Markers of Hereditary Thrombophilia with Unclear Significance

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Abstract

Thrombophilia leads to an increased risk of venous thromboembolism. Widely accepted risk factors for thrombophilia comprise deficiencies of protein C, protein S, and antithrombin, as well as the factor V "Leiden" mutation, the prothrombin G20210A mutation, dysfibrinogenemia, and, albeit less conclusive, increased levels of factor VIII. Besides these established markers of thrombophilia, risk factors of unclear significance have been described in the literature. These inherited risk factors include deficiencies or loss-of-activity of the activity of ADAMTS13, heparin cofactor II, plasminogen, tissue factor pathway inhibitor (TFPI), thrombomodulin, protein Z (PZ), as well as PZ-dependent protease inhibitor. On the other hand, thrombophilia has been linked to the gain-of-activity, or elevated levels, of α 2-antiplasmin, angiotensin-converting enzyme, coagulation factors IX (FIX) and XI (FXI), fibrinogen, homocysteine, lipoprotein(a), plasminogen activator inhibitor-1 (PAI-1), and thrombinactivatable fibrinolysis inhibitor (TAFI). With respect to the molecular interactions that may influence the thrombotic risk, more complex mechanisms have been described for endothelial protein C receptor (EPCR) and factor XIII (FXIII) Val34Leu. With focus on the risk for venous thrombosis, the present review aims to give an overview on the current knowledge on the significance of the aforementioned markers for thrombophilia screening. According to the current knowledge, there appears to be weak evidence for a potential impact of EPCR, FIX, FXI, FXIII Val34Leu, fibrinogen, homocysteine, PAI-1, PZ, TAFI, and TFPI on the thrombotic risk.

Keywords

- thrombophilia
- risk factors
- unclear evidence

Zusammenfassung

Eine Thrombophilie führt zu einem erhöhten Risiko für venöse thromboembolische Ereignisse. Weithin anerkannte Risikofaktoren für eine Thrombophilie sind ein Mangel an Protein C, Protein S und Antithrombin, sowie die Faktor-V-"Leiden"-Mutation, die Prothrombin-G20210A-Mutation, eine Dysfibrinogenämie und, weniger eindeutig, erhöhte Faktor VIII-Aktivitäten. Neben diesen etablierten Markern sind in der Literatur auch Risikofaktoren von eher unklarem Stellenwert beschrieben. Zu diesen Risikofaktoren, bzw. Markern, gehören der Funktionsverlust (loss-of-activity), bzw. der Mangel, der Aktivität von ADAMTS13, Heparin-Cofaktor II (HCII), Plasminogen, Tissue Factor Pathway Inhibitor (TFPI), Thrombomodulin (TM), Protein Z (PZ) sowie des

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Protein Z-abhängigen Proteaseinhibitors (ZPI). Andererseits wurde eine Thrombophilie mit einem qain-of-activity, bzw. erhöhten Aktivitäten, von α2-Antiplasmin (a2-AP), Angiotensin-konvertierendem Enzym (ACE), den Gerinnungsfaktoren IX (FIX) und XI (FXI), Fibrinogen, Homocystein, Lipoprotein(a), Plasminogenaktivator-Inhibitor-1 (PAI-1) sowie dem Thrombin-aktivierbaren Fibrinolyse-Inhibitor (TAFI) in Verbindung gebracht. Im Hinblick auf die zugrundeliegenden molekularen Interaktionen, die einen Einfluss auf das Thromboserisiko haben könnten, stellen sich die Mechanismen für den endothelialen Protein-C-Rezeptor (EPCR) und den Faktor-XIII-(FXIII) Val34Leu komplexer dar. In der vorliegenden Arbeit soll ein Überblick über den aktuellen Wissensstand zur Bedeutung dieser Marker für das Thrombophilie-Screening bei Patienten mit venösen thromboembolischen Ereignissen gegeben werden. Hierbei zeigt sich nach derzeitigem Kenntnisstand eine schwache Evidenz für einen möglichen Einfluss von EPCR, FIX, F XI, F XIII Val34Leu, Fibrinogen, Homocystein, PAI-1, PZ, TAFI und TFPI auf das Thromboserisiko.

Introduction

Thrombophilia refers to abnormal blood coagulation, leading to an increased risk of thromboembolism. Originally restricted to risk factors associated with venous thromboembolism (VTE), the concept of thrombophilia is now increasingly extended to thrombotic and embolic arterial events such as peripheral artery disease¹ or stroke.² In this narrative review, we will focus on thrombophilia leading to an increased risk of VTE. VTE is a common and serious blood clotting condition that includes both deep-vein thrombosis (DVT) and pulmonary embolism. DVT forms in deep veins, usually those of the legs. If the thrombus breaks free and becomes lodged to the lung arteries, DVT develops into pulmonary embolism. Thrombi in other parts of the body, such as in abdominal veins, are often excluded from clinical studies, and data on thrombophilia markers may not be directly related to these "atypical" events.

Thrombophilia can be either inherited or acquired. The two central mechanisms in inherited thrombophilia are the loss of activity of physiological anticoagulants and the gain of activity of physiological procoagulants. Widely accepted loss-of-activity thrombophilia markers include the deficiencies of protein C, protein S, and antithrombin while accepted gain-of-activity markers include the factor V "Leiden" (FVL) mutation R506Q, the prothrombin G20210A mutation, certain types of dysfibrinogenemia, and, to some extent, increased levels of factor VIII (FVIII; Table 1).³⁻⁵

Besides these classical and well-established markers of thrombophilia, a huge number of additional risk factors of unclear significance have been proposed and discussed in the literature. In the present review, based on the functions of the molecules, we aim to give an overview on the current knowledge of the significance of these markers for thrombophilia screening. We assume that a thrombophilia marker is unlikely to have clinical relevance if related to an odds ratio (OR) of less than 2.0 and that such a marker has no additional value in clinical decision making.6

The Fibrinolytic System

The fibrinolytic system plays a pivotal role in the regulation of hemostasis; because many other functions are associated with its active components, the fibrinolytic system is also referred to as the "plasminogen-plasmin system." Plasminogen is activated to plasmin by urokinase-type plasminogen activator or tissue-type plasminogen activator in humans.⁸ Plasmin is a serine protease that cleaves fibrin to break down blood clots but also cleaves fibronectin, thrombospondin, laminin, and von Willebrand factor (vWF). 9,10 The plasminogen activators are modulated by their inhibitor, called plasminogen activator inhibitor (PAI), of which three forms have been described: PAI-1, PAI-2, and PAI-3, as well as by nexin.¹¹ The active enzyme plasmin is modulated by its inhibitors α 2-antiplasmin and α 2-macroglobulin. ¹² A third modulator is thrombin-activatable fibrinolysis inhibitor (TAFI), which can be activated by thrombin and inhibits clot lysis by removing carboxy-terminal lysine residues from fibrin, which subsequently prevents plasminogen from binding to fibrin.¹³ A decreased fibrinolytic potential has been demonstrated to be a risk factor for venous and arterial thrombosis.¹⁴ Low levels of plasminogen and high levels of inhibitors (PAI-1, TAFI, and a2-antiplasmin) are likely to contribute to this association. In fact, variation on clot lysis time could be explained by variations of these four molecules in more than 75% of patients investigated for clot lysis. 15 In the same study, elevated levels of PAI-1 and TAFI were associated with an increased risk of thrombosis, whereas the association for plasminogen and t-PA was lost after adjusting for markers of inflammation.

Plasminogen

Both quantitative and qualitative defects of plasminogen have been described. Although homozygous deficiency of plasminogen results in failure of the body to remove fibrin deposits in various organs, leading to pseudomembranous conjunctivitis, obstructive airway disorders, and other sequelae, there is no evidence for an increased risk of

Table 1 Current evidence for an association between markers of hereditary thrombophilia with unclear significance and VTE

Thrombophilia	Accepted markers (approx. relative risk)	Markers of unclear significance	
		Name	Current evidence for association with VTE
Loss-of-activity	AT deficiency (40–50) Protein C deficiency (15) Protein S deficiency (10)	ADAMTS13 α2-Antiplasmin EPCR mutations Heparin cofactor II MTHFR A1298C MTHFR C677T Plasminogen Protein Z TFPI TM mutations ZPI	No No Rather yes No No No No Rather yes Rather yes Ro No No
Gain-of-activity	Factor V Leiden (heterozygous, 7; homozygous, 11–80) Prothrombin G20210A (heterozygous, 3–4; homozygous, 7–30) High FVIII (4)	ACE Clotting factor IX Clotting factor XI Clotting factor XIII Val34Leu Fibrinogen Homocysteine Lp(a) PAI-1 4G/5G Polymorphism TAFI	No Rather yes Rather yes Rather yes ^a Rather yes Yes No ^b Rather yes

Abbreviations: ACE, angiotensin-converting enzyme; EPCR, endothelial protein C receptor; PAI-1, plasminogen activator inhibitor-1; TAFI, thrombin-activatable fibrinolysis inhibitor; TM, thrombomodulin; VTE, venous thromboembolism; ZPI, protein Z-dependent protease inhibitor. Notes: Risk markers are divided according to loss- or gain-of-activity of underlying proteins/enzymes and presented in alphabetical order. For widely accepted markers, the approximate relative risk for a first VTE is provided (see text for references). "No" indicates that available meta-analyses or the majority of published studies found no significant association. "Yes" indicates that available meta-analyses or the majority of published studies demonstrated an association with a statistically significant OR \geq 2.0, that is, an at least twofold increased chance for VTE. "Rather yes" indicates that meta-analyses or the majority of studies demonstrated a statistically significant OR \geq 1.0, but <2, that is, a chance that is only discreetly increased. "This is a protective polymorphism.

thrombosis, neither for individuals with homozygous nor for individuals with heterozygous plasminogen deficiency. ^{16,17} In addition, various qualitative defects of plasminogen were identified in patients with thromboembolic events, but the majority of family members, though affected, were not symptomatic. ¹⁸ In summary, there is no evidence that plasminogen deficiency contributes to thrombophilia.

Plasminogen Activator Inhibitor 1

PAIs interfere with plasminogen activation, and elevated levels of PAIs should predispose to thromboembolism. At least three polymorphic variations in the human PAI-1 gene SERPIN1 have been reported. Specific alleles were associated with altered levels of PAI-1 in plasma. The biological consequences of elevated PAI-1 levels are not fully clear. The deletion/insertion (4G/5G) polymorphism in the promoter region leading to elevated levels of circulating PAI-1 has been studied extensively. 19 In a meta-analysis, the 4G allele was associated with unexplained VTE with an OR of 1.153 (95% confidence interval [CI], 1.068-1.246).²⁰ A slightly increased arterial thrombotic risk (OR, 1.088; 95% CI, 1.007-1.175) was also reported,²¹ but an association of this polymorphism with stroke could not be established.²² In the end, there is preliminary evidence that the PAI-1 4G/5G polymorphism may be associated with a slightly increased risk of thromboembolism under yet-to-be-defined circumstances.

α2-Antiplasmin

There is no evidence for an association between elevated $\alpha 2\text{-antiplasmin}$ levels and the risk of thrombosis. Two smaller studies on thrombotic events postsurgery did not show an association, 23,24 and elevated $\alpha 2\text{-antiplasmin}$ levels were also not associated with increased risk of thrombosis in a study of fibrinolytic markers. 15 These findings are in accordance with the assumption that $\alpha 2\text{-antiplasmin}$, because it is present in abundance, cannot represent a limiting factor of fibrinolysis, while rather regulating steps before the final activation of plasminogen are more likely to be critical. 25

Thrombin-Activatable Fibrinolysis Inhibitor

TAFI inhibits fibrinolysis when converted to its active form by thrombin (or plasmin).²⁶ Elevated levels of TAFI should therefore reduce clot lysis and lead to an elevated risk of thromboembolism. In the Leiden Thrombophilia Study (LETS), TAFI levels above the 90th percentile of the controls were associated with a slightly increased risk for thrombosis (OR, 1.7; 95% CI, 1.1–2.5), and adjustments for various possible confounders did not substantially affect this estimate.²⁷ Elevated levels of TAFI were also reported as a mild risk factor for recurrent VTE, even after adjustment for potentially confounding variables (risk ratio, 1.06; 95% CI, 1.0–1.16).²⁸

^bRetinal vein occlusions are an exception and an association has been established for this specific patient group.

A comparable association between elevated TAFI levels was also reported for patients from the MEGA study in multiple assays of fibrinolytic markers (OR, 1.6; 95% CI, 1.2–2.1 for the highest quartile). 15 In contrast, a retrospective family study on 1,940 patient relatives identified 187 individuals with high TAFI levels, but the adjusted relative risk did not show an association with thromboembolic events (relative risk, 0.8; 95% CI, 0.5-1.3).²⁹ In summary, a minor effect of elevated TAFI levels on the risk of thrombosis cannot be excluded.

Lipoprotein(a)

Lipoprotein(a) (Lp(a)) is not part of the fibrinolytic system, but this cholesterol-rich lipoprotein consists of a low-density lipoprotein domain attached to apolipoprotein(a), which has partial homology to plasminogen. Lp(a) and plasminogen compete for fibrin binding, which in theory should impair fibrinolysis whenever Lp(a) is available in higher-than-normal concentration. Recent in vitro evidence, however, questions the relevance of this mechanism.³⁰ Without doubt, Lp (a) is an independent risk factor for atherosclerotic cardiovascular diseases through mechanisms associated with increased atherogenesis and thrombosis, but its contribution to VTE is less clear.31 In a systematic review and metaanalysis based on case-control studies, a significant, but modest association between Lp(a) and VTE has been reported (OR, 1.56; 95% CI, 1.36-1.79).32 In contrast, in a pooled analysis of population-based cohort studies, the fully adjusted hazard ratio for VTE was nonsignificant (hazard ratio, 1.00; 95% CI, 0.94-1.07).33 In addition, genetic association studies were also unable to demonstrate an association between defined genetic variants which lead to elevated Lp (a) and the risk VTE. ^{34,35} Despite this, a recent meta-analysis of studies that aimed to lower Lp(a) concentrations using evolocumab, a monoclonal antibody blocking the PCSK9 enzyme, demonstrated a significant VTE risk reduction in patients whose baseline Lp(a) was above the median (hazard ratio, 0.52; 95% CI, 0.30-0.89), whereas evolocumab had no such effect in the low Lp(a) group. 36 This does not prove an effect of Lp(a) itself; additional metabolic changes following PCSK9-blockage may be responsible for the observed effects.

Unlike in other VTEs, thrombophilia does not seem to play a major role in retinal vein occlusion (RVO),³⁷ while common cardiovascular risk factors, such as hypertension, diabetes, and hyperlipemia, were reported to be predisposing factors.³⁸ RVO is therefore often considered a complication of atherosclerosis rather than a form of venous thrombosis. According to a recent meta-analysis, elevated Lp(a) also seems to have effects on an unusual thrombotic event (OR, 2.38; 95% CI, 1.7-3.34).³⁹ Although we do not think that Lp(a) is a relevant thrombophilia marker for VTE in general, we recommend including Lp (a) in the thrombophilia workup for RVOs.

Elevated Levels of Coagulation Factors

An elevated activity level of FVIII is a widely accepted thrombophilia marker. It is well documented through several studies that the thrombotic risk is approximately three- to fivefold increased for FVIII levels ≥150 IU/dL.⁴⁰ In addition, the impact of elevated FVIII levels on the risk of recurrent thrombosis is also well documented.⁴¹ Only recently, in a large population-based case-control study, elevated FVIII levels were associated with a high risk of VTE after orthopaedic surgery (OR, 18.6; 95% CI, 7.4-46.9), comparable to the risk associated with the presence of FVL (OR, 17.5, 95% CI, 4.1–73.6).⁴² Despite solid evidence for an association between elevated FVIII levels and VTE, it is still a matter of debate whether or not FVIII should be included into thrombophilia screening. The widespread use of one-stage clotting assays with their analytical shortcomings (mainly overestimating FVIII levels), the large intra-individual variation, the potential influence of a previous thrombotic event, any acute phase reaction, any underlying disease such as cancer, and the lack of a defined cut-off value make the use of FVIII as a thrombophilia factor problematic. It has been demonstrated that a shortened activated partial thromboplastin time (aPTT) is a risk factor for thrombosis and that this association between a short aPTT and VTE is only partially mediated by high FVIII levels. 43 Accordingly, it could be worth to look at other factors that affect aPTT, especially FXI, FIX, and fibrinogen.

Factor XI

The LETS demonstrated an OR of 2.2 (95% CI, 1.5-3.2) for VTE for individuals with a FXI level above the 90th percentile.⁴⁴ In a subsequent nested case-control study where the authors looked at levels of factors IX through XIII in 462 cases and 1,047 controls, only elevated levels of factors IX and XI were associated with an increased risk of VTE after adjustment for age, sex, ethnicity, and study. 45 With further adjustment for body mass index and diabetes, only elevated FXI levels remained associated with an elevated VTE risk: OR, 1.8 (95% CI, 1.3-2.7). Finally, a prospective study where 16,299 participants, initially free of VTE, were followed up over time, 606 VTEs occurred; the age, ethnicity, sex, and study-adjusted hazard ratio was 1.51 (95% CI, 1.16-1.97) for the highest versus lowest quintile of FXI, indicating a mild association.⁴⁶ Only few studies on plasmatic FXI levels and the risk of VTE did not find an association.⁴⁷ In contrast, studies that focused on polymorphisms in the F11 gene gave contradictory results. Regarding F11 rs2289252, a Swedish case-control study found an association (hazard ratio, 1.8; 95% CI, 1.1-3.0), whereas, using the same SNP, no association was identified in a Portuguese study (OR, 1.09; 95% CI, 0.75-1.59).^{48,49} We conclude that elevated FXI levels are most likely a mild risk factor for VTE.

Factor IX

The LETS identified elevated levels of FIX as a risk factor for VTE.⁵⁰ Using the 90th percentile measured in control subjects (P(90) = 129 U/dL) as the cut-off, a two- to threefold increased risk for individuals with elevated FIX levels was identified. Adjustment for age, sex, oral contraceptive use, and high levels of factors VIII and XI did not affect this association. Even after exclusion of individuals with known genetic disorders, the OR for elevated FIX levels was still 2.5 (95% CI, 1.6-3.9). Assessing the risk of future VTE, Cushman's longitudinal, nested case–control study demonstrated only a very mild association of high FIX levels with VTE after adjustment for age, sex, ethnicity, and study (OR, 1.4; 95% CI, 1.0–2.0).⁴⁵ This association was lost after additional adjustment for diabetes mellitus and body mass index. A mild effect of elevated FIX levels on the risk of venous thrombosis cannot be excluded.

Fibrinogen

Fibrinogen is the major procoagulatory protein in plasma by mass: its levels are between 1.5 and 3.5 g/L. An association between increasing levels of fibrinogen (for each g/L) and VTE risk was reported in a small study group of 199 patients with a first episode of thrombosis and 199 controls (OR, 1.4; 95% CI, 1.02-1.95).⁵¹ Notably, when the study group was divided in strata according to their fibrinogen levels, the 95% CIs indicated insufficient evidence to conclude that patients and controls differed significantly. Fibrinogen is an acutephase reactant and its levels increase in inflammation, but also under environmental stimuli (e.g., smoking). Therefore, a subsequent study included 474 thrombosis patients and 474 matched controls, which were part of the LETS.⁵² Highsensitive C-reactive protein (CRP) was included in the laboratory panel. The CRP-adjusted OR was 4.3 (95% CI, 1.7-10.5) for a fibrinogen level greater than 5 g/L and 1.6 (95% CI, 1.0-2.6) for a fibrinogen level greater than 4 g/L and was basically comparable to the unadjusted OR, indicating that the mild risk associated with elevated fibrinogen levels could not be attributed to inflammatory stimuli. Using the same set of data, stratification by age showed that the VTE risk associated with high levels of fibrinogen was mainly increased in older patients.⁵³ Even the highest fibringen levels (>5 g/L) were not associated with VTE risk in patients younger than 45 years (OR, 1.5; 95% CI, 0.7-3.3), whereas in patients older than 45 years, even fibrinogen levels greater than 3 g/L were associated with VTE risk (OR, 1.8; 95% CI, 1.3-2.6).

Factor XIII

FXIII becomes activated through thrombin and crosslinks fibrin γ - and α -chains, stabilizing the blood clot. The FXIII B subunit is highly polymorphic. Of three common single nucleotide polymorphisms (A8259G, C29470T, and A30899G), one (A8259G) results in an amino acid substitution, His95Arg. The relationship between His95Arg and VTE was investigated in two independent study cohorts, including 685 patients and 763 controls, revealing a pooled OR of 1.5 (95% CI, 1.1–2.0). The FXIII A subunit carries a polymorphism, Val34Leu, which was associated with elevated FXIII levels and moderate protection from VTE in some, 55 but not all, studies. A meta-analysis calculated a combined ORs for VTE of 0.63 (95% CI, 0.46–0.86) for homozygotes and 0.89 (95% CI, 0.80–0.99) for heterozygotes. 57

Anticoagulant Mechanisms

Tissue Factor Pathway Inhibitor

TFPI is a protease inhibitor and exerts its inhibitory effect mainly by inhibiting the tissue factor-FVIIa complex in a FXa-

dependent manner.⁵⁸ Accordingly, low plasma concentrations of TFPI might increase the risk for thrombosis. The OR for VTE in subjects who had free antigen levels of TFPI below the 10th percentile, as compared with those who had levels above this value, was 1.7 (95% CI, 1.1-2.6) in the Leiden Thrombophilia study.⁵⁹ A subsequently performed nested case-control study demonstrated relevant effects of various confounding factors: compared with those in the upper 95%, the bottom 5% of free TFPI levels had an age-, sex-, ethnicity-, and study-adjusted, nonsignificant OR for VTE of 1.35 (95% CI, 0.86-2.12).60 Additional adjustment for coagulation factor activities retrieved a moderate risk in this analysis (OR, 1.93; 95% CI, 1.05-3.53). In addition, moderate risk for recurrent VTE was identified in a prospective trial and patients after VTE were followed up after withdrawal of anticoagulation.⁶¹ Genetic markers for TFPI levels are more difficult to interpret. In a recent meta-analysis, the TFPI rs8176592 polymorphism was associated with an increased risk of VTE, while no association was found for rs10931292 or rs10153820 and venous thrombosis. However, associations varied between ethnical groups.62

Thrombomodulin

Thrombomodulin (TM) is an endothelial thrombin receptor that modulates the activity of thrombin toward activation of anticoagulant protein C as well as TAFI. Accordingly, lack of function of TM results in abnormalities of regulation of the coagulation-fibrinolysis system.⁶³ Single nucleotide polymorphisms or mutations of the TM gene that influence protein expression or function have been associated with complement-mediated thrombotic microangiopathy and are potentially linked to arterial thrombosis. 64-68 With respect to the risk of VTE, sequence variations in the promotor as well as in the coding regions of the TM gene (THBD) have been found to be rare and/or could not be identified as independent risk factors for VTE. 69,70 On the contrary, an even reduced risk of VTE has been described for the c.1418C > T polymorphism, which leads to replacement of Ala455 by Val in the TM molecule. This finding was substantiated by lower levels of soluble thrombomodulin (sTM) as well as gain-ofactivity of the membrane-bound molecule. 71 However, other studies failed to demonstrate an association of the c.1418C > T polymorphism with the risk of VTE. 72-74 Overall, there is currently no evidence that sequence variations in THBD correlate with an increased risk of VTE.

Endothelial Protein C Receptor

Activation of protein C on endothelial cells is augmented by the endothelial cell protein C receptor (EPCR), which promotes binding and alignment of the molecule to the thrombin–TM complex.⁷⁵ A soluble form of EPCR (sEPCR), which lacks the transmembrane domain and cytoplasmic tail, is present in plasma.⁷⁶ In contrast to transmembrane EPCR, sEPCR acts as an inhibitor of the protein C pathway by binding to both, protein C and activated protein C (APC) with similar affinity, thereby interfering with activation of the zymogen as well as the anticoagulant activity of the enzyme.⁷⁷ Sequence variants in the EPCR gene (*PROCR*) have

been found to be associated with both, cellular EPCR expression and sEPCR plasma levels. For instance, a 23bp insertion in exon 3 of PROCR that results in a stop codon downstream the insertion site has been described.⁷⁸ The EPCR 23bp insertion is very rare in patients with VTE as well as in patients with myocardial infarction. Accordingly, no association between the EPCR 23bp insertion and an increased arterial or venous thrombotic risk could be established.^{79–81} Furthermore, two PROCR haplotypes (H1 and H3) with opposed functional phenotypes have been identified. Haplotype H1, usually tagged by the 4678G > C sequence variation (3' UTR), is associated with increased plasma levels of anticoagulant APC and reduced levels of sEPCR. Haplotype H3, usually tagged by 4600A > G (exon 4, resulting in Ser219Gly), is associated with increased shedding of EPCR from the endothelium and, thus, increased levels of procoagulant sEPCR. 82 However, data from the LETS did not reveal a strong association between EPCR haplotypes and VTE risk.83 A comprehensive meta-analysis demonstrated a weak association between the H3 haplotype and the risk of VTE (OR, 1.22; 95% CI, 1.11-1.33).84 More recently, it has been described that both haplotypes may play a role with respect to the thrombotic risk in the younger population, 85 while a protective role against arterial thrombosis was suggested in patients with antiphospholipid syndrome.⁸⁶ In conclusion, there is currently only weak evidence for an association between EPCR haplotypes and the risk for VTE.

Protein Z and Protein Z-Dependent Protease Inhibitor

Protein Z (PZ) acts as a cofactor during inhibition of FXa by the PZ-dependent protease inhibitor (ZPI).87 Recent data suggest that ZPI/PZ also functions as a physiological inhibitor of prothrombinase-bound FXa, questioning the current idea that FXa within the prothrombinase complex is protected from inhibition by protease inhibitors.⁸⁸ Accordingly, low plasma levels of either PZ or ZPI may increase the thrombotic risk. Several PZ (PROZ) and ZPI gene (SERPINA10) polymorphisms that influence functional protein plasma levels have been described, while their impact on the risk of VTE remains debatable.^{89,90} In 2001, Vasse et al demonstrated a link between PZ deficiency and ischemic stroke, while no such association was found in patients with VTE.⁹¹ Regarding both PZ and ZPI plasma levels, a relationship with venous thrombosis was also not detected in the LETS. 92 In contrast to these findings, a meta-analysis comprising 28 case-control studies revealed a significant association between low PZ levels and venous thromboembolic diseases (OR, 2.18; 95% CI, 1.19-4.00).93 Furthermore, it has been proposed that low plasma levels of PZ may increase the risk of VTE associated with established thrombophilic risk factors (e.g., the FV Leiden mutation). 94,95 Regarding ZPI, several sequence variations in SERPINA10, including two stop mutations (R67X and W303X) have been identified with higher frequency in patients with thrombosis. 90,96 However, a subsequently performed meta-analysis did not reveal an association between these mutations and an increased risk for VTE.97 Also other identified SERPINA10 sequence variations showed no association with the risk for

VTE. 98,99 In conclusion, there is weak evidence for an association between low levels of PZ and VTE.

Heparin Cofactor II

Heparin cofactor II (HCII) is a member of the serine protease inhibitor (SERPIN) family that binds to dermatan sulfate on endothelial cells and inhibits thrombin. 100 However, although contradictory reports have been published, 101,102 even homozygous deficiency of HCII has not been associated with an increased risk of thrombosis. 103-106

Homocysteine Metabolism

Homocysteine is an intermediate in the metabolism of the essential amino acid methionine, which, among other food, is present in meat and dairy products. Homocysteine is toxic in higher concentrations and needs to be metabolized in the human body by one of the two following pathways: remethylation or transsulfuration. Remethylation (from homocysteine back to methionine) requires folate which will be converted to 5-methyltetrahydrofolate by the activity of an enzyme, methylene tetrahydrofolate reductase (MTHFR). Vitamin B12 is a relevant cofactor for this reaction. Transsulfuration (from homocysteine to cysteine) uses a different set of enzymes and depends on the availability of vitamin B6. Finally, the kidneys remove remaining homocysteine from the plasma. According to this complex metabolism, the following factors can affect homocysteine levels in the patient's plasma: food or food supplementation as a source of methionine, homocysteine, folate, vitamin B12, and vitamin B6; genetic polymorphisms affecting MTHFR activity; and renal function. 107

MTHFR C677T, MTHFR A1298C

Genetic polymorphisms of MTHFR can affect enzyme activity and may lead to higher levels of homocysteine, an effect which can be overcome by folate substitution. 108 Indeed, it has long been suspected that in populations with high oral intake of folate, reduced activity of MTHFR does not correlate with an increased risk of VTE. 109 A meta-analysis including 11,000 VTE cases did not find a correlation between the MTHFR C677T polymorphism and VTE.⁵ A more recent metaanalysis had two interesting findings: the MTHFR C677T polymorphism was associated with the risk of VTE, but only in Asians, not in Europeans; and the MTHFR A1298C polymorphism was not associated with the risk of VTE. 110 It is very likely that the difference between Asian and European patients is an effect of nutrition rather than of ethnicity: many countries in the western world enrich industrially manufactured food with folate. 111 Because of the aforementioned complex metabolism of homocysteine, and because of the evidence from meta-analyses, MTHFR polymorphisms should not be included in the diagnostic workup of thrombophilia.

Homocysteine Plasma Level

The mechanisms by which homocysteine increases the risk of VTE are unclear but may include effects on endothelial cells, platelets, and coagulation factors.¹¹² An association between hyperhomocysteinemia and the risk of VTE has been demonstrated in a meta-analysis (OR, 2.95; 95% CI, 2.08–4.17),¹¹³ an association that was underlined in various smaller studies thereafter.^{114–116} It should be noted that lowering the homocysteine level has no clear impact on patients with arterial thrombotic events,¹¹⁷ and evidence for VTE patients is lacking.

Other Thrombophilic Markers

Angiotensin-Converting Enzyme

The angiotensin-converting enzyme (ACE) plays a major role in the regulation of blood pressure by converting angiotensin I to angiotensin II. 118 ACE may also exert prothrombotic effects by inhibition of fibrinolysis and activation of platelets. 119 High ACE activity could therefore be associated with an increased risk of VTE. Half the variance of ACE activity is related to an insertion/deletion polymorphism, 120 and this polymorphism has been analyzed in most association studies. A meta-analysis based on 14 studies including 3,448 cases did not show evidence for an association between the ACE insertion/deletion genotype and VTE (OR, 1.206; 95% CI, 0.951-1.531). 121 In contrast, two studies in patients who were taking ACE inhibitors or angiotensin II receptor blockers and suffered from atherosclerosis or atrial fibrillation showed a reduction in the risk of venous thrombus formation. 122,123 However, no ACE polymorphism and/or data on enzyme activity levels were provided. Thus, the described effects could be based on a mechanism that differs from processes directly mediated by ACE.

Apolipoprotein E

Apolipoprotein E (ApoE) is a polymorphic glycoprotein that plays a role in cholesterol transport as well as cell membrane maintenance and repair. While polymorphisms/genotypes of the ApoE gene (APOE) are discussed to be associated with atherosclerosis, cardiovascular disease, or ischemic stroke, 125–127 data on a corresponding risk of VTE are limited and contradictory. For instance, the ApoE*2 allele was described to be linked to deep venous thrombotic events in women. 128 Furthermore, the ApoE*3/4 genotype was found to be associated with an increased risk of VTE in a small pilot study (OR, 1.31; 95% CI, 1.30–10.48). 129 In contrast, other studies did not find any significant associations. 130,131

ADAMTS13

ADAMTS13 (a disintegrin and metalloproteinase with a thrombospondin type 1 motif, member 13) is a plasma glycoprotein that regulates the size distribution of vWF.¹³² Due to the presence of uncleaved, ultra-large vWF multimers, severe lack of ADAMTS13 activity leads to thrombocytopenic purpura, a specific type of thrombotic microangiopathy.¹³³ Potential association between decreased ADAMTS13 activity and an increased risk for VTE is thought to be due to correspondingly increased vWF plasma levels.¹³⁴ Indeed, it has been shown that patients with an ADAMTS13 activity less than 1st quartile of normal showed an increased risk of VTE (OR,

1.6; 95% CI, 1.05–2.55), while the combination of low ADAMTS13 activity and high vWF antigen levels was associated with a 15-fold increased risk (95% CI, 7.80–33.80). The ruthermore, analysis by next-generation sequencing in patients with VTE without identified established risk factors revealed the presence of rare coding single-nucleotide variants within the ADAMTS13 gene (*ADAMTS13*). However, the association of these sequence variations with the risk of VTE needs to be verified. 136,137

Conclusion

When determining the causes of VTE in a patient, it can be useful to include thrombophilia screening, bearing in mind that VTE is a multifactorial disorder and numerous other factors may play a more prominent role in triggering VTE in an individual patient, such as cancer or major surgery. Evidence of thrombophilia may have impact on the duration of anticoagulation. 138 In addition, in families with a history of VTE and thrombophilia, screening of family members (and subsequent primary antithrombotic prophylaxis in situations of high thrombotic risk) may help reduce the incidence of VTE and VTE-related disease burden. 139 While established markers of thrombophilia represent the fundamentals of thrombophilia screening, they are absent in most VTE patients. In search of alternative explanations for VTE in these patients, numerous additional markers have been described. In an attempt to appraise these markers, we reviewed the corresponding literature as described in the present paper. At present, none of the markers does add significantly to clinical routine thrombophilia testing. As summarized in Table 1, current evidence suggests that, for some of these markers, an association with VTE has not been demonstrated: plasminogen, α2-antiplasmin, TM gene mutations, ZPI, MTHFR gene polymorphisms, ADAMTS13, HCII, Lp(a), and ACE. We believe that these markers are of no benefit in patient care outside studies. In cases of supposed evidence of a given association between the presence of the marker and thrombophilia, analysis may be considered, although most associations are weak or very weak and further evidence from well-designed studies is warranted: TFPI, EPCR mutations, PZ, PAI-1, TAFI, homocysteine, FIX, FXI, fibrinogen, FXIII Val34Leu.

Other perspectives are needed to better understand the potential impact of novel markers. For instance, with respect to the interpretation of genetic association studies, most approaches looked at polymorphisms as markers of plasma levels. As discussed elsewhere, it will be necessary to look more closely for effects certain mutations may have on function or activity. ¹⁴⁰ In addition, well-designed future studies must include extensive multiparameter testing, since it is evident that several players in coagulation, but also inflammation and metabolism, have significant impact on the risk of a first or recurrent VTE, also through mutual influence, in both a positive and negative sense.

Conflict of Interest

None declared.

References

- 1 Klarin D, Lynch J, Aragam K, et al; VA Million Veteran Program. Genome-wide association study of peripheral artery disease in the Million Veteran Program. Nat Med 2019;25(08):1274–1279
- 2 Chiasakul T, De Jesus E, Tong J, et al. Inherited thrombophilia and the risk of arterial ischemic stroke: a systematic review and meta-analysis. J Am Heart Assoc 2019;8(19):e012877
- 3 Mannucci PM, Franchini M. Classic thrombophilic gene variants. Thromb Haemost 2015;114(05):885–889
- 4 Linnemann B, Hart C. Laboratory diagnostics in thrombophilia. Hamostaseologie 2019;39(01):49–61
- 5 Simone B, De Stefano V, Leoncini E, et al. Risk of venous thromboembolism associated with single and combined effects of Factor V Leiden, prothrombin 20210A and methylenetetrahydrofolate reductase C677T: a meta-analysis involving over 11,000 cases and 21,000 controls. Eur J Epidemiol 2013;28 (08):621–647
- 6 Anderson FA Jr, Spencer FA. Risk factors for venous thromboembolism. Circulation 2003;107(23, Suppl 1):19–116
- 7 Law RHP, Abu-Ssaydeh D, Whisstock JC. New insights into the structure and function of the plasminogen/plasmin system. Curr Opin Struct Biol 2013;23(06):836–841
- 8 Ismail AA, Shaker BT, Bajou K. The plasminogen-activator plasmin system in physiological and pathophysiological angiogenesis. Int | Mol Sci 2021;23(01):337
- 9 Draxler DF, Sashindranath M, Medcalf RL. Plasmin: a modulator of immune function. Semin Thromb Hemost 2017;43(02):143–153
- 10 van der Vorm LN, Remijn JA, de Laat B, Huskens D. Effects of plasmin on von Willebrand factor and platelets: a narrative review. TH Open 2018;2(02):e218–e228
- 11 Kwaan HC. The role of fibrinolytic system in health and disease. Int J Mol Sci 2022;23(09):5262
- 12 Banbula A, Zimmerman TP, Novokhatny VV. Blood inhibitory capacity toward exogenous plasmin. Blood Coagul Fibrinolysis 2007;18(03):241–246
- 13 Juhan-Vague I, Renucci JF, Grimaux M, et al. Thrombin-activatable fibrinolysis inhibitor antigen levels and cardiovascular risk factors. Arterioscler Thromb Vasc Biol 2000;20(09):2156–2161
- 14 Lisman T, de Groot PG, Meijers JC, Rosendaal FR. Reduced plasma fibrinolytic potential is a risk factor for venous thrombosis. Blood 2005;105(03):1102–1105
- 15 Meltzer ME, Lisman T, de Groot PG, et al. Venous thrombosis risk associated with plasma hypofibrinolysis is explained by elevated plasma levels of TAFI and PAI-1. Blood 2010;116(01):113–121
- 16 Mingers AM, Heimburger N, Zeitler P, Kreth HW, Schuster V. Homozygous type I plasminogen deficiency. Semin Thromb Hemost 1997;23(03):259–269
- 17 Schuster V, Hügle B, Tefs K. Plasminogen deficiency. J Thromb Haemost 2007;5(12):2315–2322
- 18 Tsutsumi S, Saito T, Sakata T, Mlyata T, Ichinose A. Genetic diagnosis of dysplasminogenemia: detection of an Ala601-Thr mutation in 118 out of 125 families and identification of a new Asp676-Asn mutation. Thromb Haemost 1996;76(02):135–138
- 19 Dawson SJ, Wiman B, Hamsten A, Green F, Humphries S, Henney AM. The two allele sequences of a common polymorphism in the promoter of the plasminogen activator inhibitor-1 (PAI-1) gene respond differently to interleukin-1 in HepG2 cells. J Biol Chem 1993;268(15):10739-10745
- 20 Tsantes AE, Nikolopoulos GK, Bagos PG, et al. Association between the plasminogen activator inhibitor-1 4G/5G polymorphism and venous thrombosis. A meta-analysis. Thromb Haemost 2007;97(06):907–913
- 21 Nikolopoulos GK, Bagos PG, Tsangaris I, et al. The association between plasminogen activator inhibitor type 1 (PAI-1) levels, PAI-1 4G/5G polymorphism, and myocardial infarction: a Mendelian randomization meta-analysis. Clin Chem Lab Med 2014; 52(07):937–950

- 22 Tsantes AE, Nikolopoulos GK, Bagos PG, et al. Plasminogen activator inhibitor-1 4G/5G polymorphism and risk of ischemic stroke: a meta-analysis. Blood Coagul Fibrinolysis 2007;18(05): 497–504
- 23 Mellbring G, Dahlgren S, Reiz S, Wiman B. Fibrinolytic activity in plasma and deep vein thrombosis after major abdominal surgery. Thromb Res 1983;32(06):575–584
- 24 Páramo JA, Alfaro MJ, Rocha E. Postoperative changes in the plasmatic levels of tissue-type plasminogen activator and its fast-acting inhibitor-relationship to deep vein thrombosis and influence of prophylaxis. Thromb Haemost 1985;54(03): 713–716
- 25 Booth NA. Fibrinolysis and thrombosis. Best Pract Res Clin Haematol 1999;12(03):423–433
- 26 Sillen M, Declerck PJ. Thrombin activatable fibrinolysis inhibitor (TAFI): an updated narrative review. Int J Mol Sci 2021;22(07): 3670
- 27 van Tilburg NH, Rosendaal FR, Bertina RM. Thrombin activatable fibrinolysis inhibitor and the risk for deep vein thrombosis. Blood 2000;95(09):2855–2859
- 28 Eichinger S, Schönauer V, Weltermann A, et al. Thrombin-activatable fibrinolysis inhibitor and the risk for recurrent venous thromboembolism. Blood 2004;103(10):3773–3776
- 29 Folkeringa N, Coppens M, Veeger NJ, et al. Absolute risk of venous and arterial thromboembolism in thrombophilic families is not increased by high thrombin-activatable fibrinolysis inhibitor (TAFI) levels. Thromb Haemost 2008;100(01):38–44
- 30 Boffa MB, Marar TT, Yeang C, et al. Potent reduction of plasma lipoprotein (a) with an antisense oligonucleotide in human subjects does not affect ex vivo fibrinolysis. J Lipid Res 2019; 60(12):2082–2089
- 31 Reyes-Soffer G, Ginsberg HN, Berglund L, et al; American Heart Association Council on Arteriosclerosis, Thrombosis and Vascular Biology; Council on Cardiovascular Radiology and Intervention; and Council on Peripheral Vascular Disease. Lipoprotein(a): a genetically determined, causal, and prevalent risk factor for atherosclerotic cardiovascular disease: a scientific statement from the American Heart Association. Arterioscler Thromb Vasc Biol 2022;42(01):e48–e60
- 32 Dentali F, Gessi V, Marcucci R, Gianni M, Grandi AM, Franchini M. Lipoprotein(a) as a risk factor for venous thromboembolism: a systematic review and meta-analysis of the literature. Semin Thromb Hemost 2017;43(06):614–620
- 33 Kunutsor SK, Mäkikallio TH, Kauhanen J, Voutilainen A, Laukkanen JA. Lipoprotein(a) is not associated with venous thromboembolism risk. Scand Cardiovasc J 2019;53(03):125–132
- 34 Helgadottir A, Gretarsdottir S, Thorleifsson G, et al. Apolipoprotein(a) genetic sequence variants associated with systemic atherosclerosis and coronary atherosclerotic burden but not with venous thromboembolism. J Am Coll Cardiol 2012;60(08): 722–729
- 35 Kamstrup PR, Tybjærg-Hansen A, Nordestgaard BG. Genetic evidence that lipoprotein(a) associates with atherosclerotic stenosis rather than venous thrombosis. Arterioscler Thromb Vasc Biol 2012;32(07):1732–1741
- 36 Marston NA, Gurmu Y, Melloni GEM, et al. The effect of PCSK9 (proprotein convertase subtilisin/kexin type 9) inhibition on the risk of venous thromboembolism. Circulation 2020;141(20): 1600–1607
- 37 Romiti GF, Corica B, Borgi M, et al. Inherited and acquired thrombophilia in adults with retinal vascular occlusion: a systematic review and meta-analysis. J Thromb Haemost 2020;18 (12):3249–3266
- 38 Ponto KA, Scharrer I, Binder H, et al. Hypertension and multiple cardiovascular risk factors increase the risk for retinal vein occlusions: results from the Gutenberg Retinal Vein Occlusion Study. J Hypertens 2019;37(07):1372–1383

- 39 Paciullo F, Giannandrea D, Virgili G, Cagini C, Gresele P. Role of increased lipoprotein (a) in retinal vein occlusion: a systematic review and meta-analysis. TH Open 2021;5(03):e295–e302
- 40 Kamphuisen PW, Eikenboom JCJ, Bertina RM. Elevated factor VIII levels and the risk of thrombosis. Arterioscler Thromb Vasc Biol 2001;21(05):731–738
- 41 Kyrle PA, Minar E, Hirschl M, et al. High plasma levels of factor VIII and the risk of recurrent venous thromboembolism. N Engl J Med 2000;343(07):457–462
- 42 Zambelli R, Nemeth B, Touw CE, Rosendaal FR, Rezende SM, Cannegieter SC. High risk of venous thromboembolism after orthopedic surgery in patients with thrombophilia. J Thromb Haemost 2021;19(02):444–451
- 43 Tripodi A, Chantarangkul V, Martinelli I, Bucciarelli P, Mannucci PM. A shortened activated partial thromboplastin time is associated with the risk of venous thromboembolism. Blood 2004; 104(12):3631–3634
- 44 Meijers JCM, Tekelenburg WLH, Bouma BN, Bertina RM, Rosendaal FR. High levels of coagulation factor XI as a risk factor for venous thrombosis. N Engl J Med 2000;342(10):696–701
- 45 Cushman M, O'Meara ES, Folsom AR, Heckbert SR. Coagulation factors IX through XIII and the risk of future venous thrombosis: the longitudinal investigation of thromboembolism etiology. Blood 2009;114(14):2878–2883
- 46 Folsom AR, Tang W, Roetker NS, Heckbert SR, Cushman M, Pankow JS. Prospective study of circulating factor XI and incident venous thromboembolism: the longitudinal investigation of thromboembolism etiology (LITE). Am J Hematol 2015;90(11):1047–1051
- 47 Libourel EJ, Bank I, Meinardi JR, et al. Co-segregation of thrombophilic disorders in factor V Leiden carriers; the contributions of factor VIII, factor XI, thrombin activatable fibrinolysis inhibitor and lipoprotein(a) to the absolute risk of venous thromboembolism. Haematologica 2002;87(10):1068–1073
- 48 Bruzelius M, Ljungqvist M, Bottai M, et al. F11 is associated with recurrent VTE in women. A prospective cohort study. Thromb Haemost 2016;115(02):406–414
- 49 Manco L, Silva C, Fidalgo T, Martinho P, Sarmento AB, Ribeiro ML. Venous thromboembolism risk associated with ABO, F11 and FGG loci. Blood Coagul Fibrinolysis 2018;29(06):528–532
- 50 van Hylckama Vlieg A, van der Linden IK, Bertina RM, Rosendaal FR. High levels of factor IX increase the risk of venous thrombosis. Blood 2000;95(12):3678–3682
- 51 Koster T, Rosendaal FR, Reitsma PH, van der Velden PA, Briët E, Vandenbroucke JP. Factor VII and fibrinogen levels as risk factors for venous thrombosis. A case-control study of plasma levels and DNA polymorphisms the Leiden Thrombophilia Study (LETS). Thromb Haemost 1994;71(06):719–722
- 52 Kamphuisen PW, Eikenboom JCJ, Vos HL, et al. Increased levels of factor VIII and fibrinogen in patients with venous thrombosis are not caused by acute phase reactions. Thromb Haemost 1999;81 (05):680–683
- 53 van Hylckama Vlieg A, Rosendaal FR. High levels of fibrinogen are associated with the risk of deep venous thrombosis mainly in the elderly. J Thromb Haemost 2003;1(12):2677–2678
- 54 Komanasin N, Catto AJ, Futers TS, van Hylckama Vlieg A, Rosendaal FR, Ariëns RAS. A novel polymorphism in the factor XIII B-subunit (His95Arg): relationship to subunit dissociation and venous thrombosis. J Thromb Haemost 2005;3(11):2487–2496
- 55 Franco RF, Reitsma PH, Lourenço D, et al. Factor XIII Val34Leu is a genetic factor involved in the etiology of venous thrombosis. Thromb Haemost 1999;81(05):676–679
- 56 Wells PS, Anderson JL, Rodger MA, Carson N, Grimwood RL, Doucette SP. The factor XIII Val34Leu polymorphism: is it protective against idiopathic venous thromboembolism? Blood Coagul Fibrinolysis 2006;17(07):533–538
- 57 Wells PS, Anderson JL, Scarvelis DK, Doucette SP, Gagnon F. Factor XIII Val34Leu variant is protective against venous thromboem-

- bolism: a HuGE review and meta-analysis. Am J Epidemiol 2006; 164(02):101–109
- 58 Ellery PER, Adams MJ. Tissue factor pathway inhibitor: then and now. Semin Thromb Hemost 2014;40(08):881–886
- 59 Dahm A, Van Hylckama Vlieg A, Bendz B, Rosendaal F, Bertina RM, Sandset PM. Low levels of tissue factor pathway inhibitor (TFPI) increase the risk of venous thrombosis. Blood 2003;101 (11):4387–4392
- 60 Zakai NA, Lutsey PL, Folsom AR, Heckbert SR, Cushman M. Total tissue factor pathway inhibitor and venous thrombosis. The Longitudinal Investigation of Thromboembolism Etiology. Thromb Haemost 2010;104(02):207–212
- 61 Hoke M, Kyrle PA, Minar E, et al. Tissue factor pathway inhibitor and the risk of recurrent venous thromboembolism. Thromb Haemost 2005;94(04):787–790
- 62 Zhang Y, Pang A, Zhao L, et al. Association of TFPI polymorphisms rs8176592, rs10931292, and rs10153820 with venous thrombosis: a meta-analysis. Medicine (Baltimore) 2019;98 (12):e14978
- 63 Okada M, Tominaga N, Honda G, et al. A case of thrombomodulin mutation causing defective thrombin binding with absence of protein C and TAFI activation. Blood Adv 2020;4(12):2631–2639
- 64 Delvaeye M, Noris M, De Vriese A, et al. Thrombomodulin mutations in atypical hemolytic-uremic syndrome. N Engl J Med 2009;361(04):345–357
- 65 Ohlin AK, Norlund L, Marlar RA. Thrombomodulin gene variations and thromboembolic disease. Thromb Haemost 1997;78 (01):396–400
- 66 Ireland H, Kunz G, Kyriakoulis K, Stubbs PJ, Lane DA. Thrombomodulin gene mutations associated with myocardial infarction. Circulation 1997;96(01):15–18
- 67 Doggen CJ, Kunz G, Rosendaal FR, et al. A mutation in the thrombomodulin gene, 127G to A coding for Ala25Thr, and the risk of myocardial infarction in men. Thromb Haemost 1998;80 (05):743–748
- 68 Norlund L, Holm J, Zöller B, Ohlin AK. A common thrombomodulin amino acid dimorphism is associated with myocardial infarction. Thromb Haemost 1997;77(02):248–251
- 69 Le Flem L, Picard V, Emmerich J, et al. Mutations in promoter region of thrombomodulin and venous thromboembolic disease. Arterioscler Thromb Vasc Biol 1999;19(04):1098–1104
- 70 Heit JA, Petterson TM, Owen WG, Burke JP, DE Andrade M, Melton LJ III. Thrombomodulin gene polymorphisms or haplotypes as potential risk factors for venous thromboembolism: a population-based case-control study. J Thromb Haemost 2005;3(04): 710–717
- 71 Navarro S, Medina P, Bonet E, et al. Association of the thrombomodulin gene c.1418C>T polymorphism with thrombomodulin levels and with venous thrombosis risk. Arterioscler Thromb Vasc Biol 2013;33(06):1435–1440
- 72 van der Velden PA, Krommenhoek-Van Es T, Allaart CF, Bertina RM, Reitsma PH. A frequent thrombomodulin amino acid dimorphism is not associated with thrombophilia. Thromb Haemost 1991;65(05):511–513
- 73 Aleksic N, Folsom AR, Cushman M, Heckbert SR, Tsai MY, Wu KK. Prospective study of the A455V polymorphism in the thrombomodulin gene, plasma thrombomodulin, and incidence of venous thromboembolism: the LITE Study. J Thromb Haemost 2003;1(01):88–94
- 74 Ahmad A, Sundquist K, Zöller B, Svensson PJ, Sundquist J, Memon AA. Thrombomodulin gene c.1418C>T polymorphism and risk of recurrent venous thromboembolism. J Thromb Thrombolysis 2016;42(01):135–141
- 75 Stearns-Kurosawa DJ, Kurosawa S, Mollica JS, Ferrell GL, Esmon CT. The endothelial cell protein C receptor augments protein C activation by the thrombin-thrombomodulin complex. Proc Natl Acad Sci U S A 1996;93(19):10212–10216

- 76 Kurosawa S, Stearns-Kurosawa DJ, Hidari N, Esmon CT. Identification of functional endothelial protein C receptor in human plasma. J Clin Invest 1997;100(02):411–418
- 77 Liaw PC, Neuenschwander PF, Smirnov MD, Esmon CT. Mechanisms by which soluble endothelial cell protein C receptor modulates protein C and activated protein C function. J Biol Chem 2000;275(08):5447–5452
- 78 Biguzzi E, Merati G, Liaw PC, et al. A 23bp insertion in the endothelial protein C receptor (EPCR) gene impairs EPCR function. Thromb Haemost 2001;86(04):945–948
- 79 von Depka M, Czwalinna A, Eisert R, et al. Prevalence of a 23bp insertion in exon 3 of the endothelial cell protein C receptor gene in venous thrombophilia. Thromb Haemost 2001;86(06): 1360–1362
- 80 Van de Water NS, French JK, McDowell J, Browett PJ. The endothelial protein C receptor (EPCR) 23bp insert in patients with myocardial infarction. Thromb Haemost 2001;85(04): 749–751
- 81 Eroĝlu A, Ulu A, Kurtman C, Cam R, Akar N. 23-bp endothelial protein C receptor (EPCR) gene insertion mutation in cancer patients with and without thrombosis. Am J Hematol 2006;81 (03):220
- 82 Medina P, Navarro S, Bonet E, et al. Functional analysis of two haplotypes of the human endothelial protein C receptor gene. Arterioscler Thromb Vasc Biol 2014;34(03):684–690
- 83 Uitte de Willige S, Van Marion V, Rosendaal FR, Vos HL, de Visser MC, Bertina RM. Haplotypes of the EPCR gene, plasma sEPCR levels and the risk of deep venous thrombosis. J Thromb Haemost 2004;2(08):1305–1310
- 84 Dennis J, Johnson CY, Adediran AS, et al. The endothelial protein C receptor (PROCR) Ser219Gly variant and risk of common thrombotic disorders: a HuGE review and meta-analysis of evidence from observational studies. Blood 2012;119(10): 2392–2400
- 85 Anastasiou G, Politou M, Rallidis L, et al. Endothelial protein C receptor gene variants and risk of thrombosis. Clin Appl Thromb Hemost 2016;22(02):199–204
- 86 Plasín-Rodríguez MA, Rodríguez-Pintó I, Patricio P, et al. The H1 haplotype of the endothelial protein C receptor protects against arterial thrombosis in patients with antiphospholipid syndrome. Thromb Res 2018;169:128–134
- 87 Broze GJ Jr. Protein Z-dependent regulation of coagulation. Thromb Haemost 2001;86(01):8–13
- 88 Huang X, Swanson R, Kroh HK, Bock PE. Protein Z-dependent protease inhibitor (ZPI) is a physiologically significant inhibitor of prothrombinase function. J Biol Chem 2019;294(19):7644–7657
- 89 Bafunno V, Santacroce R, Margaglione M. The risk of occurrence of venous thrombosis: focus on protein Z. Thromb Res 2011;128 (06):508-515
- 90 Van de Water N, Tan T, Ashton F, et al. Mutations within the protein Z-dependent protease inhibitor gene are associated with venous thromboembolic disease: a new form of thrombophilia. Br J Haematol 2004;127(02):190–194
- 91 Vasse M, Guegan-Massardier E, Borg JY, Woimant F, Soria C. Frequency of protein Z deficiency in patients with ischaemic stroke. Lancet 2001;357(9260):933–934
- 92 Al-Shanqeeti A, van Hylckama Vlieg A, Berntorp E, Rosendaal FR, Broze GJJr. Protein Z and protein Z-dependent protease inhibitor. Determinants of levels and risk of venous thrombosis. Thromb Haemost 2005;93(03):411–413
- 93 Sofi F, Cesari F, Abbate R, Gensini GF, Broze G Jr, Fedi S. A metaanalysis of potential risks of low levels of protein Z for diseases related to vascular thrombosis. Thromb Haemost 2010;103(04): 749–756
- 94 Kemkes-Matthes B, Nees M, Kühnel G, Matzdorff A, Matthes KJ.
 Protein Z influences the prothrombotic phenotype in factor V
 Leiden patients. Thromb Res 2002;106(4-5):183–185

- 95 Martinelli I, Razzari C, Biguzzi E, Bucciarelli P, Mannucci PM. Low levels of protein Z and the risk of venous thromboembolism. J Thromb Haemost 2005;3(12):2817–2819
- 96 Corral J, González-Conejero R, Soria JM, et al. A nonsense polymorphism in the protein Z-dependent protease inhibitor increases the risk for venous thrombosis. Blood 2006;108(01): 177–183
- 97 Dentali F, Gianni M, Lussana F, Squizzato A, Cattaneo M, Ageno W. Polymorphisms of the Z protein protease inhibitor and risk of venous thromboembolism: a meta-analysis. Br J Haematol 2008; 143(02):284–287
- 98 Young LK, Birch NP, Browett PJ, et al. Two missense mutations identified in venous thrombosis patients impair the inhibitory function of the protein Z dependent protease inhibitor. Thromb Haemost 2012;107(05):854–863
- 99 Gorski MM, Lotta LA, Pappalardo E, et al. Single nucleotide variant rs2232710 in the protein Z-dependent protease inhibitor (ZPI, SERPINA10) gene is not associated with deep vein thrombosis. PLoS One 2016;11(03):e0151347
- 100 Briginshaw GF, Shanberge JN. Identification of two distinct heparin cofactors in human plasma. Separation and partial purification. Arch Biochem Biophys 1974;161(02):683–690
- 101 Lopaciuk S, Bykowska K, Kopeć M Prevalence of heparin cofactor II deficiency in patients with a history of venous thrombosis. Pol J Pharmacol 1996;48(01):109–111
- 102 Weisdorf DJ, Edson JR. Recurrent venous thrombosis associated with inherited deficiency of heparin cofactor II. Br J Haematol 1991;77(01):125–126
- 103 Bertina RM, van der Linden IK, Engesser L, Muller HP, Brommer EJ. Hereditary heparin cofactor II deficiency and the risk of development of thrombosis. Thromb Haemost 1987;57(02): 196–200
- 104 Villa P, Aznar J, Vaya A, et al. Hereditary homozygous heparin cofactor II deficiency and the risk of developing venous thrombosis. Thromb Haemost 1999;82(03):1011–1014
- 105 Tollefsen DM. Heparin cofactor II deficiency. Arch Pathol Lab Med 2002;126(11):1394–1400
- 106 Corral J, Aznar J, Gonzalez-Conejero R, et al. Homozygous deficiency of heparin cofactor II: relevance of P17 glutamate residue in serpins, relationship with conformational diseases, and role in thrombosis. Circulation 2004;110(10):1303–1307
- 107 Finkelstein JD, Martin JJ. Homocysteine. Int J Biochem Cell Biol 2000;32(04):385–389
- 108 Holmes MV, Newcombe P, Hubacek JA, et al. Effect modification by population dietary folate on the association between MTHFR genotype, homocysteine, and stroke risk: a meta-analysis of genetic studies and randomised trials. Lancet 2011;378(9791):584–594
- 109 Den Heijer M, Lewington S, Clarke R. Homocysteine, MTHFR and risk of venous thrombosis: a meta-analysis of published epidemiological studies. J Thromb Haemost 2005;3(02):292–299
- 110 Gao M, Feng N, Zhang M, Ti X, Zuo X. Meta-analysis of the relationship between methylenetetrahydrofolate reductase C677T and A1298C polymorphism and venous thromboembolism in the Caucasian and Asian. Biosci Rep 2020;40(07):40
- 111 Revuelta JL, Serrano-Amatriain C, Ledesma-Amaro R, Jiménez A. Formation of folates by microorganisms: towards the biotechnological production of this vitamin. Appl Microbiol Biotechnol 2018;102(20):8613–8620
- 112 D'Angelo A, Selhub J. Homocysteine and thrombotic disease. Blood 1997;90(01):1–11
- 113 Ray JG. Meta-analysis of hyperhomocysteinemia as a risk factor for venous thromboembolic disease. Arch Intern Med 1998;158 (19):2101–2106
- 114 Langman LJ, Ray JG, Evrovski J, Yeo E, Cole DEC. Hyperhomocyst (e)inemia and the increased risk of venous thromboembolism: more evidence from a case-control study. Arch Intern Med 2000; 160(07):961–964

- 115 Božič M, Stegnar M, Fermo I, et al. Mild hyperhomocysteinemia and fibrinolytic factors in patients with history of venous thromboembolism. Thromb Res 2000;100(04):271–278
- 116 Kosch A, Koch HG, Heinecke A, Kurnik K, Heller C, Nowak-Göttl UChildhood Thrombophilia Study Group. Increased fasting total homocysteine plasma levels as a risk factor for thromboembolism in children. Thromb Haemost 2004;91(02):308–314
- 117 Martí-Carvajal AJ, Solà I, Lathyris D, Salanti G. Homocysteine lowering interventions for preventing cardiovascular events. Cochrane Database Syst Rev 2009;(04):CD006612
- 118 Bernstein KE, Giani JF, Shen XZ, Gonzalez-Villalobos RA. Renal angiotensin-converting enzyme and blood pressure control. Curr Opin Nephrol Hypertens 2014;23(02):106–112
- 119 Brown NJ, Vaughan DE. Prothrombotic effects of angiotensin. Adv Intern Med 2000;45:419-429
- 120 Rigat B, Hubert C, Alhenc-Gelas F, Cambien F, Corvol P, Soubrier F. An insertion/deletion polymorphism in the angiotensin I-converting enzyme gene accounting for half the variance of serum enzyme levels. J Clin Invest 1990;86(04):1343–1346
- 121 Hsiao F-C, Hsu L-A. Meta-analysis of association between insertion/deletion polymorphism of the angiotensin I-converting enzyme gene and venous thromboembolism. Clin Appl Thromb Hemost 2011;17(01):51–57
- 122 Chae YK, Khemasuwan D, Dimou A, et al. Inhibition of renin angiotensin axis may be associated with reduced risk of developing venous thromboembolism in patients with atherosclerotic disease. PLoS One 2014;9(01):e87813–e87813
- 123 Suo Y, Zhang Y, Wang Y, et al. Renin-angiotensin system inhibition is associated with reduced risk of left atrial appendage thrombosis formation in patients with atrial fibrillation. Cardiol J 2018;25(05):611–620
- 124 Mahley RW. Apolipoprotein E: cholesterol transport protein with expanding role in cell biology. Science 1988;240(4852):622–630
- 125 Cattin L, Fisicaro M, Tonizzo M, et al. Polymorphism of the apolipoprotein E gene and early carotid atherosclerosis defined by ultrasonography in asymptomatic adults. Arterioscler Thromb Vasc Biol 1997;17(01):91–94
- 126 Eichner JE, Dunn ST, Perveen G, Thompson DM, Stewart KE, Stroehla BC. Apolipoprotein E polymorphism and cardiovascular disease: a HuGE review. Am J Epidemiol 2002;155(06):487–495
- 127 Qiao SY, Shang K, Chu YH, et al. Apolipoprotein E ε4 polymorphism as a risk factor for ischemic stroke: a systematic review and meta-analysis. Dis Markers 2022;2022:1407183

- 128 Nagato LC, de Souza Pinhel MA, de Godoy JM, Souza DR. Association of ApoE genetic polymorphisms with proximal deep venous thrombosis. J Thromb Thrombolysis 2012;33(01): 116–119
- 129 Katrancioglu N, Manduz S, Ozen F, et al. Association between ApoE4 allele and deep venous thrombosis: a pilot study. Clin Appl Thromb Hemost 2011;17(02):225–228
- 130 Zhu S, Wang Z, Wu X, Shu Y, Lu D. Apolipoprotein E polymorphism is associated with lower extremity deep venous thrombosis: color-flow Doppler ultrasound evaluation. Lipids Health Dis 2014;13:21
- 131 Rastogi P, Kumar N, Ahluwalia J, et al. Thrombophilic risk factors are laterally associated with apolipoprotein E gene polymorphisms in deep vein thrombosis patients: an Indian study. Phlebology 2019;34(05):324–335
- 32 South K, Lane DA. ADAMTS-13 and von Willebrand factor: a dynamic duo. J Thromb Haemost 2018;16(01):6–18
- 33 Joly BS, Coppo P, Veyradier A. Thrombotic thrombocytopenic purpura. Blood 2017;129(21):2836–2846
- 34 Bittar LF, de Paula EV, Mello TB, Siqueira LH, Orsi FL, Annichino-Bizzacchi JM. Polymorphisms and mutations in vWF and ADAMTS13 genes and their correlation with plasma levels of FVIII and vWF in patients with deep venous thrombosis. Clin Appl Thromb Hemost 2011;17(05):514–518
- 135 Pagliari MT, Boscarino M, Cairo A, et al. ADAMTS13 activity, high VWF and FVIII levels in the pathogenesis of deep vein thrombosis. Thromb Res 2021;197:132–137
- 136 Lotta LA, Tuana G, Yu J, et al. Next-generation sequencing study finds an excess of rare, coding single-nucleotide variants of ADAMTS13 in patients with deep vein thrombosis. J Thromb Haemost 2013;11(07):1228–1239
- 137 Pagliari MT, Cairo A, Boscarino M, et al. Role of ADAMTS13, VWF and F8 genes in deep vein thrombosis. PLoS One 2021;16(10): e0258675
- 138 Connors JM. Thrombophilia testing and venous thrombosis. N Engl J Med 2017;377(12):1177–1187
- 139 Mazzolai L, Duchosal MA. Hereditary thrombophilia and venous thromboembolism: critical evaluation of the clinical implications of screening. Eur J Vasc Endovasc Surg 2007;34(04): 483–488
- 140 Lowe G. Factor IX and deep vein thrombosis. Haematologica 2009;94(05):615–617