# First Total Synthesis and Investigation of the X-ray Crystal Structure of the Pyrano[3,2-a]carbazole Alkaloid Clausenalansine A

Valerie Lösle[a]
Olga Kataeva[b]
Hans-Joachim Knölker*[a]

[a] Faculty of Chemistry, Technische Universität Dresden, Bergstraße 66, 01069 Dresden, Germany
 hans-joachim.knoelker@tu-dresden.de
[b] A. E. Arbuzov Institute of Organic and Physical Chemistry, FRC Kazan Scientific Center, Russian Academy of Sciences, Arbuzov Str. 8, Kazan 420088, Russian Federation

Dedicated to Professor Tomáš Hudlický on the occasion of his 71st birthday
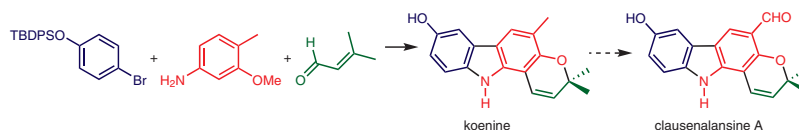
koenine   clausenalansine A

Abstract   We describe the first total synthesis of the recently discovered pyrano[3,2-a]carbazole alkaloid clausenalansine A. The synthetic strategy for the construction of this formylpyrano[3,2-a]carbazole is based on a sequence of Buchwald–Hartwig coupling, palladium(II)-catalyzed oxidative cyclization, Lewis acid promoted annulation of the pyran ring, and chemoselective oxidation of a methyl to a formyl group.

Key words carbazoles, alkaloids, annulation, catalysis, cyclization, natural products, total synthesis, palladium

Carbazole alkaloids have been the focus of research for a long time because of their broad range of useful biological activities. They have been isolated from various natural sources including terrestrial plants, microorganisms, slime molds, and algae.[1,2] Among the former, especially plants of the genera *Murraya*, *Clausena*, and *Glycosmis*, all belonging to the family Rutaceae, were found to be extremely rich in diverse carbazole alkaloids. Biogenetically, all the carbazole natural products from terrestrial plants derive from 3-methylcarbazole as parent compound. Subsequent oxygenation at different positions of the carbazole framework, oxidation of the methyl group, and prenylation or geranylation followed by annulation of additional rings along the biogenetic pathway provide the broad structural variety of carbazoles found in plants of the Rutaceae family.[2] Over the past decades, many examples of monooxygenated pyrano[3,2-a]carbazole alkaloids with a C_1 moiety at the 3-position as represented by compounds **1–4** have been isolated from nature (Figure 1).[3–6] Only very recently, Fu and co-workers described the isolation of the novel clausenalansine A (**1**), along with the previously known pyrano[3,2-a]carbazoles clauraila E (**3**)[5] and murrayamine-A (mukoenine-C) (**4**),[6] from the fruits of *Clausena lansium*.[3] The

fruits of this evergreen tree growing in Southeast Asia are very popular and reported to have health-promoting effects. In fact, biological screenings revealed a remarkable neuroprotective activity of clausenalansine A (**1**).[3] Thus, compound **1** might represent a potential lead structure for the development of new agents for the prevention and treatment of neurodegenerative diseases like Parkinson's.[3]


1 clausenalansine A   **2** koenine

3 clauraila E   **4** murrayamine-A (mukoenine-C)
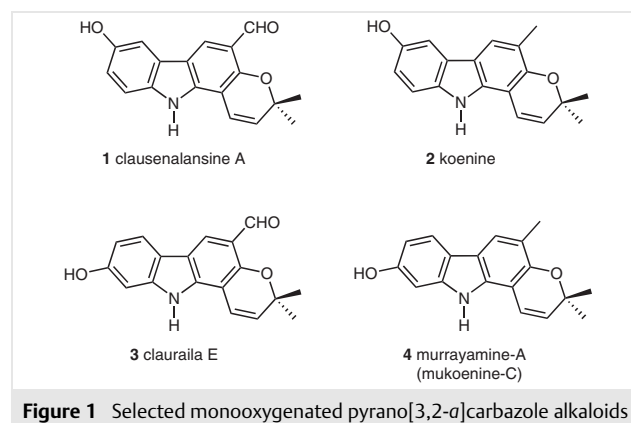
Figure 1   Selected monooxygenated pyrano[3,2-a]carbazole alkaloids

Clausenalansine A (**1**) is an oxidized derivative of koenine (**2**), a known pyrano[3,2-a]carbazole alkaloid first isolated by Narasimhan and co-workers from the leaves of *Murraya koenigii*.[4] In 2016, we described an efficient palladium-catalyzed total synthesis of koenine (**2**).[7] The two related alkaloids, clauraila E (**3**) and murrayamine-A (**4**),[5,6] were previously also synthesized by our group.[8,9] In the present work,[10] we describe the first synthesis of clausenalansine A (**1**) and an investigation of its X-ray crystal structure.

Retrosynthetic analysis of clausenalansine A (**1**) leads to silyl-protected koenine **5** as precursor which by oxidation of the methyl group and subsequent deprotection should afford **1** (Scheme 1). The pyrano[3,2-a]carbazole **5** would

derive from the orthogonally diprotected 2,6-dihydroxycar-bazole **6** via regioselective deprotection of the 2-hydroxy group and annulation of the pyran ring. Compound **6** can be prepared from the protected *p*-bromophenol **7** and aryl-amine **8** via a sequence of Buchwald–Hartwig coupling[11] and palladium(II)-catalyzed oxidative cyclization.[7,12]
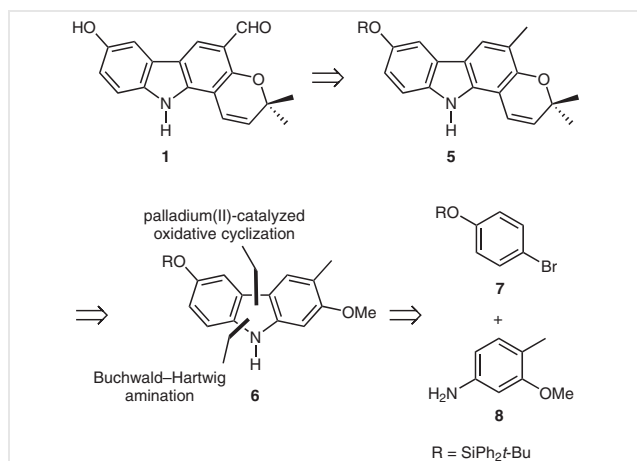
Scheme 1 Retrosynthetic analysis of clausenalansine A (**1**)

The two starting materials were readily available. Silyla-tion of commercially available 4-bromophenol provided si-lyl-protected *p*-bromophenol **7** and Béchamp reduction of 2-methoxy-4-nitrotoluene gave 3-methoxy-4-methyl-aniline (**8**).[7,13] Buchwald–Hartwig coupling of compound **7** with arylamine **8** in the presence of catalytic amounts of DavePhos [2-dicyclohexylphosphino-2′-(*N*,*N*-dimethylami-no)biphenyl] afforded diarylamine **9** (Scheme 2). Palladi-um(II)-catalyzed oxidative cyclization of **9** using pivalic acid as solvent under microwave conditions provided the or-thogonally diprotected 2,6-dihydroxycarbazole **6**. The pres-ent protocol for this transformation is better scalable and reproducible compared to the previous one.[7] Cleavage of the methyl ether of **6** using boron tribromide afforded 2-hy-droxycarbazole **10**. Annulation of the pyran moiety by treatment with prenal and titanium tetraisopropoxide fol-lowing Casiraghi's method[13,14] assembled the pyrano[3,2-*a*]carbazole framework and led to silyl-protected koenine **5**. Subsequently, we investigated the oxidation of **5** with 2,3-dichloro-5,6-dicyano-1,4-benzoquinone (DDQ) to the cor-responding formyl derivative **11** (Table 1). Using 2.2 equiva-lents of the oxidizing agent and our previously optimized conditions (reaction in a solvent mixture of MeOH, THF, and H$_2$O at room temperature),[15] the conversion proceeded sur-prisingly fast. However, after a reaction time of 1 hour, com-pound **11** was obtained in only 37% yield with no recovered starting material. A lower reaction temperature (0 °C) led to an increase of the yield to 80%. Further attempts to improve the yield by extension of the reaction time gave no better results.
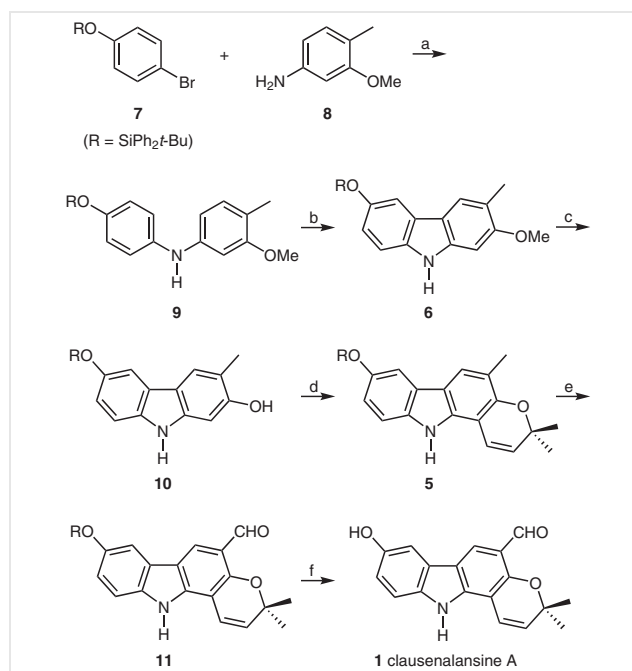
Scheme 2 Synthesis of clausenalansine A (**1**). *Reagents and conditions*: a) **8** (1.3 equiv), Pd$_2$(dba)$_3$ (6 mol%), DavePhos (13 mol%), NaO*t*-Bu (1.5 equiv), toluene, reflux, 16 h (93%); b) Pd(OAc)$_2$ (21 mol%), Cu(OAc)$_2$ (2.6 equiv), PivOH, 130 °C, MW (300 W), 1 h (70%); c) BBr$_3$ (5.0 equiv), CH$_2$Cl$_2$, −78 °C to r.t., 26 h (80%); d) prenal (2.2 equiv), Ti(O*i*-Pr)$_4$ (3.9 equiv), toluene, −78 °C to r.t., 4 h (72%); e) DDQ (2.2 equiv), MeOH/THF/H$_2$O (10:3:1), 0 °C, 1 h (80%); f) TBAF (1.2 equiv), DMF, −5 °C, 20 min (99%).

Table 1 Oxidation of Silyl-Protected Koenine **5** to Formyl Derivative **11**

| Reagents and conditions	Yield (%) of **11**
DDQ (2.2 equiv), MeOH/THF/H$_2$O (10:3:1), r.t., 1 h	37
DDQ (2.2 equiv), MeOH/THF/H$_2$O (10:3:1), 0 °C, 1 h	80
DDQ (2.2 equiv), MeOH/THF/H$_2$O (10:3:1), 0 °C, 2 h	70

Finally, deprotection of the hydroxyl group of **11** by re-action with tetrabutylammonium fluoride (TBAF) in DMF provided clausenalansine A (**1**). A comparison of the spec-troscopic data of our synthetic **1** with those described for the isolated natural product showed a perfect agreement.[3]

The structure of clausenalansine A (**1**) has been addi-tionally confirmed by an X-ray crystal structure determina-tion of single crystals (Figure 2).[16]

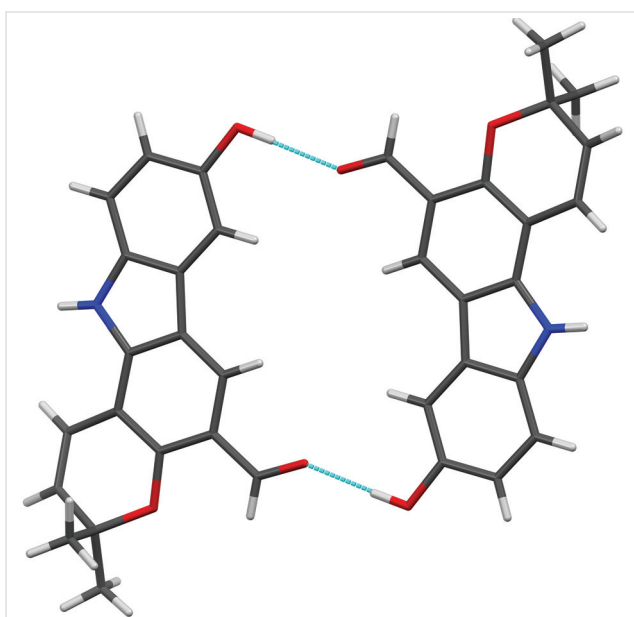The crystal contains two independent molecules A and B in the asymmetric part of the unit cell, which are bound by weak C–H···π and N–H···π hydrogen bonds (Figure 3). Symmetrically equivalent molecules are bound by strong hydrogen bonds and form 'head-to-tail' dimers (Figure 4). Moreover, symmetrically equivalent molecules in the crys-tal form stack via π···π interactions with a distance between
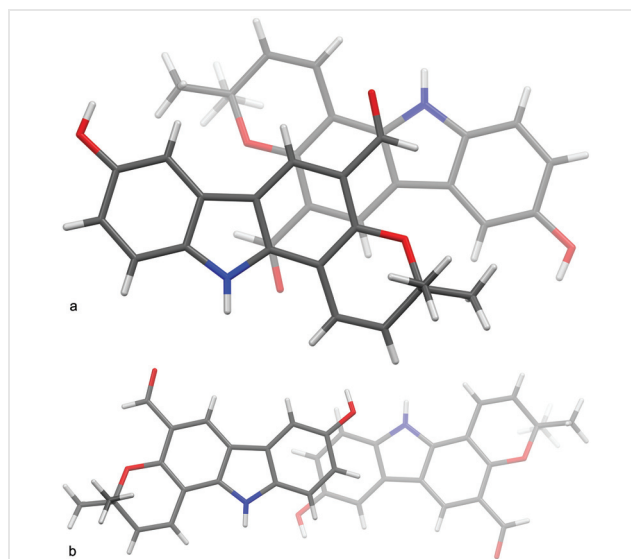
V. Lösle et al.
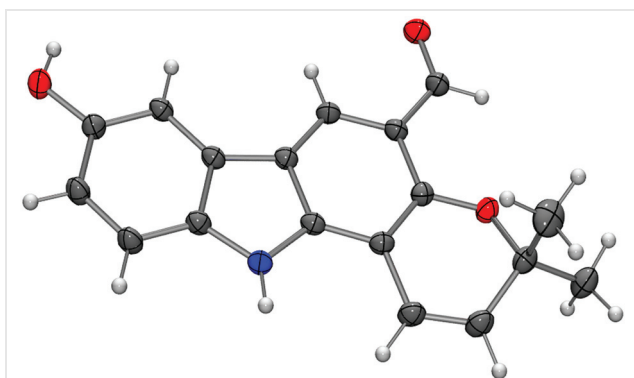
Figure 2 Molecular structure of clausenalansine A (**1**) in the crystal (triclinic, *P*1); ORTEP plot showing thermal ellipsoids at the 50% probability level.
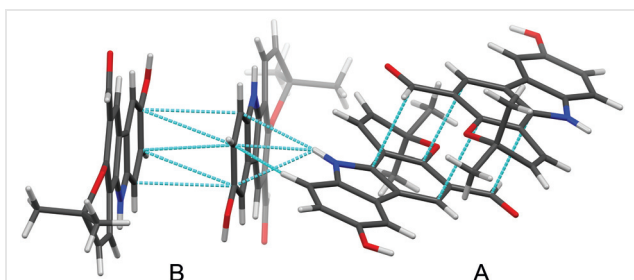
Figure 3 Intermolecular contacts of A and B molecules in the crystal showing π···π, N–H···π, and C–H···π interactions

Figure 4 Head-to-tail dimers of clausenalansine A (**1**) in the crystal

Figure 5 Top views of a fragment of the crystal packing showing the overlap of two independent molecules of A (a) and B (b)

the interacting carbon atoms and the planes of the neighboring carbazole fragments of about 3.4 Å (A molecules) and 3.3 Å (B molecules). It is noteworthy that the A molecules exhibit an ideal 'ring-over-atom' overlap of the inner six-membered rings (Figure 5a), whereas the B molecules overlap with their terminal benzo rings (Figure 5b).

In conclusion, we have described the first total synthesis of clausenalansine A (**1**) in seven steps and 30% overall yield starting from commercially available 4-bromophenol. Our synthetic approach includes the following key steps: Buchwald–Hartwig coupling, palladium(II)-catalyzed oxidative cyclization, Lewis acid promoted annulation of the pyran ring, and oxidation of a methyl group. The spectroscopic data of our synthetic clausenalansine A are in good agreement with those reported by Fu and co-workers for the natural product.[3] Thus, our total synthesis confirms the structural assignment for the natural product. Additionally, we have investigated the crystal structure of clausenalansine A (**1**) by X-ray analysis which has revealed a remarkable packing motif formed by π···π, N–H···π, and C–H···π interactions.

All reactions were carried out in oven-dried glassware under argon atmosphere using anhydrous solvents, unless stated otherwise. Pd(OAc)$_2$ was recrystallized from glacial acetic acid. All other chemicals were used as received from commercial sources. Microwave irradiations were carried out using a CEM DISCOVER microwave apparatus with a maximum power of 300 W and a maximum pressure of 20 bar. Flash chromatography was performed using silica gel from Acros Organics (0.035–0.070 mm). TLC was performed with TLC plates from Merck (60 F$_{254}$) using UV light for visualization. Melting points were measured on a Gallenkamp MPD 350 melting point apparatus. Ultraviolet spectra were recorded on a PerkinElmer 25 UV/Vis spectro-

V. Lösle et al.

meter. Fluorescence spectra were obtained using a Varian Cary Eclipse spectrometer. IR spectra were recorded on a Thermo Nicolet Avatar FT-IR spectrometer using the ATR method. NMR spectra were recorded on Bruker DXR 500 and Avance III 600 spectrometers. Chemical shifts δ are reported in ppm with the solvent signal as internal standard. Standard abbreviations are used to denote the multiplicities of the signals. EI mass spectra were recorded by GC-MS coupling using an Agilent Technologies 6890 N GC System equipped with a 5973 Mass Selective Detector (70 eV). ESI mass spectra were recorded on an Esquire LC system with an ion trap detector from Bruker; positive and negative ions were detected. Elemental analyses were measured on a EuroVector EuroEA3000 elemental analyzer. The X-ray crystal structure analysis was performed with a Bruker AXS D8 Venture instrument.

4-Bromophenyl *tert*-Butyldiphenylsilyl Ether (7)

Imidazole (2.37 g, 34.8 mmol) was added to a solution of 4-bromophenol (3.05 g, 17.6 mmol) in DMF (36 mL) and the mixture was stirred for 10 min at r.t., before *t*-BuPh$_2$SiCl (7.0 mL, 7.4 g, 27 mmol) was added dropwise. The mixture was stirred for a further 19 h at r.t. and then aq 1 M HCl (40 mL) was added. The aqueous layer was separated and extracted with CH$_2$Cl$_2$. The combined organic layers were washed with sat. aq NH$_4$Cl, then with brine, and dried over MgSO$_4$. Removal of the solvent in vacuo and column chromatography (silica gel, isohexane/EtOAc, 15:1 to 12:1) of the residue provided compound **7** as a colorless oil; yield: 7.24 g (100%).

^{1}H NMR (500 MHz, CDCl$_3$): δ = 1.09 (s, 9 H), 6.61–6.64 (m, 2 H), 7.15–7.18 (m, 2 H), 7.35–7.38 (m, 4 H), 7.41–7.45 (m, 2 H), 7.68–7.69 (m, 4 H).

^{13}C NMR and DEPT (125 MHz, CDCl$_3$): δ = 19.40 (C), 26.42 (3 CH$_3$), 113.32 (C), 121.46 (2 CH), 127.84 (4 CH), 130.03 (2 CH), 132.06 (2 CH), 132.40 (2 C), 135.44 (4 CH), 154.72 (C).

MS (EI): *m/z* (%) = 412 (9), 410 (9, [M$^+$]), 355 (100), 353 (100), 273 (76), 197 (29), 152 (23), 57 (25).

For further spectroscopic data, see ref. 7.

N-[4-(*tert*-Butyldiphenylsilyloxy)phenyl]-3-methoxy-4-methyl-aniline (9)

A solution of bromoarene **7** (2.01 g, 4.89 mmol) in toluene (28 mL) was added to a vigorously stirred solution of 3-methoxy-4-methylaniline (**8**)^{13} (878 mg, 6.40 mmol), Pd$_2$(dba)$_3$ (260 mg, 284 μmol), DavePhos (248 mg, 630 μmol), and NaO*t*-Bu (701 mg, 7.29 mmol) in toluene (66 mL) at reflux temperature. Stirring was continued at reflux for 16 h under argon and then for 4 h at r.t. under air. The reaction mixture was filtered over Celite and washed with EtOAc. The organic layer was washed with brine, dried over MgSO$_4$, and concentrated in vacuo. Column chromatography (silica gel, isohexane/EtOAc, 20:1) of the residue provided diarylamine **9** as a brown viscous oil; yield: 2.12 g (93%).

^{1}H NMR (500 MHz, DMSO-d_6): δ = 1.01 (s, 9 H), 1.98 (s, 3 H), 3.64 (s, 3 H), 6.37 (dd, J = 7.9, 2.2 Hz, 1 H), 6.43 (dd, J = 2.2, 0.3 Hz, 1 H), 6.59–6.62 (m, 2 H), 6.79–6.82 (m, 2 H), 6.84 (d, J = 8.2 Hz, 1 H), 7.39–7.46 (m, 6 H), 7.63–7.67 (m, 5 H).

^{13}C NMR and DEPT (125 MHz, DMSO-d_6): δ = 15.33 (CH$_3$), 18.94 (C), 26.38 (3 CH$_3$), 54.80 (CH$_3$), 99.13 (CH), 106.98 (CH), 115.62 (C), 118.90 (2 CH), 119.76 (2 CH), 127.96 (4 CH), 130.10 (2 CH), 130.49 (CH), 132.48 (2 C), 135.09 (4 CH), 137.41 (C), 143.72 (C), 148.48 (C), 157.77 (C).

MS (EI): *m/z* (%) = 467 (98, [M$^+$]), 410 (100), 205 (17), 136 (21).

For further spectroscopic data, see ref. 7.

6-(*tert*-Butyldiphenylsilyloxy)-2-methoxy-3-methyl-9*H*-carbazole (6)

Method A: A microwave reaction vial (10 mL pressure vessel, Pyrex, CEM Discover) was filled with diarylamine **9** (57.1 mg, 122 μmol), Pd(OAc)$_2$ (5.8 mg, 26 μmol), Cu(OAc)$_2$ (57.6 mg, 317 μmol), and PivOH (376 mg) under air, and the mixture was heated at 130 °C by microwave irradiation (300 W) for 1 h. After cooling to r.t., the mixture was washed with sat. aq K$_2$CO$_3$ and the aqueous layer was extracted several times with EtOAc. The combined organic layers were washed with brine, dried over MgSO$_4$, and concentrated in vacuo. Column chromatography (silica gel, pentane/EtOAc, 9:1 to 5:1) of the residue provided carbazole **6** as a brown viscous oil; yield: 39.7 mg (70%).

Method B: A round-bottom flask was filled with diarylamine **9** (3.29 g, 7.03 mmol), Pd(OAc)$_2$ (332 mg, 1.48 mmol), Cu(OAc)$_2$ (3.30 g, 18.2 mmol), and PivOH (22.0 g) under air, and the mixture was heated at 130 °C for 2 h while stirring vigorously. After cooling to r.t., the mixture was washed with sat. aq K$_2$CO$_3$ and the aqueous layer was extracted several times with EtOAc. The combined organic layers were washed with brine and dried over MgSO$_4$. Removal of the solvent and column chromatography (silica gel, pentane/EtOAc, 9:1 to 5:1) of the residue afforded carbazole **6** as a brown viscous oil; yield: 1.64 g (50%).

^{1}H NMR (500 MHz, DMSO-d_6): δ = 1.05 (s, 9 H), 2.18 (s, 3 H), 3.81 (s, 3 H), 6.65 (dd, J = 8.5, 2.6 Hz, 1 H), 6.84 (s, 1 H), 7.10 (d, J = 8.5 Hz, 1 H), 7.30 (d, J = 2.5 Hz, 1 H), 7.38–7.46 (m, 6 H), 7.53 (s, 1 H), 7.69–7.70 (m, 4 H), 10.73 (s, 1 H).

^{13}C NMR and DEPT (125 MHz, DMSO-d_6): δ = 16.97 (CH$_3$), 19.51 (C), 26.93 (3 CH$_3$), 55.70 (CH$_3$), 92.98 (CH), 109.04 (CH), 111.12 (CH), 115.38 (C), 116.52 (CH), 117.43 (C), 121.46 (CH), 123.50 (C), 128.37 (4 CH), 130.47 (2 CH), 133.27 (2 C), 135.01 (C), 135.59 (4 CH), 140.77 (C), 148.33 (C), 157.24 (C).

MS (EI): *m/z* (%) = 465 (51, [M$^+$]), 408 (100), 330 (8), 204 (12).

For further spectroscopic data, see ref. 7.

6-(*tert*-Butyldiphenylsilyloxy)-2-hydroxy-3-methyl-9*H*-carbazole (10)

To a solution of carbazole **6** (51.9 mg, 111 μmol) in CH$_2$Cl$_2$ (7.5 mL) at –78 °C was added 1 M BBr$_3$ in CH$_2$Cl$_2$ (0.56 mL, 560 μmol) dropwise over a period of 20 min. Stirring was continued for 30 min at –78 °C and then for 2 h at –10 °C. The mixture was allowed to warm to r.t. and stirred for a further 23 h. Then, MeOH (5.0 mL) was added under cooling. The organic layer was washed with water and brine, and the combined aqueous layers were extracted with CH$_2$Cl$_2$. The combined organic layers were dried over MgSO$_4$ and concentrated in vacuo. Column chromatography (silica gel, pentane/EtOAc, 4:1) of the residue provided 2-hydroxycarbazole **10** as a brown viscous oil; yield: 39.9 mg (80%).

^{1}H NMR (500 MHz, DMSO-d_6): δ = 1.04 (s, 9 H), 2.15 (s, 3 H), 6.62 (dd, J = 8.6, 2.6 Hz, 1 H), 6.75 (s, 1 H), 7.03 (d, J = 8.6 Hz, 1 H), 7.25 (d, J = 2.1 Hz, 1 H), 7.38–7.44 (m, 7 H), 7.68–7.70 (m, 4 H), 9.31 (s, 1 H), 10.53 (s, 1 H).

^{13}C NMR and DEPT (125 MHz, DMSO-d_6): δ = 16.43 (CH$_3$), 19.06 (C), 26.49 (3 CH$_3$), 95.85 (CH), 108.41 (CH), 110.36 (CH), 114.64 (C), 115.65 (CH), 115.97 (C), 121.01 (CH), 123.38 (C), 127.91 (4 CH), 130.00 (2 CH), 132.87 (2 C), 134.51 (C), 135.15 (4 CH), 140.51 (C), 147.74 (C), 154.75 (C).

MS (EI): *m/z* (%) = 451 (47, [M$^+$]), 394 (100), 316 (14), 197 (24).

For further spectroscopic data, see ref. 7.

V. Lösle et al.

8-(*tert*-Butyldiphenylsilyloxy)-3,3,5-trimethyl-3,11-dihydropyrano[3,2-*a*]carbazole (5)

To a stirred solution of 2-hydroxycarbazole **10** (70.3 mg, 156 µmol) in toluene (3.0 mL) at –78 °C was added 3-methylbut-2-enal (prenal) (28.7 mg, 341 µmol) and then slowly Ti(O*i*-Pr)$_4$ (0.18 mL, 0.17 g, 0.61 mmol). The reaction mixture was stirred for 10 min at –78 °C, then allowed to warm to r.t., and stirred for an additional 4 h. After hydrolysis by addition of water (3.0 mL), the organic layer was separated, and washed first with sat. aq NH$_4$Cl and then with water. The combined aqueous layers were extracted with EtOAc and the combined organic layers were then washed with brine and dried over MgSO$_4$. Removal of the solvent and column chromatography (silica gel, pentane/EtOAc, 8:1) of the residue provided pyrano[3,2-*a*]carbazole **5** as a yellow viscous oil; yield: 58.4 mg (72%).

^{1}H NMR (500 MHz, DMSO-*d*$_6$): δ = 1.07 (s, 9 H), 1.40 (s, 6 H), 2.18 (s, 3 H), 5.75 (dd, *J* = 9.8, 0.4 Hz, 1 H), 6.70 (ddd, *J* = 8.5, 2.5, 0.3 Hz, 1 H), 6.85 (d, *J* = 9.8 Hz, 1 H), 7.13 (dd, *J* = 8.5, 0.3 Hz, 1 H), 7.33 (dd, *J* = 2.5, 0.3 Hz, 1 H), 7.41–7.46 (m, 7 H), 7.71–7.73 (m, 4 H), 10.94 (s, 1 H).

^{13}C NMR and DEPT (125 MHz, DMSO-*d*$_6$): δ = 15.86 (CH$_3$), 19.12 (C), 26.53 (3 CH$_3$), 27.38 (2 CH$_3$), 75.63 (C), 104.28 (C), 108.82 (CH), 110.78 (CH), 115.81 (C), 116.45 (C, CH), 117.88 (CH), 120.86 (CH), 123.38 (C), 127.98 (4 CH), 128.89 (CH), 130.08 (2 CH), 132.86 (2 C), 134.88 (C), 135.21 (4 CH), 135.98 (C), 148.09 (C), 149.03 (C).

MS (EI): *m/z* (%) = 517 (33, [M$^+$]), 503 (75), 502 (100), 460 (19), 444 (17), 222 (28).

For further spectroscopic data, see ref. 7.

8-(*tert*-Butyldiphenylsilyloxy)-5-formyl-3,3-dimethyl-3,11-di-hydropyrano[3,2-*a*]carbazole (11)

Pyrano[3,2-*a*]carbazole **5** (93.1 mg, 180 µmol) was dissolved in a mixture of MeOH/THF/H$_2$O (10:3:1, 21 mL) and the solution was cooled to 0 °C. DDQ (92.5 mg, 407 µmol) was then added in portions and stirring was continued for 1 h at 0 °C. The reaction was quenched by addition of 10% aq NaOH, and the aqueous layer was separated and extracted with EtOAc. The combined organic layers were washed with brine, dried over MgSO$_4$, and concentrated in vacuo. Column chromatography (silica gel, pentane/EtOAc, 5:1 to 3:1) of the residue provided compound **11** as a light-yellow solid; yield: 76.5 mg (80%); mp 234–234.5 °C.

IR (ATR): 3281, 2959, 2929, 2855, 1589, 1460, 1139, 1114, 970, 890, 699, 613 cm^{-1}.

^{1}H NMR (600 MHz, DMSO-*d*$_6$): δ = 1.08 (s, 9 H), 1.48 (s, 6 H), 5.91 (d, *J* = 9.9 Hz, 1 H), 6.77 (dd, *J* = 8.7, 2.3 Hz, 1 H), 6.89 (d, *J* = 9.9 Hz, 1 H), 7.21 (dd, *J* = 8.7, 0.4 Hz, 1 H), 7.42–7.48 (m, 6 H), 7.54 (d, *J* = 2.3 Hz, 1 H), 7.71–7.73 (m, 4 H), 8.11 (s, 1 H), 10.31 (s, 1 H), 11.57 (s, 1 H).

^{13}C NMR and DEPT (151 MHz, DMSO-*d*$_6$): δ = 19.13 (C), 26.52 (3 CH$_3$), 27.22 (2 CH$_3$), 77.13 (C), 104.13 (C), 110.14 (CH), 111.48 (CH), 116.77 (CH), 117.25 (2 C), 117.37 (C), 118.12 (CH), 119.27 (CH), 123.88 (C), 128.03 (4 CH), 129.93 (CH), 130.17 (2 CH), 132.63 (C), 135.23 (4 CH), 135.68 (C), 141.22 (C), 149.18 (C), 153.77 (C), 187.91 (CHO).

MS (ESI, +50 V): *m/z* = 532.6 [M + H$^+$].

UV (MeOH): λ = 224, 288, 311, 370 (sh) nm.

Fluorescence (MeOH): λ$_{ex}$ = 311 nm, λ$_{em}$ = 358 nm.

Anal. Calcd for C$_{34}$H$_{33}$NO$_3$Si: C, 76.80; H, 6.26; N, 2.63. Found: C, 76.61; H, 6.17; N, 2.67.

Clausenalansine A (1)

A solution of compound **11** (30.4 mg, 57.2 µmol) in DMF (1.75 mL) was stirred at –5 °C under air and 1 M TBAF in THF (70 µL, 70 µmol) was added dropwise. The reaction mixture was stirred for 20 min at –5 °C and then water was added under cooling. The solution was diluted with EtOAc, and the organic layer was separated and washed with water and brine. The combined aqueous layers were extracted with EtOAc and the combined organic layers were dried over MgSO$_4$. Removal of the solvent and column chromatography (silica gel, pentane/EtOAc, 2:1, 1 vol % AcOH) of the residue provided **1** as a yellow solid; yield: 16.7 mg (99%); mp >240 °C (dec).

IR (ATR): 3413, 3393, 3298, 2972, 2868, 2030, 2010, 1658, 1639, 1586, 1464, 1258, 1146, 1116, 891, 866, 802, 718, 679 cm^{-1}.

^{1}H NMR (600 MHz, DMSO-*d*$_6$): δ = 1.49 (s, 6 H), 5.91 (d, *J* = 9.8 Hz, 1 H), 6.87 (dd, *J* = 8.6, 2.3 Hz, 1 H), 6.91 (d, *J* = 9.8 Hz, 1 H), 7.27 (d, *J* = 8.6 Hz, 1 H), 7.40 (dd, *J* = 2.3, 0.4 Hz, 1 H), 8.20 (d, *J* = 0.4 Hz, 1 H), 9.08 (br s, 1 H), 10.33 (s, 1 H), 11.46 (s, 1 H).

^{13}C NMR and DEPT (151 MHz, DMSO-*d*$_6$): δ = 27.24 (2 CH$_3$), 77.01 (C), 103.89 (C), 105.32 (CH), 111.59 (CH), 114.84 (CH), 116.84 (CH), 116.97 (C), 117.59 (C), 119.17 (CH), 124.00 (C), 129.70 (CH), 134.45 (C), 141.13 (C), 151.64 (C), 153.59 (C), 187.86 (CHO).

MS (EI): *m/z* (%) = 293 (13, [M$^+$]), 279 (19), 278 (100), 276 (23), 249 (6), 220 (9), 139 (6).

UV (MeOH): λ = 227, 288, 312, 372 (sh) nm.

Anal. Calcd for C$_{18}$H$_{15}$NO$_3$: C, 73.71; H, 5.15; N, 4.78. Found: C, 73.73; H, 5.29; N, 4.69.

X-ray Crystallographic Data for Compound 1

Single crystals of clausenalansine A (**1**) were obtained by crystallization from pentane/MeOH. C$_{18}$H$_{15}$NO$_3$, *M* = 293.31 g mol^{-1}, crystal size: 0.090 × 0.153 × 0.564 mm^3, triclinic, space group: P$\overline{1}$, *a* = 8.429(2) Å, *b* = 10.978(3) Å, *c* = 17.384(4) Å, α = 71.708(9)°, β = 78.951(9)°, γ = 74.748(10)°, *V* = 1463.0(6) Å^3, *Z* = 4, ρ$_{calcd}$ = 1.332 g cm^{-3}, µ = 0.091 mm^{-1}, λ = 0.71073 Å, *T* = 150(2) K, θ range = 2.49–28.00°, reflections collected: 71077, independent reflections: 7039 (*R*$_{int}$ = 0.0947), 6193 observed reflections [*I* ≥ 2σ(*I*)], 418 parameters. The structure was solved by direct methods and refined by full-matrix least squares on *F*^{2}; final *R* indices [*I* > 2σ(*I*)]: *R*$_1$ = 0.0500, *wR*$_2$ = 0.1341; max. residual electron density: 0.540 e Å^{-3}.[16]

Supporting Information

Supporting information for this article is available online at https://doi.org/10.1055/s-0040-1706551.

References

(1) (a) Chakraborty, D. P.; Roy, S. In *Progress in the Chemistry of Organic Natural Products, Vol. 57*; Herz, W.; Grisebach, H.; Kirby, G. W.; Steglich, W.; Tamm, C., Ed.; Springer-Verlag: Wien, **1991**, 71. (b) Chakraborty, D. P. In *The Alkaloids, Vol. 44*; Cordell, G. A., Ed.; Academic Press: New York, **1993**, 257.

(2) (a) Knölker, H.-J.; Reddy, K. R. *Chem. Rev.* **2002**, *102*, 4303. (b) Knölker, H.-J. *Top. Curr. Chem.* **2005**, *244*, 115. (c) Knölker, H.-J.; Reddy, K. R. In *The Alkaloids, Vol. 65*; Cordell, G. A., Ed.; Academic Press: Amsterdam, **2008**, 1. (d) Bauer, I.; Knölker, H.-J. *Top. Curr. Chem.* **2012**, *309*, 203. (e) Schmidt, A. W.; Reddy, K. R.; Knölker, H.-J. *Chem. Rev.* **2012**, *112*, 3193.

(3) Liu, Y.-P.; Guo, J.-M.; Liu, Y.-Y.; Hu, S.; Yan, G.; Qiang, L.; Fu, Y.-H. *J. Agric. Food Chem.* **2019**, *67*, 5764.

(4) (a) Narasimhan, N. S.; Paradkar, M. V.; Kelkar, S. L. *Indian J. Chem.* **1970**, *8*, 473. (b) Narasimhan, N. S.; Paradkar, M. V.; Chitguppi, V. P.; Kelkar, S. L. *Indian J. Chem.* **1975**, *13*, 993.

(5) Sriphana, U.; Thongsri, Y.; Prariyachatigul, C.; Pakawatchai, C.; Yenjai, C. *Arch. Pharmacal Res.* **2013**, *36*, 1078.

(6) (a) Wu, T. S. *Phytochemistry* **1991**, *30*, 1048. (b) Ito, C.; Thoyama, Y.; Omura, M.; Kajiura, I.; Furukawa, H. *Chem. Pharm. Bull.* **1993**, *41*, 2096. (c) Songue, J. L.; Kouam, E. D.; Mpondo, T. N.; White, R. L. *Molecules* **2012**, *17*, 13673.

(7) Schuster, C.; Rönnefahrt, M.; Julich-Gruner, K. K.; Jäger, A.; Schmidt, A. W.; Knölker, H.-J. *Synthesis* **2016**, *48*, 150.

(8) Hesse, R.; Kataeva, O.; Schmidt, A. W.; Knölker, H.-J. *Chem. Eur. J.* **2014**, *20*, 9504.

(9) (a) Julich-Gruner, K. K.; Kataeva, O.; Schmidt, A. W.; Knölker, H.-J. *Chem. Eur. J.* **2014**, *20*, 8536. (b) Schuster, C.; Julich-Gruner, K. K.; Schnitzler, H.; Hesse, R.; Jäger, A.; Schmidt, A. W.; Knölker, H.-J. *J. Org. Chem.* **2015**, *80*, 5666.

(10) Part 147 of Transition Metals in Organic Synthesis; for Part 146, see: Lösle, V.; Knölker, H.-J. *Arkivoc* **2020**, *vii*, 192.

(11) (a) Charles, M. D.; Schultz, P.; Buchwald, S. L. *Org. Lett.* **2005**, *7*, 3965. (b) Surry, D. S.; Buchwald, S. L. *Angew. Chem. Int. Ed.* **2008**, *47*, 6338. (c) Ruiz-Castillo, P.; Buchwald, S. L. *Chem. Rev.* **2016**, *116*, 12564.

(12) (a) Knölker, H.-J.; O'Sullivan, N. *Tetrahedron* **1994**, *50*, 10893. (b) Knölker, H.-J. *Chem. Lett.* **2009**, *38*, 8. (c) Gensch, T.; Rönnefahrt, M.; Czerwonka, R.; Jäger, A.; Kataeva, O.; Bauer, I.; Knölker, H.-J. *Chem. Eur. J.* **2012**, *18*, 770. (d) Gensch, T.; Richter, N.; Theumer, G.; Kataeva, O.; Knölker, H.-J. *Chem. Eur. J.* **2016**, *22*, 11186.

(13) Hesse, R.; Gruner, K. K.; Kataeva, O.; Schmidt, A. W.; Knölker, H.-J. *Chem. Eur. J.* **2013**, *19*, 14098.

(14) (a) Bandaranayake, W. M.; Begley, M. J.; Brown, B. O.; Clarke, D. G.; Crombie, L.; Whiting, D. A. *J. Chem. Soc., Perkin Trans. 1* **1974**, 998. (b) Sartori, G.; Casiraghi, G.; Bolzoni, L.; Casnati, G. *J. Org. Chem.* **1979**, *44*, 803.

(15) Gruner, K. K.; Hopfmann, T.; Matsumoto, K.; Jäger, A.; Katsuki, T.; Knölker, H.-J. *Org. Biomol. Chem.* **2011**, *9*, 2057.

(16) CCDC 2017103 (compound **1**) contains the supplementary crystallographic data for this paper. The data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/getstructures.