

Radiological Features of Joubert's Syndrome

Giovanni Stroschio¹ Caterina Cuppari² Maria Domenica Ceravolo² Annamaria Salpietro³
 Francesco Battaglia⁴ Alessia Sallemi² Monica Fusco² Antonio Ceravolo⁵ Giulia Iapadre⁶ Elisa Cali⁷
 Daniela Impollonia¹ Francesca Granata¹

¹Unit of Radiology, Department of Human Pathology in Adulthood and Childhood "G. Barresi," University Hospital of Messina, Messina, Italy

²Unit of Pediatric Emergency, Department of Human Pathology of the Adult and Developmental Age "Gaetano Barresi," University of Messina, Messina, Italy

³ASST-Spedali Civili of Brescia, Pediatrics Clinic, Brescia, Italy

⁴Orthopaedic and Traumatology Department, "S. Anna" Hospital, University of Ferrara, Ferrara, Italy

⁵Department of Pediatrics, Cinquefrondi, Reggio Calabria, Italy

⁶Department of Pediatrics, University of L'Aquila, L'Aquila, Italy

⁷Department of Neuromuscular Diseases, UCL Queen Square Institute of Neurology, London, United Kingdom

Address for correspondence Caterina Cuppari, MD, Unit of Emergency Pediatrics, Department of Human Pathology in Adulthood and Childhood "G. Barresi," University of Messina, "G. Martino" Policlinic, Via Consolare Valeria, 98124 Messina, Italy (e-mail: caterina.cuppari@polime.it).

J Pediatr Neurol 2023;21:73–77.

Abstract

Joubert syndrome (JS) is a rare autosomal recessive disorder. All patients affected by this syndrome presented a characteristic picture of cranial fossa malformations, called "molar tooth sign." This sign is defined by the presence in axial section at the level of a deck/midbrain, of hypo/dysplasia of the cerebellar vermis, abnormally deep interpeduncular fossa and horizontalized thickened and elongated superior cerebellar peduncles. Although "molar tooth sign" is peculiar of JS, other radiological findings have been also reported in these patients. Here, the authors briefly assumed the principal magnetic resonance imaging findings of JS.

Keywords

- ▶ Joubert syndrome
- ▶ radiological findings
- ▶ "molar tooth sign"

Introduction

Joubert syndrome (JS) is a rare congenital systemic disease,¹ described for the first time in 1968 by Dr. Marie Joubert in patients with agenesis of the cerebellar vermis, episodic hyperpnea, abnormal eye movements, ataxia, mental retardation, and occipital meningoencephalocele. Similar to other syndromes,^{2–5} this condition is sometimes associated with other eye abnormalities (such as retinal dystrophy, which can cause vision loss), kidney disease, liver disease, skeletal abnormalities (such as the presence of extra fingers and toes), and hormone (endocrine) problems.^{6–8} Vermian hypoplasia and abnormalities of the pontomesencephalic junction are the distinguishing features that lead to the diagnosis of JS.^{9,10}

All patients presented a characteristic picture of cranial fossa malformations, called "molar tooth sign" (MTS), which has been reported in approximately 85% patients with JS (–Fig. 1). This sign is defined by the presence in axial section at the level of a deck/midbrain, of hypo/dysplasia of the cerebellar vermis, abnormally deep interpeduncular fossa, and horizontalized thickened and elongated superior cerebellar peduncles. Since the first description, many cases were reported with expansion of the phenotype and nosological and genetic evolution.

Several disorders are more difficult to analyze because their genetic causes are often unclear, and they do not follow the patterns of inheritance. Fortunately, molecular characterization can contribute to valuable information in terms of the patient's diagnosis, prognosis, and the potential for

received

August 22, 2022

accepted after revision

October 27, 2022

article published online

January 5, 2023

© 2023, Thieme. All rights reserved.

Georg Thieme Verlag KG,

Rüdigerstraße 14,

70469 Stuttgart, Germany

DOI <https://doi.org/>

10.1055/s-0042-1760241.

ISSN 1304-2580.

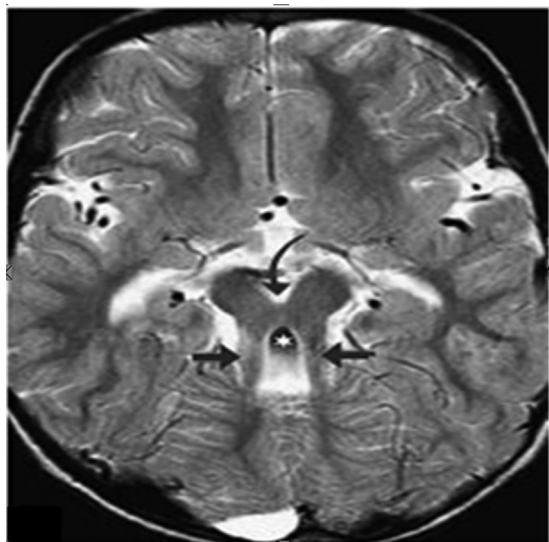


Fig. 1 The roots of this “tooth” are represented by thick and perpendicular non-decussated superior cerebellar peduncles, while the crus cerebri (cerebral peduncles) of the brain stem, with the deep interpeduncular fossa in between, represent the body of the “tooth.”

treatment with targeted therapy.¹¹ In the last decade, omic-based and experimental investigations yielded an increasing molecular and pathway complexity as causative of several pediatric intellectual disabilities and congenital brain anomalies, including ciliopathies and a wide array of neurodevelopmental disorders.^{12–20} This is observed in the context of advances brought by basic sciences in regard to the molecular understanding of many pediatric neurological conditions, both rare and ultra-rare, associated with either genetic and non-genetic etiologies.^{21–31} Several pathomechanisms have been identified, allowing a better pediatric-patient care in terms of clinical diagnosis, prognosis, and treatment.^{32–39} Pediatric developmental disorders can often result by the combination of environmental and genetic factors and their aberrant interplay may sometimes lead to disease expression and metabolic and neurological abnormalities as part of the clinical phenotype in some cases.^{40–48} These molecularly complex disorders may include a variety of monogenic as well as polygenic/genetically complex conditions with increasingly widened molecular heterogeneity and implicating

a wide array of disease mechanisms that could potentially affect brain development.^{49–56}

Although several genetic mechanisms are involved in the pathogenesis of JS,^{57–62} the abovementioned neuroradiological characteristics are exclusively reported in JS. Moreover, other syndromes with posterior-fossa malformations, including Dekaban-Arima, Senior-Löken and cerebellar vermis hypoplasia/aplasia, oligophrenia, ataxia, coloboma and hepatic fibrosis (COACH), frequently lead to diagnostic dilemmas. Here, the authors briefly assumed the principal radiological findings of JS.

The Principal MRI Findings of Joubert Syndrome

The diagnosis is based on characteristic imaging features on computed tomography (CT) or magnetic resonance imaging (MRI). The principal MRI findings of JS are: the MTS (deep interpeduncular fossa with a narrowing of isthmus, fourth ventricle deformity with hypoplasia of the vermis, thickening of the superior cerebellar peduncle, fastigium rostral shift, and sagittal vermian cleft) and the fourth ventricle batwing, resulting in distortion and dilatation of the fourth ventricle. The fastigium is shifted rostrally, and the fourth ventricle assumes a rectangular instead of a normal triangular shape, as seen on midline sagittal images (→ **Fig. 2**). Additionally, cerebellar folial disorganization, temporal lobe hypoplasia, ventriculomegaly, occipital encephalocele, atretic encephalocele, callosal dysgenesis, periventricular, subcortical heterotopia, and hypomyelination were also reported. The vermian lobules are generally dysplastic, ranging from hypoplasia to complete dysplasia. Cerebellar hemispheres, occupying most of the posterior fossa, are less involved and are nearly normal and, in axial images, are separated only by a thin cleft. On the coronal images, due to absence of the posterior vermian lobe, the cerebellar hemispheres, being divided only by a midline cleft, can be appreciated as “buttock sign” (→ **Fig. 3**). Generally, the superior cerebellar peduncles are thin (approximately 1–2 mm), dysplastic, asymmetric, and are oriented obliquely downward. In JS, the interpeduncular fossa is deep and the isthmus is variably thinner than normal and there is a lack of decussation of the superior cerebellar peduncles. Dilatation of the posterior fossa cisterns (wide prepontine cistern) was noted in almost all

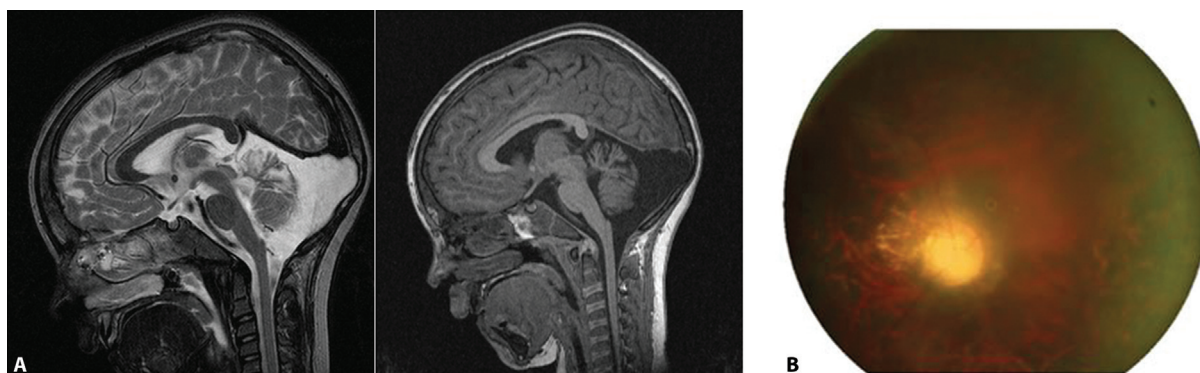


Fig. 2 (A, B) Dilatation of the fourth ventricle in midline sagittal images.

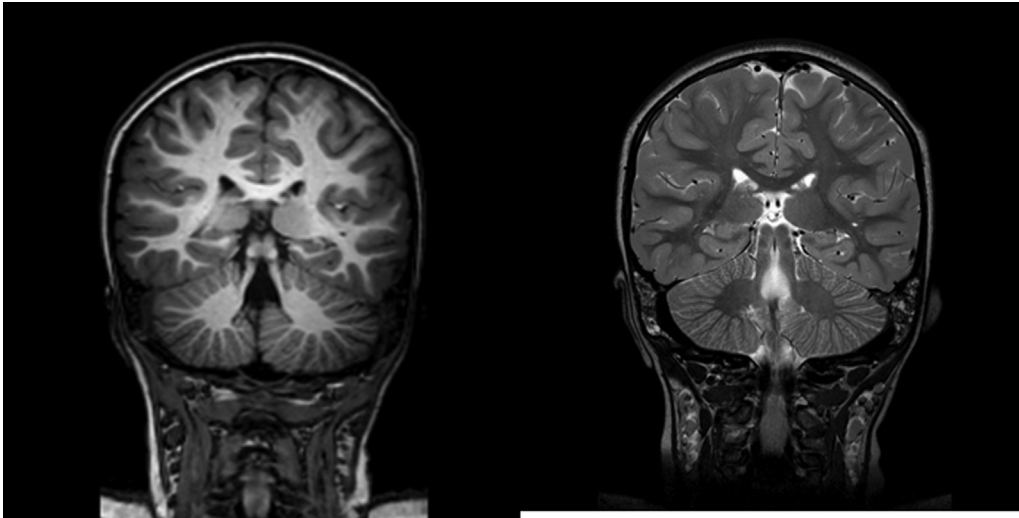


Fig. 3 The “butterfly sign” on the coronal images.

the patients with a giant cisterna magna.⁶³ Among other findings, it has also reported dysgenesis of the corpus callosum, mild to moderate cerebral atrophy with prominent ventricles, absent septum pellucidum,^{64,65} and misleading presentation of cases as pseudo-tumor cerebri.⁶⁶

The Diffusion Tensor Imaging Technique and Joubert Syndrome

Although all patients reported “MTS,” radiologic-genotype correlation in JS is a difficult task because of the low estimated prevalence. Recently, diffusion tensor imaging (DTI) technique has given an insight into the underlying complex radiological findings in JS, especially on fiber tract abnormalities including absence and/or thinning of the dorsal pontocerebellar tract, abnormal thickening of the ventral pontocerebellar tract, abnormal decussation of superior cerebellar peduncles, and finally, the absence of red dot sign.⁶⁷

The Principal Ultrasound Findings of Joubert Syndrome

The shepherd's crook sign must be also considered when evaluating patients for suspected JS. The arc of the crook is made by abnormal superior cerebellar peduncle and cerebellar hemisphere.⁶⁸

There was a shepherd's crook in sagittal views of posterior fossa where the shaft of crook is made by the brainstem and pons. By ultrasound, the shepherd's crook sign was seen through the posterior fontanelle only. CT imaging also showed the shepherd's crook sign.⁶⁹

Radiological Prenatal Findings and Joubert Syndrome

Fetal cerebral MRI can be also useful when evaluating patients for suspected JS, often in families at high risk of recurrence and/or on the ultrasound finding of abnormal posterior fossa

anatomy or the presence of associated suggestive features. However, prenatal sonographic findings in fetuses are relatively aspecific and can include increased nuchal translucency, enlarged cisterna magna, cerebellar vermian agenesis, occipital encephalocele, ventriculomegaly, hypoplastic phallus, renal cysts, and polydactyly. Authors have described in-utero visualization of the “MTS” with ultrasound⁷⁰ between 22 and 27 weeks of gestation. However, it is not possible to demonstrate vermian hypoplasia before the 18th gestational week.⁷¹ The mature normal fourth ventricle can be assessed from 18 weeks of gestation onward. However, the features of abnormal fourth ventricle are rarely described. The fourth ventricle, an ependymal cavity situated in the bulbopontine area of the brain, appeared enlarged and, in sagittal view, appears rounded, lacking the characteristic posterior “fastigial point” and with a convex roof. Moreover, due to the lack of normal decussation of cerebellar peduncles, floor of fourth ventricle is also abnormal.^{71,72}

Authors' Contribution

C.C. did the conceptualization. CC and MDC did the investigation. G.S. and E.C. contributed toward the resources. A.S., G.I., D.I., and E.G. did the data curation. E.D. and F. G. wrote and prepared the original draft. A.S. and A.C. wrote the review and edited it. M.F. and F.B. did the supervision. All authors have read and agreed to the published version of the manuscript.

Data Availability Statement

The data presented in this study are available on request from the corresponding author.

Conflict of Interest

None declared.

References

- 1 Brancati F, Dallapiccola B, Valente EM. Joubert syndrome and related disorders. *Orphanet J Rare Dis* 2010;5:20

- 2 Salpietro CD, Briuglia S, Rigoli L, Merlino MV, Dallapiccola B. Confirmation of Nablus mask-like facial syndrome. *Am J Med Genet A* 2003;121A(03):283–285
- 3 Salpietro V, Phadke R, Saggari A, et al. Zellweger syndrome and secondary mitochondrial myopathy. *Eur J Pediatr* 2015;174(04):557–563
- 4 Salpietro V, Zollo M, Vandrovicova J, et al; SYNAPS Study Group. The phenotypic and molecular spectrum of PEHO syndrome and PEHO-like disorders. *Brain* 2017;140(08):e49
- 5 Damiano Salpietro C, Briuglia S, Valeria Merlino M, Piraino B, Valenzise M, Dallapiccola B. Hallerman-Streiff syndrome: patient with decreased GH and insulin-like growth factor-1. *Am J Med Genet A* 2004;125A(02):216–218
- 6 Lacquaniti A, Chirico V, Donato V, et al. NGAL as an early biomarker of kidney disease in Joubert syndrome: three brothers compared. *Ren Fail* 2012;34(04):495–498
- 7 Travaglini L, Brancati F, Attie-Bitach T, et al; International JSRD Study Group. Expanding CEP290 mutational spectrum in ciliopathies. *Am J Med Genet A* 2009;149A(10):2173–2180
- 8 Travaglini L, Brancati F, Silhavy J, et al; International JSRD Study Group. Phenotypic spectrum and prevalence of INPP5E mutations in Joubert syndrome and related disorders. *Eur J Hum Genet* 2013;21(10):1074–1078
- 9 Valente EM, Salpietro DC, Brancati F, et al. Description, nomenclature, and mapping of a novel cerebello-renal syndrome with the molar tooth malformation. *Am J Hum Genet* 2003;73(03):663–670
- 10 Poretti A, Snow J, Summers AC, et al; NISC Comparative Sequencing Program. Joubert syndrome: neuroimaging findings in 110 patients in correlation with cognitive function and genetic cause. *J Med Genet* 2017;54(08):521–529
- 11 Molinari E, Srivastava S, Sayer JA, Ramsbottom SA. From disease modelling to personalised therapy in patients with *CEP290* mutations. *F1000 Res* 2017;6:669
- 12 Manole A, Jaunmuktane Z, Hargreaves I, et al. Clinical, pathological and functional characterization of riboflavin-responsive neuropathy. *Brain* 2017;140(11):2820–2837
- 13 Leu C, Stevelink R, Smith AW, et al; Epi25 Consortium. Polygenic burden in focal and generalized epilepsies. *Brain* 2019;142(11):3473–3481
- 14 Bruno L, Ceravolo G, Ceravolo MD, et al. Genetic cardiac channelopathies and SIDS. *J Biol Regul Homeost Agents* 2020;34(4, suppl 2):55–58
- 15 Piard J, Umanah GKE, Harms FL, et al. A homozygous *ATAD1* mutation impairs postsynaptic AMPA receptor trafficking and causes a lethal encephalopathy. *Brain* 2018;141(03):651–661
- 16 Chelban V, Wilson MP, Warman Chardon J, et al; Care4Rare Canada Consortium and the SYNAPS Study Group. PDXK mutations cause polyneuropathy responsive to pyridoxal 5'-phosphate supplementation. *Ann Neurol* 2019;86(02):225–240
- 17 Pellerin D, Ellezam B, Korathanakhun P, et al. Multisystem proteinopathy associated with a VCP G156S mutation in a French Canadian Family. *Can J Neurol Sci* 2020;47(03):412–415
- 18 Nascimben F, Peri FM, Impellizzeri P, et al. Role of oxidative stress in the pathogenesis of congenital cardiopathies. *J Biol Regul Homeost Agents* 2020;34(4, suppl 2):85–90
- 19 Nicita F, Ruggieri M, Polizzi A, et al. Seizures and epilepsy in Sotos syndrome: analysis of 19 Caucasian patients with long-term follow-up. *Epilepsia* 2012;53(06):e102–e105
- 20 Carotenuto M, Roccella M, Pisani F, et al. Polysomnographic findings in fragile X syndrome children with EEG abnormalities. *Behav Neurol* 2019;2019:5202808
- 21 Coleman J, Jouannot O, Ramakrishnan SK, et al. PRRT2 regulates synaptic fusion by directly modulating SNARE complex assembly. *Cell Rep* 2018;22(03):820–831
- 22 Salpietro V, Lin W, Delle Vedove A, et al; SYNAPS Study Group. Homozygous mutations in *VAMP1* cause a presynaptic congenital myasthenic syndrome. *Ann Neurol* 2017;81(04):597–603
- 23 Giacobbe A, Granese R, Grasso R, et al. Association between maternal serum high mobility group box 1 levels and pregnancy complicated by gestational diabetes mellitus. *Nutr Metab Cardiovasc Dis* 2016;26(05):414–418
- 24 Pavone P, Briuglia S, Falsaperla R, et al. Wide spectrum of congenital anomalies including choanal atresia, malformed extremities, and brain and spinal malformations in a girl with a de novo 5.6-Mb deletion of 13q12.11–13q12.13. *Am J Med Genet A* 2014;164A(07):1734–1743
- 25 Efthymiou S, Salpietro V, Malintan N, et al; SYNAPS Study Group. Biallelic mutations in neurofascin cause neurodevelopmental impairment and peripheral demyelination. *Brain* 2019;142(10):2948–2964
- 26 Cuppari C, Amatruda M, Ceravolo G, et al. Myocarditis in children – from infection to autoimmunity. *J Biol Regul Homeost Agents* 2020;34(4, suppl 2):37–41
- 27 Steel D, Salpietro V, Phadke R, et al. Whole exome sequencing reveals a *MLL* de novo mutation associated with mild developmental delay and without 'hairy elbows': expanding the phenotype of Wiedemann-Steiner syndrome. *J Genet* 2015;94(04):755–758
- 28 Pinchefskey EF, Accogli A, Shevell MI, Saint-Martin C, Srouf M. Developmental outcomes in children with congenital cerebellar malformations. *Dev Med Child Neurol* 2019;61(03):350–358
- 29 Toldo I, Brunello F, Morao V, et al. First attack and clinical presentation of hemiplegic migraine in pediatric age: a multicenter retrospective study and literature review. *Front Neurol* 2019;10:1079
- 30 Chimenz R, Chirico V, Cuppari C, et al. Fabry disease and kidney involvement: starting from childhood to understand the future. *Pediatr Nephrol* 2022;37(01):95–103
- 31 Greco M, Ferrara P, Farello G, Striano P, Verrotti A. Electroclinical features of epilepsy associated with 1p36 deletion syndrome: a review. *Epilepsy Res* 2018;139:92–101
- 32 Bell S, Rousseau J, Peng H, et al. Mutations in *ACTL6B* cause neurodevelopmental deficits and epilepsy and lead to loss of dendrites in human neurons. *Am J Hum Genet* 2019;104(05):815–834
- 33 Papandreou A, Schneider RB, Augustine EF, et al. Delineation of the movement disorders associated with *FOXG1* mutations. *Neurology* 2016;86(19):1794–1800
- 34 Chirico V, Rigoli L, Lacquaniti A, et al. Endocrinopathies, metabolic disorders, and iron overload in major and intermedia thalassemia: serum ferritin as diagnostic and predictive marker associated with liver and cardiac T2* MRI assessment. *Eur J Haematol* 2015;94(05):404–412
- 35 Sestito S, Roppa K, Parisi F, et al. The heart in Anderson-Fabry disease. *J Biol Regul Homeost Agents* 2020;34(4, suppl 2):63–69
- 36 Ghosh SG, Becker K, Huang H, et al. Biallelic mutations in *ADPRHL2*, encoding ADP-ribosylhydrolase 3, lead to a degenerative pediatric stress-induced epileptic ataxia syndrome. *Am J Hum Genet* 2018;103(03):431–439
- 37 Granata F, Morabito R, Mormina E, et al. 3T double inversion recovery magnetic resonance imaging: diagnostic advantages in the evaluation of cortical development anomalies. *Eur J Radiol* 2016;85(05):906–914
- 38 Chimenz R, Manti S, Fede C, et al. Primary nocturnal enuresis in children with allergic rhinitis and severe adenotonsillar hypertrophy: a single center pilot study. *J Biol Regul Homeost Agents* 2015;29(2, suppl 1):73–79
- 39 Accogli A, Iacomino M, Pinto F, et al. Novel *AMPD2* mutation in pontocerebellar hypoplasia, dysmorphisms, and teeth abnormalities. *Neurol Genet* 2017;3(05):e179
- 40 Sheldon CA, Paley GL, Xiao R, et al. Pediatric idiopathic intracranial hypertension: age, gender, and anthropometric features at diagnosis in a large, retrospective, multisite cohort. *Ophthalmology* 2016;123(11):2424–2431

- 41 Riva A, Gambadauro A, Dipasquale V, et al. Biallelic variants in *KIF17* associated with microphthalmia and coloboma spectrum. *Int J Mol Sci* 2021;22(09):4471
- 42 Ruggieri M, Polizzi A, Strano S, et al. Mixed vascular nevus syndrome: a report of four new cases and a literature review. *Quant Imaging Med Surg* 2016;6(05):515–524
- 43 Salpietro V, Ruggieri M. Pseudotumor cerebri pathophysiology: the likely role of aldosterone. *Headache* 2014;54(07):1229
- 44 Casto C, Dipasquale V, Ceravolo I, et al. Prominent and regressive brain developmental disorders associated with Nance-Horan syndrome. *Brain Sci* 2021;11(09):1150
- 45 Pavlidou E, Salpietro V, Phadke R, et al. Pontocerebellar hypoplasia type 2D and optic nerve atrophy further expand the spectrum associated with selenoprotein biosynthesis deficiency. *Eur J Paediatr Neurol* 2016;20(03):483–488
- 46 Salpietro V, Efthymiou S, Manole A, et al. A loss-of-function homozygous mutation in *DDX59* implicates a conserved DEAD-box RNA helicase in nervous system development and function. *Hum Mutat* 2018;39(02):187–192
- 47 Shaheen R, Mark P, Prevost CT, et al. Biallelic variants in *CTU2* cause DREAM-PL syndrome and impair thiolation of tRNA wobble U34. *Hum Mutat* 2019;40(11):2108–2120
- 48 Tassano E, Accogli A, Pavanello M, et al. Interstitial 9p24.3 deletion involving only *DOCK8* and *KANK1* genes in two patients with non-overlapping phenotypic traits. *Eur J Med Genet* 2016;59(01):20–25
- 49 Pedullà M, Miraglia Del Giudice M, Fierro V, et al. Atopy as a risk factor for thyroid autoimmunity in children. *J Biol Regul Homeost Agents* 2012;26(Suppl 1):S9–S14
- 50 Niccolini F, Mencacci NE, Yousaf T, et al. *PDE10A* and *ADCY5* mutations linked to molecular and microstructural basal ganglia pathology. *Mov Disord* 2018;33(12):1961–1965
- 51 Gramaglia SMC, Cuppari C, Salpietro C, et al. Congenital heart disease in Down syndrome. *J Biol Regul Homeost Agents* 2020;34(4, suppl 2):31–35
- 52 Salpietro V, Ruggieri M, Sancetta F, et al. New insights on the relationship between pseudotumor cerebri and secondary hyperaldosteronism in children. *J Hypertens* 2012;30(03):629–630
- 53 Mitsumoto H, Turner MR, Ajroud-Driss S, et al; all Delegates of the PLS Conference. Preface: promoting research in PLS: current knowledge and future challenges. *Amyotroph Lateral Scler Frontotemporal Degener* 2020;21(Suppl 1):1–2
- 54 Ruggieri M, Polizzi A, Schepis C, et al. Cutis tricolor: a literature review and report of five new cases. *Quant Imaging Med Surg* 2016;6(05):525–534
- 55 Chirico V, Lacquaniti A, Salpietro V, Buemi M, Salpietro C, Arrigo T. Central precocious puberty: from physiopathological mechanisms to treatment. *J Biol Regul Homeost Agents* 2014;28(03):367–375
- 56 Loddo I, Cutrupi MC, Concolino D, et al. Cardiac defects in RASopathies: a review of genotype–phenotype correlations. *J Biol Regul Homeost Agents* 2020;34(4, suppl 2):23–30
- 57 Valente EM, Marsh SE, Castori M, et al. Distinguishing the four genetic causes of Joubert syndrome-related disorders. *Ann Neurol* 2005;57(04):513–519
- 58 Valente EM, Brancati F, Silhavy JL, et al; International JSRD Study Group. *AHI1* gene mutations cause specific forms of Joubert syndrome-related disorders. *Ann Neurol* 2006;59(03):527–534
- 59 Valente EM, Logan CV, Mougou-Zerelli S, et al. Mutations in *TMEM216* perturb ciliogenesis and cause Joubert, Meckel and related syndromes. *Nat Genet* 2010;42(07):619–625
- 60 Brancati F, Barrano G, Silhavy JL, et al; International JSRD Study Group. *CEP290* mutations are frequently identified in the ocular form of Joubert syndrome-related disorders. *Am J Hum Genet* 2007;81(01):104–113
- 61 Brancati F, Travaglini L, Zablocka D, et al; International JSRD Study Group. *RPGRIP1L* mutations are mainly associated with the cerebello-renal phenotype of Joubert syndrome-related disorders. *Clin Genet* 2008;74(02):164–170
- 62 Brancati F, Iannicelli M, Travaglini L, et al; International JSRD Study Group. *MKS3/TMEM67* mutations are a major cause of *COACH* Syndrome, a Joubert Syndrome related disorder with liver involvement. *Hum Mutat* 2009;30(02):E432–E442
- 63 Briguglio M, Pinelli L, Giordano L, et al; CBCD Study Group. Pontine tegmental cap dysplasia: developmental and cognitive outcome in three adolescent patients. *Orphanet J Rare Dis* 2011;6:36
- 64 Arrigoni F, Romaniello R, Peruzzo D, et al. Anterior mesencephalic cap dysplasia: novel brain stem malformative features associated with Joubert syndrome. *AJNR Am J Neuroradiol* 2017;38(12):2385–2390
- 65 Enokizono M, Aida N, Niwa T, et al. Neuroimaging findings in Joubert syndrome with *C5orf42* gene mutations: a milder form of molar tooth sign and Vermian hypoplasia. *J Neurol Sci* 2017;376:7–12
- 66 Seylanian Toosi F, Boloursaz S, Abbasi B, Hekmat R, Mortazavi Ardestani R, Mohajerzadeh M. Joubert syndrome; misleading presentation of two cases as pseudo-tumor cerebri and literature review. *J Renal Inj Prev* 2016;6(02):76–79
- 67 Hsu CC, Kwan GN, Bhuta S. High-resolution diffusion tensor imaging and tractography in Joubert syndrome: beyond molar tooth sign. *Pediatr Neurol* 2015;53(01):47–52
- 68 Accogli A, Addour-Boudrahem N, Srouf M. Neurogenesis, neuronal migration, and axon guidance. *Handb Clin Neurol* 2020;173:25–42
- 69 Manley AT, Maertens PM. The Shepherd's Crook sign: a new neuroimaging pareidolia in Joubert syndrome. *J Neuroimaging* 2015;25(03):510–512
- 70 Pugash D, Oh T, Godwin K, et al. Sonographic 'molar tooth' sign in the diagnosis of Joubert syndrome. *Ultrasound Obstet Gynecol* 2011;38(05):598–602
- 71 Quarello E, Molho M, Garel C, et al. Prenatal abnormal features of the fourth ventricle in Joubert syndrome and related disorders. *Ultrasound Obstet Gynecol* 2014;43(02):227–232
- 72 Accogli A, Addour-Boudrahem N, Srouf M. Diagnostic approach to cerebellar hypoplasia. *Cerebellum* 2021;20(04):631–658