A Prospective Multicenter Randomized Controlled Trial to Evaluate the Efficacy of Chitosan Hydrogel Paste in Comparison to Commercial Hydroactive Gel as a Wound Bed Preparation

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Abstract

Background This clinical trial aimed to evaluate the clinical efficacy of chitosan derivative hydrogel paste (CDHP) as a wound bed preparation for wounds with cavities.

Methods This study enrolled 287 patients, with 143 patients randomized into the CDHP group (treatment) and 144 patients randomized into the commercial hydroactive gel (CHG) group (control). The granulation tissue, necrotic tissue, patient comfort, clinical signs, symptoms, and patient convenience during the application and removal of the dressing were assessed.

Results The study was completed by 111 and 105 patients from the treatment and control groups, respectively. Both groups showed an increasing mean percentage of wound granulation over time when the initial wound size and comorbidity were adjusted ($F(10,198) = 4.61; p < 0.001$), but no significant difference was found between the groups ($F(1,207) = 0.043; p = 0.953$). The adjusted mean percentage of necrotic tissue of both groups showed a significant decrease over time ($F(10,235) = 5.65; p < 0.001$), but no significant differences were found between the groups ($F(1,238) = 0.487; p = 0.486$).

Conclusion CDHP is equivalent to CHG and is an alternative in wound management and wound bed preparation for wounds with cavities.

Keywords ► chitosan dressing ► full-thickness wound ► cavity wounds ► granulation

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Introduction

Chitosan is a biodegradable, nontoxic, complex carbohydrate derivative of chitin (poly-N-acetyl-D-glucosamine),1,2 a naturally occurring material found in crustacean exoskeletons (such as crabs and shrimp) and the cell walls of fungi.2–4 Chitosan possesses various biological properties, such as hemostasis5–8 and acedesyne activity (analgesic), wound healing properties,9,10 scar reduction, bacteriostasis,11,12 biocompatibility, and biodegradability.13–17 These properties make chitosan a suitable candidate as a prospective material for wound dressings.

Chitosan is water insoluble but can be made water soluble either through derivatization of its functional groups or hydrolysis of the long polymeric chains into shorter chain length chitosan products called oligochitosan.18 One such derivative is N,O-CMC, a water-soluble chitosan derivative bearing carboxymethyl substituents at some of both amine and 6-hydroxyl sites of glucosamine units. Moisture retention, gel performance capability, low toxicity, and good biocompatibility are a few of the physical and biological features that make chitosan a promising biomaterial.19

Various dressings are available, indicating that no single dressing material can be considered the gold standard and suitable for all wound types.20,21 Ideally, wound dressings should be selected based on their ability to enhance re-epithelialization as well as angiogenesis and extracellular matrix (ECM) synthesis. To enhance blood flow for angiogenesis, the ideal dressing must be sufficiently permeable to allow gas exchange while maintaining tissue temperature.22

Furthermore, the dressing must be nonadherent and easy to remove, preventing damage to the new epithelialized and granulation tissues, and the dressing must provide a debridement factor for easy removal of necrotic tissue.20,21,23,24 Wound dressings are generally classified as passive, interactive, or bioactive (advance)25,26 products that contain an active compound that aids wound healing.

Gel dressings are widely used as a wound treatment. The most common hydroactive gels include Intrasite, Hydrosorb, Purilon, and DuoDERM hydroactive gels. Commercial hydroactive gel (CHG) is a viscous hydrogel that is preservative-free and transparent. It has been recommended for partial thickness wounds with a cavity that required wound bed preparation for a subsequent surgery/procedure to close the wound at a later stage.

Patients and Methods

This prospective, multicenter, randomized controlled clinical trial involved patients at our study centers with full-thickness wounds with a cavity that required wound bed preparation for a subsequent surgery/procedure to close the wound at a later stage.

Selection of Patients

Patients of both sexes, aged 16 to 70 years, who presented with a full-thickness wound with a cavity at the four medical centers in Malaysia were screened for the study. Patients with severely contaminated or infected wounds, allergies to seafood, uncontrolled diabetes (random blood glucose > 10 mmol/L), noncompliance, pregnancy, and any skin pathology (such as eczema) were excluded from the study. Patients who fulfilled the inclusion criteria were enrolled in the study after providing signed consent and were randomized into either the treatment groups with CDHP dressings or control group with commercially available CHG dressings. Randomization was conducted using web-based software (http://www.randomization.com).

Preparation of CDHP Dressing

This dressing was manufactured in a good manufacturing practice-compliant pilot plant facility by our collaborator, Standard and Industrial Research Institute Malaysia (SIRIM Malaysia). This product was granted a Malaysian patent with grant no. MY-145085-A on 30 December 2011 under the Intellectual Property Corporation of Malaysia (MyIPO).

Hydroactive Dressing

The hydroactive gel dressing was purchased from our local supplier.

Dressing Application

Wound toilets were treated with 0.9% normal saline and 0.5% chlorhexidine aqueous solution until clean and free of dirt or foreign bodies. The dressing material, either CDHP or CHG, was applied to the wound, and a secondary dressing was applied using an Opsite Flexigrid with gauze on top and secured with a bandage. The dressing was changed every 1 to 3 days based on the wound condition that was decided by the treating surgeon/physician. The wounds were photographed during wound inspection and before a new dressing was applied (Fig. 1).

Measurement Outcomes

The patient details and all clinical characteristics were recorded on a case report form by our trained research assistants (staff nurses/medical assistants). The time until the wound bed is ready, granulation tissue, necrotic tissue, patient comfort (pain and itching), clinical appearance (wound drainage, erythema, localized warmth, and edema), convenience during application and removal of the dressings, such as pain following removal, exudate, adherence, ease of removal, and odor, were all noted. Additionally, notes were made regarding allergies and complications caused by the dressings.

The formation of granulation tissue (percentage of area), quality of granulation tissue, wound contraction (percentage of area), and exudates and pain were evaluated using a scale of 0 to 3 (where 0 denotes nil/none and 3 denotes maximum).27 Patient comfort (pain and itching) and pain following removal of the dressings were assessed using a visual analog scale of 0 to 10 (where 0 denotes nil/none and 10 denotes maximum).
Statistical Methods
The data were analyzed using IBM SPSS Statistic version 24. Numerical variables were summarized as means and standard deviations, and categorical variables were presented as frequencies and percentages. Baseline comparisons of variables were performed using Pearson’s chi-squared test, independent t-test, and one-way analysis of variance. Differences in scores between groups at any time point were analyzed using independent t-test. Repeated measures analysis of covariance was used to analyze the time effect, treatment effect, time–treatment interaction effect for the granulation, necrotic tissue percentages, the assessments of wound characteristics, dressing, granulation tissue, and complications. All analyses were adjusted for initial wound size and diabetes comorbidity. The level of significance was set at a p-value of 0.05.

Results
A total of 287 patients were included during the study period from April 2012 until May 2019. Of these, 143 patients were randomized into the treatment group, and 144 patients were randomized into the control group. One hundred eleven and 105 patients from the treatment and control groups, respectively, completed the study, while 58 patients (25 and 33 from the treatment and control groups, respectively) were discontinued for various reasons.

A comparison of the patients’ background characteristics and baseline data is shown in –Table 1. Both groups were comparable except for the days to complete the study (healing duration) (p = 0.032) and initial wound size (p = 0.025). Because of the higher initial wound size, the duration of healing in the treatment group was longer time than that of the control group.

When the healing duration was compared further, significant mean differences were observed in the healing duration among the age group, diabetes, and wound etiology (–Table 2). The younger group and nondiabetic patients healed faster than the other groups. Those with motor vehicle accidents and trauma healed faster than those with infection and surgical complications.

The wounds were located at various parts of the body, and most were at the lower limbs for both groups with number of wounds are 95 and 76, respectively (–Fig. 2A). –Fig. 2B compares the healing times of different types of wound edges. Most of the wounds had sloping edges. Although the punch-out
wound took longer to heal, no significant mean differences were found between the groups ($p = 0.087$).

A significant time effect was observed on the percentage of granulation when the initial wound area and comorbidity were adjusted ($F_{(10, 198)} = 4.61; p < 0.001$). However, no statistical significance was observed regarding the effect of treatment ($F_{(1, 207)} = 0.043; p = 0.836$). A significant interaction effect was observed between treatments and time when other variables were adjusted ($F_{(10, 198)} = 2.17; p = 0.021$). Both the treatment and control groups showed an increasing mean percentage of wound granulation at the respective time points (►Fig. 2C). The granulation tissue formation was evident in the wound healing progress (►Fig. 3).

►Fig. 2D shows a significant time effect on the percentage of necrotic tissue when the initial wound area and comorbidity were adjusted ($F_{(10, 235)} = 5.65; p < 0.001$). However, no significant effects of the treatments ($F_{(1, 244)} = 0.487; p = 0.486$) or interaction effects, $F_{(10, 235)} = 1.44, p = 0.162$ were noted. Both groups showed a decreasing mean percentage of necrotic tissue at the respective time points.

Both the treatment and control groups had significantly decreased means in wound drainage and erythema over time. However, no significant differences were found between the groups and time–treatment interaction in all clinical signs and symptoms of the wound (►Table 3).

### Table 1 The distribution of background and baseline parameters of treatment and control groups

<table>
<thead>
<tr>
<th></th>
<th>Frequency (%)/mean (SD)</th>
<th>$p$-Value</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Treatment ($n = 136$)</td>
<td>Control ($n = 140$)</td>
</tr>
<tr>
<td>Age (y)</td>
<td>49.61 (16.71)</td>
<td>47.71 (15.11)</td>
</tr>
<tr>
<td>Sex</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Male</td>
<td>79 (52.3)</td>
<td>72 (47.7)</td>
</tr>
<tr>
<td>Female</td>
<td>57 (45.6)</td>
<td>68 (54.4)</td>
</tr>
<tr>
<td>Race</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Malays</td>
<td>123 (50.6)</td>
<td>120 (49.4)</td>
</tr>
<tr>
<td>Chinese</td>
<td>6 (40.0)</td>
<td>9 (60.0)</td>
</tr>
<tr>
<td>Indian</td>
<td>4 (36.4)</td>
<td>7 (63.6)</td>
</tr>
<tr>
<td>Others</td>
<td>3 (42.9)</td>
<td>4 (57.1)</td>
</tr>
<tr>
<td>Comorbidity</td>
<td></td>
<td></td>
</tr>
<tr>
<td>None</td>
<td>22 (44.0)</td>
<td>28 (56.0)</td>
</tr>
<tr>
<td>Recent surgical procedure</td>
<td>26 (40.6)</td>
<td>38 (59.4)</td>
</tr>
<tr>
<td>Recent trauma</td>
<td>15 (42.9)</td>
<td>20 (40.4)</td>
</tr>
<tr>
<td>Diabetes mellitus</td>
<td>62 (59.6)</td>
<td>42 (40.4)</td>
</tr>
<tr>
<td>Peripheral vascular disease</td>
<td>1 (100)</td>
<td>0</td>
</tr>
<tr>
<td>Days to complete the study</td>
<td>12.97 (10.24)</td>
<td>10.41 (9.56)</td>
</tr>
<tr>
<td>Initial wound size (cm$^2$)</td>
<td>59.54 (100.72)</td>
<td>37.17 (57.44)</td>
</tr>
</tbody>
</table>

Abbreviations: NA, not applicable; SD, standard deviation.

$^a$Independent t-test.

$^b$Chi-squared test.

### Table 2 Comparison of means of healing time (day) between variables

<table>
<thead>
<tr>
<th></th>
<th>Healing time (d) Mean (SD)</th>
<th>$p$-Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age group (y)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>&lt; 40</td>
<td>7.43 (7.72)</td>
<td>&lt;0.001$^a$</td>
</tr>
<tr>
<td>40–60</td>
<td>12.84 (10.35)</td>
<td></td>
</tr>
<tr>
<td>&gt; 60</td>
<td>14.12 (10.07)</td>
<td></td>
</tr>
<tr>
<td>Diabetes</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Yes</td>
<td>15.55 (10.26)</td>
<td>&lt;0.001$^b$</td>
</tr>
<tr>
<td>No</td>
<td>8.13 (8.25)</td>
<td></td>
</tr>
<tr>
<td>Recent surgical procedure</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Yes</td>
<td>15.55 (10.26)</td>
<td>0.721$^b$</td>
</tr>
<tr>
<td>No</td>
<td>8.13 (8.25)</td>
<td></td>
</tr>
<tr>
<td>Wound etiology</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Infection</td>
<td>12.73 (10.57)</td>
<td>0.004</td>
</tr>
<tr>
<td>Motor vehicle accidents</td>
<td>7.23 (7.64)</td>
<td></td>
</tr>
<tr>
<td>Surgical complications</td>
<td>14.00 (7.82)</td>
<td></td>
</tr>
<tr>
<td>Trauma</td>
<td>7.00 (5.41)</td>
<td></td>
</tr>
<tr>
<td>Others</td>
<td>13.67 (8.31)</td>
<td></td>
</tr>
</tbody>
</table>

Abbreviations: ANOVA, analysis of variance; SD, standard deviation.

$^a$One-way ANOVA.

$^b$Independent t-test.
Regarding the ease of removal and odor of the dressings, both groups had significant mean differences over time. The ease of dressing removal improved significantly over time, but the odor in both groups was significantly increased on day 21. At all-time points, the treatment group had significantly less pain following removal than the control group. No significant interaction time–treatment was found in any dressing assessment (Table 3).

Both groups showed a significant decrease in the mean scores regarding color, thickness, and consistency of the granulation tissue over time. No significant treatment or interaction effects were observed in any granulation tissue assessment (Table 3).

The mean scores for local and systemic complications decreased significantly for both groups over time. However, no significant treatment or interaction effects were found for complication assessments (Table 3).

**Discussion**

This study showed the effectiveness of CDHP compared with conventional CHG dressings for wound bed preparation. The use of a CDHP dressing on full-thickness wounds with cavities was supported by previous studies.\(^2\,10,28–31\) This dressing can keep the wound moist and enhance granulation tissue. This clinical trial demonstrated the effectiveness of CDHP dressings in wound bed preparation concerning granulation and removal of necrotic tissue from cavitation wounds.

However, proving that one material was superior to the other in terms of wound healing was challenging. The reason...
is that full-thickness wounds with cavities (large in size) will take a longer time for primary wound closure. Hence, this study demonstrated the use of CDHP for wound bed preparation before the secondary intention of wound closure. Almost all wounds demonstrated new granulation tissue and less necrotic tissue in the wound bed over the time points.

Although randomization of the treatment group was performed, the initial wound area was significantly larger in the CDHP group than in the control group, likely causing delayed healing. However, the difference in the baseline wound size was adjusted by conducting repeated measure analysis of covariance. Most of the wound beds were prepared and ready for the secondary intention of wound closure between days 10 and 12 in both groups.

Granulation is crucial in cavity/full-thickness wound healing. During the proliferative phase, macrophages actuate fibroblasts to release growth factors from the ECM, migrate to the wound via a fibrin–fibronectin matrix network, synthesize, and secrete the ECM. The primary composition of the initial wound matrix is fibrin, glycosaminoglycan (GAG), and hyaluronic acid. The GAG scaffold holds type I and III collagen. At this stage, the wound bed contains a dense capillary network clinically known as neoangiogenesis, giving the granulation tissue a pink, soft, and granular appearance. Hence, the wound must be covered with an effective dressing to prevent contamination and secure the process of granulation.

In our study, granulation gradually increased at the respective time points in both groups and decreased in necrotic tissue development across the time points. Thus, both dressings enhanced wound repair while providing moisture and protection to the wound via properties such as antibacterial and angiogenic properties. Another study showed that chitosan gel augmented the synthesis levels of collagen, an endogenous antioxidant, and prevented free radical-mediated tissue injury. This endogenous antioxidant may indicate that chitosan prevents the development of necrotic tissue.

Chitosan gel with a higher molecular weight and a higher degree of deacetylation accelerates wound healing. This result also suggests that chitosan accelerates the reformation of connective tissue.

This study also aims to determine the patient’s comfort and facilitate the application and removal of the dressings. Because the wounds were treated with paste and gel form dressings, leakage was observed as the paste/gel was diluted by wound exudates at the early time points of the study. The CDHP used in this trial was slightly more solid than the CHG. However, the paste was eventually liquified by exudates. Erythema was also observed during the early time points of the study because most of the wounds were from diabetic patients and might have been caused by inflammation from the prior surgical procedure. Other assessments of signs and symptoms of wound infection, such as localized warmth, pain, tenderness, edema, induration, suggested that the wounds were healthy and stable during the treatment course.

Regarding patient convenience during application and removal of the dressings, this study found that removing the CDHP was slightly more difficult than removing the CHG because it was stickier. Additionally, patients treated with CDHP experienced less pain likely because CDHP has an analgesic effect. Because infection was ruled out, the
increase in odor over time is likely attributed to the interaction of new granulation tissue, exudate, the dressing time with CDHP and CHG itself. Regardless of the dressing employed, the color, thickness, and consistency of granulation tissue exhibited a decrease in the mean score. The reason may be subjective assessment by different operators with different perceptions of the color and texture of the granulation tissues. No local or systemic problems were noted during the trial period.

Although no complications were reported, this CDHP should not be used on a patient allergic to chitosan or its derivatives, and it is not recommended for high exudate wounds or infected wounds.

**Conclusion**

This prospective randomized controlled study showed that a CDHP dressing manufactured from a deacetylated chitosan bioderivative is comparable to CHG in terms of granulation tissue formation and necrotic tissue removal. CDHP caused less pain in the patients during dressing removal. These attributes represent the positive potential of this new dressing made of natural materials.

**Authors’ Contributions**

Nur Azida Mohd Nasir was involved in writing original draft, investigation, data curation, and visualization. Ahmad Sukari Halim contributed to conceptualization, validation, investigation, resources, writing and review, supervision, project administration and acquisition. Arman Zaharil Mat Saad (USM) contributed to project administration, methodology, investigation, writing and review, supervision. Fatimah Mohd Noor (HKL and MOH) and Farrah-Hani Imran (HUKM & MOH) were involved in project administration, investigation, writing and review, and supervision. Mohd Nazri Yusof (IIUM) supervised and wrote and reviewed the manuscript. Ariff Sarifudin (IIUM) did the project administration and wrote and reviewed the manuscript. Nor Sa’adah Bachok was involved in methodology, formal analysis, and writing and review. Zanariah Ujang, Ahamd Hazri Ab Rashid, Kartini Norsal, Kamaruddin Hashim, and Norimah Yusof contributed to validation, resources, writing and review.

**Compliance with Ethics Requirements**

All procedures followed were in accordance with the ethical standards of the responsible committee on human

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**Table 3** Comparison between treatment and control groups for the assessments of wound, dressing and granulation tissue, and complications

<table>
<thead>
<tr>
<th>Assessment</th>
<th>p-Valuea</th>
<th>Treatment effect</th>
<th>Time–treatment interaction effect</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Wound</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Drainage</td>
<td>&lt; 0.001b</td>
<td>0.767</td>
<td>0.162</td>
</tr>
<tr>
<td>Itchiness</td>
<td>0.938</td>
<td>0.162</td>
<td>0.639</td>
</tr>
<tr>
<td>Erythema</td>
<td>&lt; 0.001b</td>
<td>0.365</td>
<td>0.368</td>
</tr>
<tr>
<td>Localized warmth</td>
<td>0.994</td>
<td>0.655</td>
<td>0.206</td>
</tr>
<tr>
<td>Pain/tenderness</td>
<td>0.585</td>
<td>0.162</td>
<td>0.071</td>
</tr>
<tr>
<td>Edema/induration</td>
<td>0.926</td>
<td>0.850</td>
<td>0.162</td>
</tr>
<tr>
<td><strong>Dressing</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Pain upon removal</td>
<td>0.466</td>
<td>0.019</td>
<td>0.893</td>
</tr>
<tr>
<td>Exudate</td>
<td>0.066</td>
<td>0.533</td>
<td>0.646</td>
</tr>
<tr>
<td>Adherence</td>
<td>0.936</td>
<td>0.682</td>
<td>0.843</td>
</tr>
<tr>
<td>Ease of removal</td>
<td>&lt; 0.001b</td>
<td>0.850</td>
<td>0.140</td>
</tr>
<tr>
<td>Odor</td>
<td>0.020b</td>
<td>0.231</td>
<td>0.637</td>
</tr>
<tr>
<td><strong>Granulation tissue</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Color</td>
<td>&lt; 0.001b</td>
<td>0.487</td>
<td>0.115</td>
</tr>
<tr>
<td>Thickness</td>
<td>&lt; 0.001b</td>
<td>0.263</td>
<td>0.940</td>
</tr>
<tr>
<td>Consistency</td>
<td>&lt; 0.001b</td>
<td>0.188</td>
<td>0.199</td>
</tr>
<tr>
<td><strong>Complications</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Local</td>
<td>&lt; 0.001b</td>
<td>0.354</td>
<td>0.596</td>
</tr>
<tr>
<td>Systemic</td>
<td>&lt; 0.001b</td>
<td>0.223</td>
<td>0.439</td>
</tr>
</tbody>
</table>

Abbreviation: ANCOVA, analysis of covariance.
aRepeated measure of ANCOVA.
bSignificant at \( p < 0.001 \).
cSignificant at \( p < 0.05 \).
experimentation (institutional and national) and with the Helsinki Declaration of 1975, as revised in 2008 (5). Informed consent was obtained from all patients for being included in the study. All procedures followed were in accordance with the ethical standards of the responsible committee on human experimentation from The Research Ethics Committee (Human) of Universiti Sains Malaysia (FWA Reg. No. 00007718; IRB Reg. No: 00004494) and Research Ethics Committee of Universiti Kebangsaan Malaysia (JEPUKM) (No: FF-285–2012). This study was also approved by Medical Research and Ethnic Committee, Ministry of Health Malaysia (NMRR-11-948–10565) and with the Helsinki Declaration of 1975, as revised in 2008 (5). Informed consent was obtained from all patients for being included in the study.

Clinical Trial Registration
NMRR-11–948–10565.

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Conflict of Interest
The authors declared they have no conflict of interest.

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