankner BA. Beta-amyloid neurotoxicity requires fibril formation and is inhibited by congo red. *Proc Natl Acad Sci U S A* 1994;91(25):12243–12247
- 37 Xie Y, Deng S, Chen Z, Yan S, Landry DW. Identification of small-molecule inhibitors of the A β -ABAD interaction. *Bioorg Med Chem Lett* 2006;16(17):4657–4660
- 38 Bu XL, Rao PPN, Wang YJ. Anti-amyloid aggregation activity of natural compounds: implications for Alzheimer's drug discovery. *Mol Neurobiol* 2016;53(06):3565–3575
- 39 Velander P, Wu L, Henderson F, Zhang S, Bevan DR, Xu B. Natural product-based amyloid inhibitors. *Biochem Pharmacol* 2017;139:40–55
- 40 Savelieff MG, DeToma AS, Derrick JS, Lim MH. The ongoing search for small molecules to study metal-associated amyloid- β species in Alzheimer's disease. *Acc Chem Res* 2014;47(08):2475–2482
- 41 Korshavn KJ, Jang M, Kwak YJ, et al. Reactivity of metal-free and metal-associated amyloid-beta with glycosylated polyphenols and their esterified derivatives. *Sci Rep* 2015;5:17842
- 42 Nie Q, Du XG, Geng MY. Small molecule inhibitors of amyloid β peptide aggregation as a potential therapeutic strategy for Alzheimer's disease. *Acta Pharmacol Sin* 2011;32(05):545–551
- 43 NIH. U.S National Library of Medicine. Clinical Trials.gov publication. Safety and efficacy study of ALZT-OP1 in subjects with evidence of early Alzheimer's disease (NCT02547818). Available at: <https://clinicaltrials.gov/ct2/show/NCT02547818>. Accessed September 11, 2015
- 44 NIH. U.S National Library of Medicine. Clinical Trials.gov publication. Safety, efficacy and dose titration of Sodium Oligo-mannurate capsule on mild to moderate Alzheimer's disease (NCT01453569). Available at: <https://clinicaltrials.gov/ct2/show/NCT01453569>. Accessed October 18, 2011.
- 45 NIH. U.S National Library of Medicine. Clinical Trials.gov publication. An efficacy and safety study of sodium Oligo-mannurate (GV-971) capsule for the treatment of Alzheimer's disease (NCT02293915). Available at: <https://clinicaltrials.gov/ct2/show/NCT02293915>. Accessed November 19, 2014
- 46 NIH. U.S National Library of Medicine. Clinical Trials.gov publication. A study of single and multiple doses of KHK6640 in subjects with prodromal or mild to moderate Alzheimer's disease (NCT02127476). Available at: <https://clinicaltrials.gov/ct2/show/NCT02127476>. Accessed April 30, 2014
- 47 Bachurin SO, Bovina EV, Ustyugov AA. Drugs in clinical trials for Alzheimer's disease: the major trends. *Med Res Rev* 2017;37(05):1186–1225
- 48 Sinha S, Du Z, Maiti P, et al. Comparison of three amyloid assembly inhibitors: the sugar scyllo-inositol, the polyphenol epigallocatechin gallate, and the molecular tweezer CLR01. *ACS Chem Neurosci* 2012;3(06):451–458
- 49 Maccacchini ML, Chang MY, Pan C, John V, Zetterberg H, Greig NH. Posiphen as a candidate drug to lower CSF amyloid precursor protein, amyloid- β peptide and τ levels: target engagement, tolerability and pharmacokinetics in humans. *J Neurol Neurosurg Psychiatry* 2012;83(09):894–902
- 50 Bachurin SO, Bovina EV, Ustyugov AA. Current trends in the development of drugs for the treatment of Alzheimer's disease and their clinical trials. *Biomed Chem Res Methods* 2018;1(03):e00015
- 51 Suhr OB, Lundgren E, Westermark P. Transthyretin amyloidosis: new strategies for therapeutic intervention. *Drugs Future* 2010;35:325
- 52 Vartiainen NE, Williams M, Charles ML, Stenius T, Nurmi A, Yrjanheikki J. SAN-61 protects against A β 1–42, OGD and H2O2 in rat mixed cortical cultures and inhibits GSK-3 β . Paper presented at: 39th Society for Neuroscience Meeting; October 16, 2009; Chicago, United States
- 53 Hu Q, Cam J, Lake T, et al. Identification of Exebryl-1 and other novel small molecules as tau protein aggregation inhibitors. *Alzheimer's & Dementia: J Alzheimer's Assoc* 2011;4:481
- 54 Zou K, Gong JS, Yanagisawa K, Michikawa M. A novel function of monomeric amyloid beta-protein serving as an antioxidant molecule against metal-induced oxidative damage. *J Neurosci* 2002;22(12):4833–4841
- 55 Danielsson J, Jarvet J, Damberg P, Gräslund A. The Alzheimer beta-peptide shows temperature-dependent transitions between left-

- handed 3-helix, beta-strand and random coil secondary structures. *FEBS J* 2005;272(15):3938–3949
- 56 Yang DS, Yip CM, Huang TH, Chakrabartty A, Fraser PE. Manipulating the amyloid-beta aggregation pathway with chemical chaperones. *J Biol Chem* 1999;274(46):32970–32974
- 57 Liu YH, Giunta B, Zhou HD, Tan J, Wang YJ. Immunotherapy for Alzheimer disease: the challenge of adverse effects. *Nat Rev Neurol* 2012;8(08):465–469
- 58 Xu Y, Shen J, Luo X, et al. Conformational transition of amyloid beta-peptide. *Proc Natl Acad Sci U S A* 2005;102(15):5403–5407
- 59 Landau M, Sawaya MR, Faull KF, et al. Towards a pharmacophore for amyloid. *PLoS Biol* 2011;9(06):e1001080
- 60 Rajasekhar K, Suresh SN, Manjithaya R, Govindaraju T. Rationally designed peptidomimetic modulators of $\text{A}\beta$ toxicity in Alzheimer's disease. *Sci Rep* 2015;5:8139
- 61 Arai T, Sasaki D, Araya T, Sato T, Sohma Y, Kanai M. A cyclic KLVFF-derived peptide aggregation inhibitor induces the formation of less-toxic off-pathway amyloid- β oligomers. *ChemBioChem* 2014;15(17):2577–2583
- 62 Gazit E. Mechanistic studies of the process of amyloid fibrils formation by the use of peptide fragments and analogues: implications for the design of fibrillization inhibitors. *Curr Med Chem* 2002;9(19):1725–1735
- 63 Gazit E. Global analysis of tandem aromatic octapeptide repeats: the significance of the aromatic-glycine motif. *Bioinformatics* 2002;18(06):880–883
- 64 Sarasa M, Pesini P. Natural non-transgenic animal models for research in Alzheimer's disease. *Curr Alzheimer Res* 2009;6(02):171–178
- 65 Dai X, Chang P, Liu W, et al. $\text{A}\beta$ -40 Y10F increases β fibrils formation but attenuates the neurotoxicity of amyloid- β peptide. *Int J Mol Sci* 2012;13(05):5324–5337
- 66 Barnham KJ, Haeflner F, Ciccotosto GD, et al. Tyrosine gated electron transfer is key to the toxic mechanism of Alzheimer's disease beta-amyloid. *FASEB J* 2004;18(12):1427–1429
- 67 Schöneich C. Methionine oxidation by reactive oxygen species: reaction mechanisms and relevance to Alzheimer's disease. *Biochim Biophys Acta* 2005;1703(02):111–119
- 68 Yang DS, McLaurin J, Qin K, Westaway D, Fraser PE. Examining the zinc binding site of the amyloid-beta peptide. *Eur J Biochem* 2000;267(22):6692–6698
- 69 Cheng B, Gong H, Xiao H, Petersen RB, Zheng L, Huang K. Inhibiting toxic aggregation of amyloidogenic proteins: a therapeutic strategy for protein misfolding diseases. *Biochim Biophys Acta* 2013;1830(10):4860–4871
- 70 Yanagisawa D, Taguchi H, Yamamoto A, Shirai N, Hirao K, Tooyama I. Curcuminoid binds to amyloid- β 1-42 oligomer and fibril. *J Alzheimers Dis* 2011;24(Suppl 2):33–42
- 71 Masuda Y, Fukuchi M, Yatagawa T, et al. Solid-state NMR analysis of interaction sites of curcumin and 42-residue amyloid β -protein fibrils. *Bioorg Med Chem* 2011;19(20):5967–5974
- 72 Orlando RA, Gonzales AM, Royer RE, Deck LM, Vander Jagt DL. A chemical analog of curcumin as an improved inhibitor of amyloid Abeta oligomerization. *PLoS One* 2012;7(03):e31869
- 73 Sato T, Kienlen-Campard P, Ahmed M, et al. Inhibitors of amyloid toxicity based on beta-sheet packing of Abeta40 and Abeta42. *Biochemistry* 2006;45(17):5503–5516
- 74 Zhao LN, Chiu SW, Benoit J, Chew LY, Mu Y. The effect of curcumin on the stability of $\text{A}\beta$ dimers. *J Phys Chem B* 2012;116(25):7428–7435
- 75 Rao PP, Mohamed T, Teckwani K, Tin G. Curcumin binding to beta amyloid: a computational study. *Chem Biol Drug Des* 2015;86(04):813–820
- 76 Ge JF, Qiao JP, Qi CC, Wang CW, Zhou JN. The binding of resveratrol to monomer and fibril amyloid beta. *Neurochem Int* 2012;61(07):1192–1201
- 77 Feng Y, Wang XP, Yang SG, et al. Resveratrol inhibits beta-amyloid oligomeric cytotoxicity but does not prevent oligomer formation. *Neurotoxicology* 2009;30(06):986–995
- 78 Ladiwala AR, Lin JC, Bale SS, et al. Resveratrol selectively remodels soluble oligomers and fibrils of amyloid Abeta into off-pathway conformers. *J Biol Chem* 2010;285(31):24228–24237
- 79 Ladiwala AR, Mora-Pale M, Lin JC, et al. Polyphenolic glycosides and aglycones utilize opposing pathways to selectively remodel and inactivate toxic oligomers of amyloid β . *ChemBioChem* 2011;12(11):1749–1758
- 80 Rezaei-Zadeh K, Shytle D, Sun N, et al. Green tea epigallocatechin-3-gallate (EGCG) modulates amyloid precursor protein cleavage and reduces cerebral amyloidosis in Alzheimer transgenic mice. *J Neurosci* 2005;25(38):8807–8814
- 81 Bieschke J, Russ J, Friedrich RP, et al. EGCG remodels mature alpha-synuclein and amyloid-beta fibrils and reduces cellular toxicity. *Proc Natl Acad Sci U S A* 2010;107(17):7710–7715
- 82 Rauk A. Why is the amyloid beta peptide of Alzheimer's disease neurotoxic? *Dalton Trans* 2008;(10):1273–1282
- 83 Shaw JE, Chio J, Dasgupta S, et al. $\text{A}\beta$ (1-42) assembly in the presence of scyllo-inositol derivatives: identification of an oxime linkage as important for the development of assembly inhibitors. *ACS Chem Neurosci* 2012;3(03):167–177
- 84 Sun Y, Zhang G, Hawkes CA, Shaw JE, McLaurin J, Nitz M. Synthesis of scyllo-inositol derivatives and their effects on amyloid beta peptide aggregation. *Bioorg Med Chem* 2008;16(15):7177–7184
- 85 O'Hare E, Scopes DI, Treherne JM, Norwood K, Spanswick D, Kim EM. RS-0406 arrests amyloid- β oligomer-induced behavioural deterioration in vivo. *Behav Brain Res* 2010;210(01):32–37
- 86 Nakagami Y, Nishimura S, Murasugi T, et al. A novel beta-sheet breaker, RS-0406, reverses amyloid beta-induced cytotoxicity and impairment of long-term potentiation in vitro. *Br J Pharmacol* 2002;137(05):676–682
- 87 Mohamed T, Shakeri A, Rao PP. Amyloid cascade in Alzheimer's disease: recent advances in medicinal chemistry. *Eur J Med Chem* 2016;113:258–272
- 88 López LC, Dos-Reis S, Espargaró A, et al. Discovery of novel inhibitors of amyloid β -peptide 1-42 aggregation. *J Med Chem* 2012;55(22):9521–9530
- 89 Luo W, Li YP, He Y, et al. Design, synthesis and evaluation of novel tacrine-multialkoxybenzene hybrids as dual inhibitors for cholinesterases and amyloid beta aggregation. *Bioorg Med Chem* 2011;19(02):763–770
- 90 Arai T, Araya T, Sasaki D, et al. Rational design and identification of a non-peptidic aggregation inhibitor of amyloid β based on a pharmacophore motif obtained from cyclo[$-\text{Lys-Leu-Val-Phe-Phe-}$]. *Angew Chem Int Ed Engl* 2014;53(31):8236–8239
- 91 Guziar N, Bajda M, Skrok M, et al. Development of multifunctional, heterodimeric isoindoline-1,3-dione derivatives as cholinesterase and β -amyloid aggregation inhibitors with neuroprotective properties. *Eur J Med Chem* 2015;92:738–749
- 92 Kroth H, Ansaloni A, Varisco Y, et al. Discovery and structure activity relationship of small molecule inhibitors of toxic β -amyloid-42 fibril formation. *J Biol Chem* 2012;287(41):34786–34800
- 93 Veloso AJ, Dhar D, Chow AM, et al. sym-Triazines for directed multitarget modulation of cholinesterases and amyloid- β in Alzheimer's disease. *ACS Chem Neurosci* 2013;4(02):339–349
- 94 Cummings J, Lee G, Ritter A, Sabbagh M, Zhong K. Alzheimer's disease drug development pipeline: 2019. *Alzheimers Dement (N Y)* 2019;5:272–293
- 95 Cummings J, Lee G, Ritter A, Zhong K. Alzheimer's disease drug development pipeline: 2018. *Alzheimers Dement (N Y)* 2018;4:195–214