EFSUMB Recommendations and Guidelines for Gastrointestinal Ultrasound

Part 1: Examination Techniques and Normal Findings (Long version)

EFSUMB-Empfehlungen und Leitlinien des Gastrointestinalen Ultraschalls

Teil 1: Untersuchungstechniken und Normalbefund (Langversion)

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ABSTRACT
In October 2014 the European Federation of Societies for Ultrasound in Medicine and Biology formed a Gastrointestinal Ultrasound (GIUS) task force group to promote the use of GIUS in a clinical setting. One of the main objectives of the task force group was to develop clinical recommendations and guidelines for the use of GIUS under the auspices of EFSUMB. The first part, gives an overview of the examination techniques for GIUS recommended by experts in the field. It also presents the current evidence for the interpretation of normal sonoanatomical and physiological features as examined with different ultrasound modalities.
Introduction

Transabdominal gastrointestinal ultrasound (GIUS) offers the unique opportunity to examine non-invasively and in physiological condition the bowel including extra-intestinal features such as the splanchnic vessels, mesentry, omentum and lymph nodes. For properly trained users, GIUS has been shown to have good accuracy and repeatability not only in a primary work up, but also in the follow up of chronic diseases [1, 2].

Although there is an extensive documentation for the usefulness of GIUS in clinical practice it has only been fully implemented in some European countries and expert centres. Furthermore, the lack of standardization of the examination technique, and of guidelines, makes it hard to properly train physicians.

This was the motivation behind establishing the GIUS Task Force Group in 2014 under the umbrella of the European Federation of Societies for Ultrasound in Medicine and Biology (EFSUMB) which previously have published several guidelines and recommendations [3 – 11]. The group consists of a team of international experts of GIUS and the objective is to promote the use of GIUS in a clinical setting. This will be achieved by publishing clinical guideline series and recommendations on indications and use of GIUS for the gastrointestinal (GI) tract and by stimulating the development of training networks.

A guideline-series of altogether 7 papers are in the pipeline: examination techniques and normal findings, inflammatory bowel disease, transrectal and perineal ultrasound, other inflammatory disorders, functional disorders, upper GI ultrasound and miscellaneous pathologies.

In the making of this first document the GIUS task force group agreed on the scope of the document and then assigned a responsible author to select a panel of authors from the group based on their previous publications in the relevant fields of interest and their reputation as international experts in research and in teaching GIUS. Finally, a consensus meeting was held April 2016 to discuss important aspects of the guidelines and to vote on actual recommendations.

This document is mainly focused on presenting the examination techniques for performing GIUS and the normal ultrasound (US) features of the bowel, bowel wall and surrounding structures. Examination techniques and normal ultrasound findings for the perineal region and stomach are not included, but will be addressed in upcoming guideline papers. The recommendations are based on an extensive literature review. Based on the literature a recommendation level was suggested for each guideline. The Oxford Guidelines for reporting medical evidence was used specifying the level of evidence (LoE) and the grade of recommendation (GoR) [12]. Since many of the themes in these guidelines have not been subjected to systematic studies these recommendations often have a level of evidence 4 or 5, the latter simply being expert opinion. Therefore this document also includes the level of consensus of the members in the GIUS task force group. In April 2016 members of the Task Force Group participated in a consensus meeting in Gargnano, Italy. Each recommendation was discussed, adjusted and subjected to vote by members in the GIUS task force group. Recommendations 14 and 15 were not ready before the consensus meeting and were put to the vote during the review process. Degree of consensus was graded as follows: Strong consensus = > 95 %, broad consensus = 95 – 76 %, majority consensus 75 – 50 % and dissent < 50 %.

Equipment and examination modalities

B-mode

Ultrasound scanners should have sufficient quality and screen resolution to be able to delineate the structures in the gastrointestinal wall. The resolution of an US transducer is dependent on the frequency, the speed of sound in tissue and the number of cycles in the US pulse. Since the thickness of the bowel wall layers usually is less than 1 mm [13, 14], the frequency of a transducer must be at least 5 megahertz (MHz) for wall layers to be well discriminated [15 – 17]. No head-to-head studies have been published comparing the diagnostic performance of regular low frequency range abdominal probes (frequency range around 1 – 6 MHz), mid-frequency range transducers (frequency range around 5 – 10 MHz) and high frequency range transducers (frequency range around 10 – 18 MHz) for the detection of the intestines and intestinal disease. However, according to their specifications most mid-frequency range transducers offer the investigator a good compromise between resolution and depth penetration. While a mid-frequency range transducer can have a depth penetration of about 8 – 10 cm a high-frequency range transducer rarely penetrates beyond 4 cm. At the same time the resolution of a mid-frequency range transducer is quite adequate for separating individual layers in the GI wall [15 – 17]. A low-frequency range transducer is still needed for overview for reaching deeper lying bowel segments, such as the rectum and in obese patients. Harmonic imaging should be activated when available as this may improve the delineation of bowel wall layers [18, 19]. To document longer areas of involved intestines panoramic imaging may be helpful [20, 21].
Doppler techniques

Doppler US can assess both the signal from the visceral vessels that supply the gastrointestinal tract and directly smaller vessels of the intestinal wall, but cannot detect capillary flow.

Analysis of superior and inferior mesenteric inflow by pulsed Doppler scanning (systolic and diastolic velocities, resistance index, blood flow volume) provides several quantifiable parameters [22–25]. The best place to position the sample area is 2–3 cm distally to the origin of the vessel, in a longitudinal section as it runs parallel to the aorta, proximal to any side branches [26–28]. The examiner should tilt the probe to obtain an angle <60°. A high-pass filter of 100–200 kHz should be used to eliminate low frequencies related to vessel wall movement [28, 29].

Colour or Power Doppler can both be used to evaluate bowel wall vascularity [30]. Colour or Power Doppler flow parameters should be optimized to maximize the sensitivity for the detection of vessels with low-velocity flow in the bowel wall. Although specific technical characteristics depend on the equipment, in general it is recommended that persistence of colour be set at “medium,” the wall filter adjusted to the lowest setting, and a combination of the lowest velocity scale with the colour sensitivity at high level to maximize visualization of vessels avoiding colour blooming [30–34]. This special preset optimized for slow flow detection should be programmed, and be kept constant for the follow-up studies for each patient in cases of therapy monitoring [30, 31, 33]. Finally, colour Doppler gain should be turned up until flash artefacts occur and then turned down until they disappear before assessing vascularity.

The information obtained from colour Doppler images is semi-quantitative. It is recommended to measure bowel wall vascularity according to the number of vessels detected per square centimetre [30–33, 35]. According to previously published data, vascularity is subjectively assigned a grade as follows: grade 0 = no vessels; grade 1 or barely visible flow = fewer than two signals per square centimetre; grade 2 or moderate flow = three to five signals per square centimetre; and grade 3 or readily visible flow = more than five signals per square centimetre [30–33, 35].

Colour Doppler flow is considered present when colour pixels persist throughout the observation period and/or reoccur in the same location. Pulsed Doppler obtaining an arterial or venous signal at the location of the colour pixel should be used when there is doubt, to confirm that colour signals are originated from blood vessels and not from movement artefacts [31, 33, 36, 37].

If vascularity is not detected in the pathologically thickened intestinal wall this might be due to insensitivity of the equipment, inadequate chosen Doppler parameters, high body mass index or depth penetration >40 mm with loss of sensitivity.

**RECOMMENDATIONS:**

1. For a complete examination of the bowel both a low and high resolution probe are needed, LoE 5, GoR C, Strong consensus 13/13
2. A probe with a frequency above 5 MHz should be used when measuring wall thickness, LoE 4, GoR B, Strong consensus 13/13

Contrast-enhanced ultrasound

Contrast-enhanced ultrasound (CEUS) is performed after the injection of stabilized microbubbles with gaseous content into the blood stream.

The bubbles oscillate when subjected to US and the size and stiffness of the microbubbles affects the resonance frequency. Resonating microbubbles give rise to more intensely reflected signals which are easier to separate from tissue signals [38]. The most commonly used microbubble in Europe, Sonovue, is on average 2.5 μm and has a size distribution of 1–10 μm [39]. With high frequency probes a full vial of Sonovue (4.8 ml) is commonly used while it is usually sufficient with half a vial or less with low frequency probes. Due to the broad size distribution the higher dose will make more bubbles available for imaging at the higher frequencies needed for examining the intestinal wall [39].

There are several ways of interpreting contrast-enhancement in the bowel wall: pattern of enhancement [40–43], contrast quantification at peak intensity [44–46] and dynamic contrast-enhanced ultrasound where intensity changes over time are analysed [47–51].

CEUS can be used to quantify vascularity [44, 45, 52], but also be used to separate vascular from avascular tissue which is particularly useful when trying to differentiate a phlegmon from an abscess [53, 54].

Pattern of enhancement after a bolus injection is used as a qualitative parameter. For instance, patients with no enhancement can be separated from those with enhancement or the patients can be categorized according to where in the GI-wall the enhancement is detectable [42, 43, 55, 56]. The operators’ interpretation and the sensitivity of the US equipment may, however, affect the results [57].

Since there is a linear relationship between microbubble concentration and US intensity within a certain range [58] it is possible to quantify contrast-enhancement to make the method more objective. This does not reflect pathophysiological changes. Therefore attempts have been made to use an internal reference to reduce the variability [44, 45].

Finally, the contrast-enhancement can be analysed over time. This method is called dynamic contrast-enhanced ultrasound (DCE-US) and the values obtained from these analyses are closely related to perfusion in the GI wall. There are two main methods practiced, the bolus tracking and the burst-replenishment technique [48].

Bolus tracking is performed by injecting a dose of contrast followed by a flush of saline and analysing the development of the time intensity curve after the recording has been saved to the
Elastography

Elastography is a relative new technique that depicts the stiffness of tissues and is already used in clinical practice. An overview of the different techniques and applications has been published by EFSUMB [3, 5]. Recently, elastography has also been suggested as a tool for assessing diseases in the gastrointestinal tract [59, 60].

The bowel wall is thin, surrounded by serosa and with a lumen containing gas and chyme or fecal contents. This does not make it the ideal organ to be studied with elastography. However, pathology of the GI tract such as inflammation or tumour causes bowel wall thickening and often reduces motility and luminal contents in the affected area which may facilitate sonoelastography. There is good evidence for the use of elastography in endorectal ultrasound [61 – 64], but the evidence for transabdominal elastography of the bowel is sparse. Some recent studies suggest that it can be used to differentiate between fibrotic and inflammatory stenosis in Crohn's disease [65, 66].

RECOMMENDATIONS:

4. Contrast-enhanced ultrasound of the bowel can be used to separate vascular from avascular intestinal or peri-intestinal lesions including abscesses. EL 3b, GoR B, Strong consensus 12/12

5. Ultrasound elastography can be used to evaluate the stiffness of pathological thickened bowel. LoE 4, GoR C, Broad consensus 11/12

Investigator training and learning curve

It is important to set standards for performance of GIUS and for EFSUMB to secure high quality US education and professional standards. Previously, EFSUMB defined three levels of training recommendations in its release of minimal training requirements. Appendix 5 is specifically addressing gastroenterology [67]. EFSUMB recommends that GI US should mainly be performed by operators that have considerable experience and have passed the first competence level. However, also on level 1 the operator should be able to recognise the small and large bowel, and major focal intestinal abnormalities including obstruction. On level 2, the investigator should be able to perform a comprehensive examination of the GI tract: evaluation of the small bowel for focal or diffuse disease, the large bowel for the presence of diverticular disease and its complications (tumours and obstruction), the peritoneal cavity, its mesenteries, compartments and the omentum for the presence of infectious or malignant diseases. A level 3 practitioner should spend the majority of their time undertaking gastrointestinal US or teaching, research and development and be an expert in this area.

RECOMMENDATIONS:

6. Dedicated training in bowel ultrasound is necessary and should preferably be performed following training in general abdominal ultrasound, LoE 5, GoR C, Broad consensus 11/12

Preparation

In principle, no preparation of the patient is needed to perform a GIUS. Fluid installation, laxatives, and anti-flatulent preparations do not improve results [68, 69]. There is also no clinically relevant difference in wall thickness in the small and large bowel after a meal [14].

To reduce the amount of food and air in the small bowel a fasting period of at least 4 hours is recommended, however, fasting may not significantly improve visibility except in male patients [70, 71]. Also the presence of food in the stomach and small bowel will increase the flow in the splanchnic vessels which will vary with the size, composition and time since the last meal [72 – 76]. An overnight fast (> 8 hours) will include both the effect of improved visibility and minimize the effect of the previous meal.

Activity also affects splanchnic flow and thus the patients should refrain from extensive physical activity in the period before the examination [77].
Techniques

Scanning

The scanning technique for evaluating the bowel may vary according to the clinical problem [28, 78, 79]. The investigative approach will, for instance, differ between abdominal trauma, suspected intestinal obstruction [80] or appendicitis and chronic complaints such as longstanding diarrhoea. For surgical disorders a faster, targeted approach is used whereas for other complaints a full examination is performed. In this document, however, a general approach on how to perform the examination is described. There are no comparative studies where one GIUS scanning technique has been compared with another. As such these recommendations are mostly LoE meaning they are a matter of expert opinion.

After examining the parenchymal organs in the abdomen using the low frequency abdominal US probe the gastrointestinal tract is scanned systematically. First the abdominal US probe is used to get an overview before switching to a mid-range to high-frequency probe for a detailed examination.

The rectum can be scanned behind the urinary bladder with the abdominal US probe. The normal rectum may be difficult to display if the urinary bladder is empty.

The investigator should use a combination of internal and external references to describe the findings in the gastrointestinal tract. Since the cecum, ileocecal valve and terminal ileum very often are found and identified with certainty lying over the iliopsoas muscle in the right iliac fossa the bowel is followed in the distal direction through the ascending colon, right flexure, transverse colon, left flexure, descending colon and sigmoid colon and final the rectum. By sweeping back and forth in the transverse direction the examiner gets an overview of the pathology while at the same time tracing the path of the colon. The flexures are located high in the abdomen. The right flexure can be seen both intercostally and subcostally while the left flexure is found intercostally in the region of the spleen and left kidney.

If the examiner loses track of the colon the recommendation is to return to a known location and try again or identify a more distal area and backtrack. Segments that are easy to use as reference points are the ascending colon in the right flank, the descending colon in the left flank and the proximal sigmoid colon as it crosses the left iliopsoas muscle. The transverse colon can also be easily found by moving the probe from the epigastrium caudally, until typical haustations are recognized.

The small bowel scan starts by returning the probe to the right iliac fossa and identifying the terminal ileum. The examiner should then trace the terminal ileum as far as possible proximally. The rest of the small bowel is difficult to trace and to ensure most parts of the small bowel have been included in the examination a systematic scanning approach must be adopted. The abdomen should be scanned in parallel overlapping lanes cranially and caudally (“mowing the lawn”) while applying sufficient probe pressure so the dorsal wall of the abdominal cavity can be identified. This way the examiner is certain that all bowel segments between the probe and the dorsal wall are included in the scan. If the dorsal wall is not seen, such a claim cannot be made. In addition, scanning in a horizontal direction is recommended for a complete examination of all intestines. It is particularly important to look carefully at the small bowel segments in the pelvic region as it is harder to push away overlying bowel segments. A well filled urinary bladder may help in this regard as it will tend to push the small bowel loops in the hypogastric region up towards the umbilical region.

Graded compression

Graded compression is performed by using the US probe much in the same way as when performing palpation with the fingertips. The probe is used to compress the abdomen while following the respiratory movements. This can push away overlying bowel segments with gas or intraabdominal fat and in this way enable the examiner to reach deeper with high frequency probes such as for instance in the pelvis. The concept of graded compression was introduced by Puylaert [81] for the diagnosis of appendicitis [82 – 84]. Surgeons use the technique with good results [85]. Graded compression has been used for detection of bowel wall thickening [86] and for specific diagnoses such as diverticulitis [87, 88] and polyp detection [89].

Fluid use

Luminal gas and the variable and unpredictable presence of contents in the gastrointestinal tract may interfere with its visualization and with detailed evaluation of wall structure and intraluminal lesions. This can be improved by filling the lumen with an anechoic fluid. The ingestion of adequate amounts of water en-
hances the contrast and facilitates the assessment of the stomach wall if the gastric lumen has been properly distended [90]. The distention of the colonic lumen with instillation of water into the colon, hydrocolonic ultrasound [91], and with oral administration of hyperosmotic solutions [92] allows the visualisation of the colon with US from the rectosigmoid transition to the cecum in 97 % of patients studied, making the detailed examination of the architecture of colonic wall and surrounding structure possible. Unlike the stomach and the large bowel, water and osmotic solution are not appropriate to distend the lumen of the small bowel lumen. Water and hypo-osmolar solutions containing digestible or absorbable solutes are rapidly absorbed in the proximal small intestine, so that the lumen of the distal small bowel is not distended. Hyperosmolar solutions with indigestible contents, delay gastric emptying and, stimulating the intestinal peristaltic activity, hinder the appropriate lumen distension of the entire small bowel. It is thus unlikely that the entire small bowel can be visualized using hypo-osmolar, hyperosmolar, water or caloric fluid. The examination of the small bowel after ingestion of small (250 – 500 ml) amounts of iso-osmolar polyethylene glycol (PEG) 3350 – 4000 (macrogol) solution analogous to CT- or MR-enterography is called US-enterography or Small Intestine Contrast US (SICUS). With this technique the entire small bowel from the duodenal-jejunum angle to ileo-cecal valve can be visualized [93]. After ingestion the iso-osmolar and non-caloric macrogol solution is rapidly delivered from the stomach into the duodenum, since its gastric emptying is not opposed by osmotic and caloric-sensitive duodenal receptors [94]. The relatively constant gastric emptying rate induces gradual small bowel distension irrespective of the amount of the ingested solution. The PEG solution being non-digestible and non-absorbable links the waters molecules and thus retaining fluid within the lumen distends the intestinal wall. The luminal distension induces wall contractility and the PEG solution is displaced aborally, sequentially distending every single loop of the entire small bowel. SICUS used in healthy controls independent from the amounts of oral contrast used, results in values of wall thickness (≤ 3 mm) and lumen diameter (≤ 25 mm). These normative values help to discriminate normal from abnormal findings [93]. Safety and tolerability of PEG solution have been reported to be satisfactory previously, in studies using larger solution amounts than those administered in SICUS studies, and thereafter by several studies performed also in paediatric patients [95]. US enterocolysis has also been performed after instillation of PEG solution through a nasojejunal tube, placed in the duodenum using gastroscopy [96]. However, an excellent visualization of the small bowel was achieved only for the distal part of the ileum.

Hydrocolon examination with retrograde installation of fluids has also been used to improve visualization of colon pathology [91]. However, this technique has not gained widespread acceptance in clinical practice.

**RECOMMENDATION:**

11. Oral fluid contrast can improve visualisation of small bowel disease, LoE 1b, GoR A, Strong consensus 12/12

**Safety**

Diagnostic US should be performed according to the EFSUMB clinical safety statement [97]. Ultrasound is generally considered a very safe procedure and there are no data showing harmful effects of diagnostic US in adults. However, US may cause bio-effects with cavitation and tissue heating. The risk for causing such effect increases with the acoustic output (Pulse wave Doppler> Colour Doppler> B-mode), tissue transitions with large differences in acoustic impedance such as between soft tissue and bone and exposure time, but also between soft tissue and gas which is commonly encountered when examining bowel and exposure time. One should therefore limit examination time to what is necessary for diagnostic purposes [98].

Ultrasound contrast agents (UCA) have a low incidence of side effects. Serious anaphylactoid reactions occur in less than 0.002 % of the examinations [99, 100]. As they are excreted via the lungs and through breakdown in the liver they can be used in patients with kidney failure. When combining US with a high mechanical index and UCA’s, microvascular damage has been found resulting in small haemorrhages in animal models, but in these studies both higher MI and longer exposure times are used than in diagnostic US [10].

The benefit in using UCA’s should outweigh the risks. To avoid complications resuscitation facilities should be available, off-label use in areas where small haemorrhages may have serious clinical consequences should be avoided, long exposure and high mechanical indexes should be avoided and caution should be exercised when used in patients with severe coronary heart disease or pulmonary hypertension.

**Anatomy and sonographic findings**

**Bowel wall**

**Wall thickness**

In vitro measurements of GI wall thickness with high frequency US correlate well with histological sections [101]. However, studies have shown that devitalization of tissue and tissue preparation with formalin as well as histological sectioning can cause changes in tissue dimensions. Also differences in tissue texture and temperature can cause variability in the tissue impedance thus complicating the comparison between in vivo and in vitro measurements [17, 102].

There are several studies where wall thickness in different parts of the gastrointestinal tract has been measured with GIUS without a reference standard. In recent publications of studies performed with equipment comparable to present standards the common
finding is that both the normal small and large intestine is < 2 mm when distended [13, 14, 103–107]. The exceptions are the duodenal bulb and rectum which are smaller than 3 and 4 mm, respectively [14, 106]. Since collapsed bowel loops probably lead to higher wall thickness measurements it should be reported if the measurements were made on these.

The normal appendix can be identified in about 50% of healthy subjects using graded compression [108, 109], but experience plays a significant role. Maximum wall thickness in healthy volunteers is 2 ± 0.5 mm or less than 3 mm [110]. In clinical practice usually the maximum overall appendiceal diameter is measured, which should be less than 6 mm.

Wall layers

The gastrointestinal wall consists of 5 distinct sonographic layers when examined with a high frequency probe in the range of 5–15 MHz in vitro. The echo layers are a combination of interface echoes and the echo properties of the histological layers [101, 111, 112]. When imaged in the anterior wall of a bowel loop starting from the lumen the hyperechoic layer 1 corresponds to the interface between the mucosa and the lumen and is not a part of the actual GI wall. The hypoechoic layer 2 corresponds to the mucosa without the interface between the submucosa and mucosa, the hyperechoic layer 3 to the submucosa including this interface echo, the hypoechoic layer 4 to most of the proper muscle and layer 5 to the hyperechoic interface echo between the proper muscle and the serosa.

Since interface echoes are hyperechoic and located distally to the actual tissue interface, the correspondence between histology and sonographic layers differ slightly in the dorsal wall. Notably, the interface between lumen and mucosa (layer 1) is a part of the actual mucosa and layer 2 represents the rest of the mucosa without muscularis mucosae which normally is covered by an interface echo and add thickness to layer 3. Furthermore, the interface between submucosa and the proper muscle adds thickness to layer 3 and reduces the thickness of layer 4. The interface between the proper muscle and serosa (layer 5) extends beyond the actual serosa [15, 16, 113].

During in vivo scanning it is not always possible to discern all the layers. The interface echo from the serosa is mixed in with the interface from the peritoneum and the interface between the mucosa and the lumen can be difficult to distinguish without the presence of bowel air or luminal debris. Also the posterior bowel wall often is not possible to see due to air in the lumen. The measurements should therefore be made in the anterior wall. Since the interface from the serosa is difficult to delineate the measurement should be made from the start of the hypoechoic layer of the proper muscle to the end of the hyperechoic layer of the mucosa. Compression of the bowel wall with the transducer will reduce thickness and can make it difficult to separate the wall layers [114, 115]. However, some operators practice mild compression suggesting that this improves reproducibility of measurements [103]. The examiner should also be aware of interpretation difficulties due to mucosal folds and haustrations and keep the probe angled perpendicular to the GI wall to avoid tangential measurements.

RECOMMENDATIONS:
12. A bowel wall thickness less than 2 mm (not the cut-off value for pathology) could be considered as normal, when measured in the normal filling state except in the duodenal bulb and rectum, LoE 4, GoR B, Majority consensus 9/12

Superior and inferior mesenteric artery

The normal fasting flow in the superior mesenteric artery (SMA) has been assessed in a large number of studies where the healthy volunteers mostly have been added as a control group while there is clearly less data found on the flow parameters in the inferior mesenteric artery (IMA) [26].

The mean peak systolic velocity of the SMA varies between 93 to 146 cm/s in published literature, but there is considerable interindividual variability suggesting a normal range between 80 to 220 cm/s [116–122]. Resistive index ranges from 0.80 to 0.89 and blood flow from 380 to 640 ml/min in the SMA [23, 72, 116–118, 120–129]. Some of the variability could be caused by the difficult angle between the SMA and abdominal surface. In the IMA the blood flow is between 80 – 130 ml/min and the RI 0.9[24, 26, 130].

Intramural vessels

Vessel assessment in the GI wall is relevant with regards to diseases causing changes in vascularity such as for instance tumours, ischemia and inflammatory bowel disease. In vitro studies have shown that small vessels in the gastrointestinal wall can be identified using high frequency US [131]. More common is the use of colour Doppler to detect flow in the vessels of the GI wall. Due to the comparatively slow flow and small dimensions of these vessels the velocity range of the colour Doppler has to be set very low between 2 to 5 cm/s [31, 36, 122, 132, 133]. This increases the risk of flash artefacts and the patients need to hold their breath during the acquisition. Also, due to the PRF needed to perform this exam-
in the depth where this flow can be detected is quite limited. Colour and power Doppler provide a semi-quantitative description of vessel density in the bowel wall. In the healthy bowel wall it is uncommon to detect more than one or two vessel signals with colour or power Doppler [36, 122].

Pulse wave measurements of individual arteries in the GI wall can provide indirect quantitative measurements of the local vascularity. Since the angle of a vessel in the GI wall is difficult to see the resistive index is used [36, 122, 134]. Unfortunately, the measurements are difficult to perform and the technique is not commonly used in clinical practice.

Local perfusion

DCE-US provides non-invasive measurements of the perfusion in the gastrointestinal wall. To date only one study reports absolute blood flow values from healthy volunteers with a median and range of 44.5 (6.6 – 91.2) ml/min/100 ml tissue and 39.4 (2.2 – 111.4) ml/min/100 ml of tissue[51]. The method requires much post-processing and has a substantial variability which is currently not useful in clinical practice, but seems in line with current literature [135 – 139].

Small and large bowel

Location

The duodenum passes into the small bowel at the ligament of Treitz. The small bowel has a tortuous course and is very moveable due to the mesenteric leaves. The jejunum is usually located in the left upper- and mid-abdomen, and the ileum in the right mid- and lower abdomen. The right iliac vessels are a landmark of the ileocecal region. As a result of malrotations the different parts of the small bowel can also be found in other positions.

The colon is located like a picture frame more in the periphery of the abdomen. The ascending and descending colon are usually fixed to the retroperitoneum dorsolaterally on the right and on the left side, respectively. The transverse and the sigmoid colon may have a more variable course owing to the different length of the mesocolon [140]. The transverse colon may descend down to the lower abdomen in case of an elongated mesocolon or may be located behind the stomach in case of a very short mesocolon [141]. The sigmoid colon on the other hand may have an elongated course and can cross the midline to the right iliac fossa or even extend up to the liver. The rectum is visible in its predominantly extraperitoneal position behind the urinary bladder.

Sometimes the cecum and ascending colon may be located intraperitoneal with the cecum in variable positions or the whole colon is located on the left side of the abdomen. This is important for diagnostic US because of possible misinterpretations of pathologic findings and because of allocation of findings to the wrong bowel segment. Such variations are better detected with CT or MRI than with US.

The appendix arises from the cecum about 3 cm below the ileocecal valve at the point where the three taeniae converge. It has a highly variable position such as the typical medial course over the iliopsoas muscle, a medial or lateral elevation or a retro-caecal course and it also varies with the position of the cecum [142].

Appearance

The small bowel has a length of 3 – 6 metres and is characterised by the valvulae conniventes. They decrease in number and height from the proximal jejunum to the distal ileum and are best visualised when the bowel loops are fluid-filled. In a collapsed condition bowel loops may have a predominant hypoechoic appearance or in case of intraluminal gas a hyperechoic appearance. Usually we can find both conditions side by side. Usually only high-frequency transducers allow the visualisation of the valvulae conniventes of collapsed loops.

The colon is characterised by its hausturation, which is best visible on US in longitudinal sections if the colon is filled with stool and gas and thus has a hyperechoic appearance. In a contracted condition – which is more frequently seen at the left hemicolon – the hausturation is not adequately demonstrable. The semilunar folds protrude to the lumen between the haustra and are only visible after cleansing preparation of the colon which allows the best visualisation of the colonic wall [143]. If the colon is distended and filled with stool, bowel wall layers are hardly visible even with high-frequency transducers. When we look for the colon with the abdominal probe, we are usually guided by the typical location and by the hyperechoic luminal content and not by the aspect of the colonic wall itself. The numerous epiploic appendages of the colon can only be differentiated from adjacent fatty tissue if fluid is present in the peritoneal cavity.

When examined with a high-frequency probe, the appendix usually appears as a target structure with different wall layers [144]. If the lumen is completely filled with gas, a predominant hyperechoic appearance may result. Sometimes this can be helpful to find a normal appendix even with the abdominal probe if higher frequencies cannot be applied.

Motility

The normal transit time for the small bowel ranges from 2 – 6 hours [145]. Knowledge on motility of the small bowel motility is still limited due to complex interaction between the central and enteric nervous system, sensory and motor functions and multiple gastrointestinal hormones influencing peristaltic activity [143].

After overnight fasting the motility of the small bowel is reduced [146, 147], but intake of food or fluids will induce contractions. To-and-fro movements in the bowel improves the contact between contents and the mucosa for absorption of nutritional components and is significantly more easily seen in patients with coeliac disease [143].

Even during transit of colonic contents such a to-and-fro movement is present [148]. But this is usually not noticeable on US because of the long transit time in the colon (20 – 72 hours) with very slow peristaltic movement. It is usually only under pathological conditions such as enterocolitis or bowel obstruction that contractions in the colon are visible on US. The peristalsis of the appendix is also not noticeable during examination.
Blood supply

The whole small bowel is supplied by the SMA with its jejunal and ileal branches. The blood supply of the colon occurs on the one hand via branches of the SMA and on the other hand via the IMA. The watershed between the SMA and IMA is in the transverse colon near the splenic flexure. The rectum has its arterial supply from the IMA and the internal iliac artery. This explains the typical affection of the colon from the left colonic flexure to the sigmoid colon in ischaemic colitis.

The superior mesenteric vein accompanies the superior mesenteric artery and the inferior mesenteric vein runs vertically upward and enters the splenic vein or its junction with the superior mesenteric vein to form the portal vein.

Collateral pathways are important to protect the bowel wall from potential ischaemia if arterial supply is compromised. In case of severe stenosis or occlusion at the origin, the one pathway connects the three mesenteric vessels. The other collateral pathway is formed by multiple interconnecting arterial arcades between the branches in order to warrant adequate blood supply in cases of segmental arterial occlusion [149].

Lumen

After overnight fasting, the lumen of the small bowel is frequently collapsed. Usually only small amounts of intraluminal fluid and some gas are present. Depending on nutritional components a more or less hyperechoic liquid content and more gas is visible after a meal. Small bowel obstruction and oral intake of fluids or application through a feeding tube result in hypoechoic luminal content. The normal maximum diameter of small bowel loops ranges from 2 – 2.5 cm [147, 150, 151].

At the level of the ileocecal valve, where the ileal content passes over to the colon, a still liquid content of mixed echogenicity may be visible. The faecal material gradually solidifies as it moves along in the colon and thus becomes hyperechoic. The diameter of the colon usually measures up to 5 cm, whereas that of the cecum may exceed this width [80, 152]. The width of the left hemicolon slightly decreases in an aboral direction. The colon is usually filled with stool and gas but the descending and sigmoid colon sometimes present in a mainly contracted condition which could make detection of these bowel segments more difficult.

The lumen of the normal appendix may be collapsed or contain some stool and gas. The lumen rarely exceeds a width of 4 – 5 mm. At times we can see that the lumen of a distal segment is obliterated and the hyperechoic submucosa is the predominant layer[142].

Peri-intestinal features

Peri-intestinal sonographic findings provide relevant elements, as an adjunct to the features of bowel wall to suspect, diagnose or exclude digestive diseases. Therefore, mesentery and lymph nodes should always be assessed during routine bowel investigations.

Mesentery and omentum

Mesentery extends laterally to the aorta, from the left hypochondrium to right iliac fossa. It is scanned with both regular abdominal and mid-range to high-frequency probes, depending on size of the patient, as visceral fat determines increase in attenuation thus limiting the use of high-frequency probes [153]. The normal mesentery appears at US as a series of mildly hyperechoic parallel layers, 7 – 12 mm in thickness, alternated by hyperechoic strips, resembling thickened bowel walls in a longitudinal scan. Mesentery is easily seen when ascites is present, appearing as a series of hyperechoic folds, which arise from the posterior wall of the peritoneal cavity and extend to the bowel loops, visible at their extremities.

Mesentery may be affected by several systemic and gastrointestinal diseases. As it reflects the overall visceral adiposity, increased mesenteric fat thickness (> 1 cm) may correlate with metabolic syndrome and cardiovascular diseases [154]. More important, chronic and acute inflammatory disorders (e.g. Crohn’s disease, appendicitis and diverticulitis) and some neoplastic diseases affecting the bowel may show mesenteric hypertrophy, also named fat wrapping or creeping fat presenting as a firm, abundant hyperechoic tissue, surrounding the bowel loops [155 – 159].

Despite the accuracy of US in the description and detection of mesenteric abnormalities, it is limited by inferior panoramic view compared to CT and MRI.

Lymph nodes

The detection of enlarged or even normal mesenteric lymph nodes is a common and often incidental finding of abdominal and bowel US, in particular in children and young adults [160]. The sonographic detection of regional mesenteric lymph nodes may be a normal or physiologic condition or suggest a past or ongoing, mainly inflammatory or neoplastic, disease of the abdomen.

In adults normal mesenteric lymph nodes appear as oval, elongated or U-shaped hypo- or mild hypo-echoic nodules with the shorter diameter < 4 mm and larger diameter usually < 17 mm [161 – 164]. In children, due to an activated immune response and as a result of previous intestinal infections, normal mesenteric lymph nodes may have a shorter axis with a diameter up to 10 mm, but preserved regular shape ad echogenicity [160, 165, 166].

In enlarged mesenteric nodes, the size, number, site, shape and echogenicity are not specific for the underlying diseases [167]. However, the analysis of all these features may help in discriminating between infectious, inflammatory or potential neoplastic causes [168]. Enlarged mesenteric lymph nodes may sug-
gest intestinal and systemic inflammatory conditions as well as neoplastic diseases. In particular if associated with sonographic changes of the bowel and mesenteric hypertrophy [161, 169 – 172].

**RECOMMENDATION:**
18. Ultrasound can assess lymph nodes and mesenteric tissue. LoE 4, GoR B, 4, Strong consensus 10/10

**Clinical applications**

Intestinal US is often suggested as the first imaging tool in patients with acute abdomen [88]. Systematic reviews and meta-analyses have shown that US is highly accurate in detecting acute appendicitis, although not as high as CT [174, 175]. However, as their positive predictive value is quite similar, US can be used as the first imaging tool in a conditioned US-CT strategy where patients with US positive for appendicitis, are sent directly to surgery, avoiding CT, while those with inconclusive or negative sonographic results are submitted to CT. This strategy has been proven to be as effective as immediate CT for all patients, and although potentially burdened by more false positive results, it halves the number of CTs needed, without any impact on length of hospital stay, saving radiation exposures and costs [176 – 178].

The diagnosis of acute colonic diverticulitis can be made in patients only by clinical evaluation [179]. However, additional imaging is usually required to establish the diagnosis and assess complications. Systematic reviews and meta-analyses have shown that US and CT have high and comparable accuracy in diagnosing acute diverticulitis [88, 180]. Despite the advantage of CT due to higher specificity, panoramic view and the ability to identify alternative diagnoses, a conditional strategy with CT performed after an inconclusive or negative US, is the preferable approach, endorsed also by national guidelines [179, 181].

Intestinal US accurately detects ileus, showing as dilated (> 3 cm) and fluid-filled small bowel loops. Real-time US evaluation enables also to assess the nature of ileus, if mechanic or dynamic, and may suggest the causes and severity. In particular, the reported sensitivities and specificities of US in detecting ileus is high in most prospective studies published so far both in consecutive series of patients and in selected population of Crohn’s disease patients [182 – 184].

Besides acute conditions, one of the most common uses of intestinal US is the detection and follow-up of inflammatory bowel diseases, in particular Crohn’s disease along with disease complications such as strictures, fistulas, abscesses and extra-intestinal complications. Several systematic reviews and meta-analyses have shown that US is able to detect signs of Crohn’s disease and, like CT and MRI, has a high and comparable diagnostic accuracy at the initial presentation of terminal ileal CD, as well as in monitoring the disease by assessing its activity and abdominal complications [1, 2]. US has proven to be of value in the follow up of IBD patients irrespective of symptoms [185].

Finally, when used as preliminary imaging investigation in patients with abdominal symptoms, such as abdominal pain or changes in bowel habits, US can identify abnormal intestinal findings or lesions that suggest intestinal diseases which may not primarily have been suspected [186]. In particular, US can detect signs suggesting malabsorption and celiac disease such as enlarged mesenteric lymph nodes, dilated small bowel loops with increased fluid content, and increased peristalsis with high sensitivity. The overall accuracy and the place of US in the diagnostic algorithm of celiac disease may vary upon the probability of the disease in the considered population [169, 187]. The detection of these signs in patients with abdominal complaints and changes in bowel habit can adequately drive further investigations.
Finally, intestinal US can detect masses and neoplastic lesions of the gastrointestinal tract, in particular when in advanced stage[188]. In contrast, the role of US in detecting or suggesting gastrointestinal functional disorders is not established and needs further investigation.

References

Guidelines & Recommendations


Liu KH, Chan YL, Chan WB et al. Mesenteric fat thickness is an independent determinant of metabolic syndrome and identifies subjects with increased carotid intima-media thickness. Diabetes Care 2006; 29: 379 – 384


