The Value of Partial HPV Genotyping After Conization of Cervical Dysplasias

Nutzen der partiellen HPV-Genotypisierung nach Konisation zervikaler Dysplasien

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ABSTRACT
Introduction In this retrospective study partial genotyping of human papilloma viruses (HPV) using the Abbott RealTime HighRisk HPV Test (RealTime) was compared with simple HPV detection (Qiagen Hybrid Capture 2 Test; hc2) for recurrence prediction at the first follow-up examination after conization of cervical intraepithelial neoplasia (CIN).

Methods 144 women who had undergone conization for CIN between January 2007 and December 2013 were included. HPV status was determined preoperatively and at first follow-up using hc2 in 103 women and RealTime in 41 women. Recurrent or persistent CIN was assumed when CIN2+ was confirmed histologically or on comparable cytology findings.

Results Of the 144 women with complete data 12 (8.3%) had a recurrence after conization. HPV persistence at follow-up correlated significantly with recurrence (hc2: p = 0.003; RealTime: p = 0.003) and both sensitivity and specificity were high (hc2 = 100 and 78.4% respectively; RealTime = 75.0 and 83.9%). Whereas isolated HPV testing had a relatively low positive predictive value for recurrence (hc2 16%; RealTime 54.5%), this rose to 80% with HPV 16 detection at follow-up.

Conclusion At follow-up after conization of CIN the combination of high risk HPV detection and partial genotyping of HPV 16 constitutes excellent diagnostic criteria for recurrence/persistence of CIN.

ZUSAMMENFASSUNG
Einleitung In dieser retrospektiven Studie wurde die partielle Genotypisierung humaner Papillomviren (HPV) mithilfe des Abbott RealTime HighRisk HPV Tests (RealTime) verglichen mit dem einfachen HPV-Nachweis (Qiagen Hybrid Capture 2 Test; hc2) zur Vorhersage eines Rezidivs in der 1. Follow-up-Untersuchung nach Konisation bei zervikalen intraepithelialen Neoplasien (CIN).


Ergebnisse Von 144 Frauen mit vollständigen Daten hatten 12 (8.3%) ein Rezidiv nach Konisation. Eine HPV-Persistenz im Follow-up korrelierte signifikant mit einem Rezidiv (hc2: p = 0.003 bzw. RealTime: p = 0.003) bei hoher Sensitivität bzw. Spezifität (hc2 = 100 bzw. 78.4%; RealTime = 75,0 bzw. 83.9%). Während der positiv prädiktive Wert der alleinigen HPV-Testung für ein Rezidiv relativ niedrig lag (hc2 16%; RealTime 54,5%), stieg dieser bei HPV-16-Nachweis im Follow-up auf 80% an.
Introduction

High-grade cervical intraepithelial neoplasia (CIN) and cervical carcinoma (CC) are associated with high-risk type human papilloma virus (hr-HPV) infection in over 99% of cases. The International Agency for Research on Cancer (IARC) classifies 12 (16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58 and 59) of the 150 HPV types as class one carcinogens. Genotypes 16 and 18 are among the most common worldwide and are found in over 70% of squamous cell carcinomas (85% of CC) and in over 80% of adenocarcinomas and adenosquamous carcinomas (15% of CC) [1–4].

The precancerous lesions CIN and adenocarcinoma in situ (AIS) can be effectively treated with the various available surgical interventions and laser ablation (ectocervical CIN) [5–7]. Loop excision is the most commonly used method, which, in contrast to e.g. laser ablation, allows acquisition of a cone biopsy for histological analysis and assessment of endo- and ectocervical resection margins [8, 9].

Despite surgical intervention residual findings and recurrences occur in 5 to 25% of cases following conization of a CIN-3 lesion. This is attributed to non-in-sano resection or persistent and newly acquired HPV infections [10].

For this reason the follow-up examination is essential and detection methods with high sensitivity and specificity for recurrence are required. The American Society for Colposcopy and Cervical Pathology (ASCCP) currently recommends combined cytology and HPV status testing (so-called co-testing) 12 and 24 months after conization for CIN 2+ [11]. According to the current German 53 guideline (consultation version) co-testing should be performed at 6, 12 and 24 months after conization. Should this co-testing be negative on all three occasions the return to routine screening is recommended. In the presence of co-testing positivity, however, differentiated colposcopy should be performed [12].

In recent decades the Hybrid Capture 2 HPV DNA Test (hc2; Qiagen, Hilden, Germany) was the most commonly used HPV test in clinical practice worldwide and in studies was most often the primary method for hr-HPV detection [13–15]. In addition, the hc2 is the standard against which new HPV test methods are measured; to be regarded as suitable tests non-inferiority to hc2 in terms of clinical sensitivity and specificity must be demonstrated [15, 16].

Such non-inferiority has been shown for the Abbott RealTime High Risk HPV Test (RealTime; Abbott, Wiesbaden, Germany), a PCR-based HPV detection method, in studies by Poljak et al. [17] and Carozzi et al. [18].

The aim of this retrospective study was to evaluate both above-mentioned test methods with respect to prediction of CIN recurrence after conization. In this context, using RealTime, we tested the partial course of HPV 16 infection, i.e. persistence, in particular as a predictor of recurrence. We also considered the recurrence prediction potential of co-testing of cytology and course of HPV infection.

Materials and Methods

Study population

Between January 2007 and December 2013 a total of 717 patients underwent conization at the Department of Gynaecology and Obstetrics at the Hannover Medical School, mostly in the presence of known CIN, AIS or microinvasive cervical carcinoma (also see [19]). Most of these patients were examined preoperatively in the dysplasia clinic. Many were also referred for follow-up. At colposcopy a fresh cytology swab (mostly fluid cytology; ThinPrep, Hologic, Wiesbaden) in combination with an HPV test were always performed. In 2011 the hospital's HPV test changed from hc2 to RealTime, thus forming two patient groups for comparison with one another. Since this study aimed to test the validity of pre- and postoperative testing with the same HPV test method, 573 patients (79.9%) could not be included (Fig. 1).

Data acquisition

The data were collected from patient clinical histories, examination findings as well as from surgical and pathology reports. Cytology reporting was according to the Munich nomenclature II since this was valid until 12/2013. For this study the Bethesda classification was also applied with the formation of two groups (≤ low grade squamous intraepithelial lesions ≤ LSIL) or high grade squamous intraepithelial lesions ≥ HSIL). Histology reporting was according to the CIN classification (CIN1–3, AIS, cervical carcinoma).

Persistence or recurrence was defined as histologically confirmed CIN2+ and/or cytology findings of ≥ HSIL during follow-up, the higher grade results always being regarded as relevant. In cases with no histology results, where recurrence was suspected on cytology the documented clinical course was scrutinised to determine whether progressive higher grade dysplasia developed and if so, the case was classified as a recurrence. If not, and in the presence of normalised results, these initially suspicious cytology results were not regarded as recurrences.

HPV genotyping was performed using RealTime in 41 patients (28.5%) and the remaining 103 patients (71.5%) were tested for HPV positivity using hc2. The predictive power for recurrence of the three factors HPV status (hc2 vs. RealTime), HPV persistence (hc2 vs. RealTime) and partial HPV genotyping at follow-up was analysed by forming two cohorts in each case, each with applicable criteria. E.g. for HPV 16 persistence, only cases shown to be HPV 16 positive before conization and at follow-up approx. six months later were analysed. The test quality was then calculated for each cohort.
HPV test methods

RealTime is a qualitative multiplex test based on real-time PCR used for the detection of 14 hr-HPV genotypes (16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 66, and 68) with simultaneous genotyping of HPV type 16 and 18. Hc2 is an in vitro nucleic acid hybridising assay for qualitative detection of 13 hr-HPV genotypes (16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, and 68). The tests were performed according to the manufacturers recommendations (detailed description in [20]).

Apart from HPV status and cytology, cone height (categorisation according to Kliemann et al. [21]) and weight (categorisation into two similar sized subgroups), resection margins, endocervical re-resection, menopause status and the transformation zone were also analysed in terms of correlation with recurrence.

Data were collected and analysed using Excel version 2007 (Microsoft) and IBM SPSS Statistics 22 (IBM Corporation, Armonk, NY, USA). Significance testing was performed using Fisher’s exact test and p-values of < 0.05 were regarded as significant.

Results

Patient data

The average age of the 144 patients included in the study was 36.1 ± 8.7 years (range: 21.8–68.5), 90.3% was premenopausal. Cone histology is shown in Table 1. Both cases of invasive carcinoma were microinvasive carcinomas that were only diagnosed at conization. Both cases did not require further surgery and there was no evidence of recurrence at follow-up.

The first follow-up examination was performed after 6.4 months ± 2.8 (range: 2.8–17.2), 123 patients (85.4%) having an “unremarkable result” (histology ≤ CIN 1 or cytology ≤ LSIL). In a further nine patients with only abnormal cytology at 1st follow-up recurrence was subsequently excluded so that overall 132 patients (91.7%) without recurrence were included in the analysis. In contrast a total of 12 patients were diagnosed with recurrence or persistence (8.3%) defined as CIN2+ at 1st follow-up (7 patients, 4.9%) and/or initial abnormal cytology (≥ HSIL) and subsequent histologically proven new high-grade dysplasia (5 patients, 3.5%). There were 4 recurrences in the hc2 group (4/103; 3.9%) and 8 in the RealTime group (8/41; 19.5%).

Predictors of recurrence at first follow-up examination after conization

140 patients (97.2%) had positive HPV results preoperatively whereas, following surgical intervention, 36 women (25.0%) still tested positive for HPV infection and 104 (72.2%) tested negative. Four patients (2.8%) were negative for HPV infection both pre- and postoperatively. Of the 41 patients (28.5%) who were tested with RealTime pre- and postoperatively multiple infections were detected in 9 cases (22.0%) (Table 2). Test quality criteria for the different HPV constellations are shown in Table 3.

HPV testing at first follow-up using RealTime produced false negative results in two cases with recurrence. In one of these cases the recurrence was primarily identified by pathological cytology only. CIN2 (p16 immunohistochemistry negative) was later
confirmed. The second HPV negative recurrence was detected at follow-up and confirmed as AIS at biopsy after previous HPV positive CIN3.

Positive co-testing was defined as either positive HPV results, cytology results of ≥ LSIL or a combination of both parameters. Positive co-testing with both hc2 + cytology and with RealTime + cytology correlated significantly with recurrence recognition (▶ Table 4).

Following conization cone heights of < 11 mm, < 13 mm, < 20 mm and cone weight of ≤ 1 g did not correlate with increased recurrence risk when compared to larger or heavier cone biopsies (cone height 11 mm: p = 0.732; 13 mm: p = 1.000; 20 mm: p = 1.000; cone weight p = 1.000). The transformation zone, determined during preoperative testing and at follow-up, was also not significant for the prediction of recurrence (preoperative: p = 0.147; follow-up: p = 0.702).

In 65.3% of cases (94 patients) an endocervical re-resection was performed in addition to conization (7/94 recurrences; 7.4%), the remaining cases having conization alone (4/49 recurrences; 8.2%). In this study re-resection was not shown to provide any additional benefit with respect to recurrence reduction (p = 1.000).

### Table 2 The course of multiple HPV infections detected using RealTime (n = 41).

<table>
<thead>
<tr>
<th>HPV genotypes</th>
<th>Preoperative</th>
<th>Follow-up</th>
</tr>
</thead>
<tbody>
<tr>
<td>HPV 16</td>
<td>14</td>
<td>5</td>
</tr>
<tr>
<td>HPV 18</td>
<td>1</td>
<td>0</td>
</tr>
<tr>
<td>HPV 18 + 16</td>
<td>1</td>
<td>0</td>
</tr>
<tr>
<td>HPV 18 + non-16/18</td>
<td>1</td>
<td>0</td>
</tr>
<tr>
<td>non-16/18</td>
<td>15</td>
<td>6</td>
</tr>
<tr>
<td>non-16/18 + 16</td>
<td>7</td>
<td>0</td>
</tr>
<tr>
<td>HPV negative</td>
<td>2</td>
<td>2</td>
</tr>
</tbody>
</table>

### Table 3 HPV test methods and recurrence prediction.

<table>
<thead>
<tr>
<th>Cohort</th>
<th>n</th>
<th>Recurrence</th>
<th>Test positive</th>
<th>Test negative</th>
<th>Sens. %</th>
<th>Spec. %</th>
<th>PPV %</th>
<th>NPV %</th>
<th>p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>hc2 status (1st follow-up)</td>
<td>103</td>
<td>yes</td>
<td>4</td>
<td>0</td>
<td>100</td>
<td>78.8</td>
<td>16</td>
<td>100</td>
<td>0.003</td>
</tr>
<tr>
<td></td>
<td></td>
<td>no</td>
<td>21</td>
<td>78</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>RealTime status (1st follow-up)</td>
<td>41</td>
<td>yes</td>
<td>6</td>
<td>2</td>
<td>75</td>
<td>84.8</td>
<td>54.5</td>
<td>93.3</td>
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</tr>
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<td></td>
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<td>28</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
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<tr>
<td>hc2 persistence</td>
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<td>4</td>
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<td>100</td>
<td>78.4</td>
<td>16</td>
<td>100</td>
<td>0.003</td>
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<td>76</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>RealTime persistence</td>
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<td>yes</td>
<td>6</td>
<td>2</td>
<td>75</td>
<td>83.9</td>
<td>54.5</td>
<td>92.9</td>
<td>0.003</td>
</tr>
<tr>
<td></td>
<td></td>
<td>no</td>
<td>5</td>
<td>26</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>HPV 16 status (1st follow-up)</td>
<td>41</td>
<td>yes</td>
<td>4</td>
<td>4</td>
<td>50</td>
<td>97</td>
<td>80</td>
<td>88.9</td>
<td>0.003</td>
</tr>
<tr>
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<td></td>
<td>no</td>
<td>1</td>
<td>32</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>HPV 16 persistence</td>
<td>22</td>
<td>yes</td>
<td>4</td>
<td>0</td>
<td>100</td>
<td>94.4</td>
<td>80</td>
<td>100</td>
<td>0.001</td>
</tr>
<tr>
<td></td>
<td></td>
<td>no</td>
<td>1</td>
<td>17</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>HPV non-16/18 status (1st follow-up)</td>
<td>41</td>
<td>yes</td>
<td>2</td>
<td>6</td>
<td>25</td>
<td>87.9</td>
<td>33.3</td>
<td>82.9</td>
<td>0.578</td>
</tr>
<tr>
<td></td>
<td></td>
<td>no</td>
<td>4</td>
<td>29</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>HPV non-16/18 persistence</td>
<td>23</td>
<td>yes</td>
<td>2</td>
<td>3</td>
<td>40</td>
<td>88.9</td>
<td>50</td>
<td>84.2</td>
<td>0.194</td>
</tr>
<tr>
<td></td>
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<td>no</td>
<td>2</td>
<td>16</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

n: number; hc2: Hybrid Capture 2 Assay; RealTime: Abbott RealTime Assay; PPV: positive predictive value; NPV: negative predictive value

![Fig. 2 Sensitivity and specificity of postoperative HPV status for recurrence prediction, n = 144, values in percent (%).](image-url)
The predictive value of cone biopsy resection margin status was also evaluated. 19 cases (13.2%) were excluded due to uncertain results. R1 status, i.e. non-in sano resection, was present in 13 cases (9%). Five of these 13 patients (38.5%) had a recurrence whereas six of the 112 patients (5.4%) with R0 histology had a recurrence (p = 0.002; ▶ Table 5).

Discussion

HPV status

Determination of HPV status forms part of the currently recommended standard follow-up after conization of high-grade cervical dysplasia (simple hr-HPV detection without genotyping) [11, 12]. The two test methods evaluated in our study, hc2 and RealTime, were shown to have clinical sensitivity and specificity of 100 and 78.8% (hc2)/specificity 84.8% for RealTime respectively. These are predictive values for recurrence that correlate well with other comparable studies (hc2: sensitivity 97.4% [94.7–100%], specificity 87.9% [83.3–92.6%], RealTime: specificity 89.4% [86.4–92.3%] [17, 18, 22]). Similar to our findings these studies also found a tendency towards higher specificity for RealTime compared to hc2 and higher clinical sensitivity for hc2.

Our findings differ from the above-mentioned studies in terms of a lower sensitivity for RealTime (75.0 vs. 96.7% [95.5–98.2%]), 2/8 patients (25.0%) with recurrence had negative RealTime HPV tests at follow-up. However with only eight cases of recurrence in the RealTime group this result is of limited validity. It is also questionable whether the two cases were indeed “actual recurrences”. In one case CIN2 was diagnosed at follow-up that was negative for p16 on immunohistochemistry and was thus probably not caused by hr-HPV but possibly by low-risk HPV. In the second case with a negative RealTime HPV test the follow-up biopsy (after previous CIN3) confirmed AIS. Subsequent re-conization was however unremarkable/normal. Both cases were nevertheless treated as recurrences in view of the original inclusion criteria defined at the start of the study. Without these two cases the sensitivity of RealTime would also have been 100%. A degree of selection bias can therefore not be excluded.

The significantly higher recurrence rate in the RealTime group (19.5%; 8/41 cases) compared to the hc2 collective (3.9%; 4/103 cases) can also be explained by these two specific cases and to some extent also by the small total case number.

A further limitation of the study is the relatively small case number overall with different sized groups for testing with the two methods (hc2 103 patients, RealTime 41 patients). HPV 16 could thus only be evaluated in 22 patients. This is due to the fact that only patients tested preoperatively and at follow-up with the same HPV testing method were included.

A study by Kaliterna et al. [23] that included 108 patients – a similar sized study population to the 103 patients tested with hc2 in our study – found a similar sensitivity and specificity for this HPV test (Kaliterna: 88.2 and 79% vs. hc2: 100 and 78.8%). Strand et al. in their study [24] note the potentially good predictive value of HPV genotyping for the presence of CIN2+ recurrence. In our study we evaluated the course of HPV infection using hc2 and RealTime. We initially pooled RealTime results according to HPV positivity or negativity without performing a genotype specific analysis. These results showed no recurrence prediction advantage in terms of sensitivity and specificity for the observation of the course of HPV infection using hc2 and RealTime (pooled) (hc2: sensitivity 100%, specificity 78.4%; RealTime: sensitivity 75.0%, specificity 83.9%) over determination of HPV status only at first follow-up after conization (hc2: sensitivity 100%, specificity 78.8%; RealTime: sensitivity 75.0%, specificity 84.8%) (▶ Fig. 2).

This was also not to be expected, since for both test methods (pooled HPV status) the numbers of HPV persistence matched those of HPV positive results at 1st follow-up. Likewise the num-

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**Table 4** Co-testing methods and recurrence recognition.

<table>
<thead>
<tr>
<th></th>
<th>n</th>
<th>Recurrence</th>
<th>Test positive</th>
<th>Test negative</th>
<th>Sens. %</th>
<th>Spec. %</th>
<th>PPV %</th>
<th>NPV %</th>
<th>p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cytology + hc2</td>
<td>103</td>
<td>yes</td>
<td>4</td>
<td>0</td>
<td>100</td>
<td>78.8</td>
<td>16</td>
<td>100</td>
<td>0.003</td>
</tr>
<tr>
<td></td>
<td></td>
<td>no</td>
<td>21</td>
<td>78</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Cytology + RealTime</td>
<td>41</td>
<td>yes</td>
<td>8</td>
<td>0</td>
<td>100</td>
<td>78.8</td>
<td>53.3</td>
<td>100</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td></td>
<td></td>
<td>no</td>
<td>7</td>
<td>26</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Cytology + HPV 16 persistence</td>
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<td>yes</td>
<td>4</td>
<td>0</td>
<td>100</td>
<td>88.9</td>
<td>66.7</td>
<td>100</td>
<td>0.002</td>
</tr>
<tr>
<td></td>
<td></td>
<td>no</td>
<td>2</td>
<td>16</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

PPV: positive predictive value; NPV: negative predictive value; hc2: Hybrid Capture 2 Assay; RealTime: Abbott RealTime assay; HPV: human papilloma virus

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**Table 5** Resection margin status for prediction of recurrence.

<table>
<thead>
<tr>
<th></th>
<th>R0</th>
<th>R1</th>
<th>Total</th>
<th>p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>No residual/recurrence</td>
<td>106</td>
<td>8</td>
<td>114</td>
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<tr>
<td>Residual/occurrence</td>
<td>6</td>
<td>5</td>
<td>11</td>
<td>0.002</td>
</tr>
<tr>
<td>Total</td>
<td>112</td>
<td>13</td>
<td>125</td>
<td></td>
</tr>
</tbody>
</table>

Recurrence rate 5.4% 38.5% 8.8%

PPV: positive predictive value; NPV: negative predictive value
ber of HPV eradictions corresponded to that of HPV negative results after conization.

Genotyping

RealTime enables the evaluation of the course of genotype specific infections with HPV 16 and 18. Independent of preoperative findings, exclusive testing of HPV 16 postoperatively had a sensitivity and specificity of 50.0 and 97.0% respectively for the detection of CIN2+. Testing for this specific HPV type achieved the highest specificity of all parameters assessed in this study (1st follow-up: HPV 16 97.0%, hc2 78.8%, RealTime – total 84.8%, non-16/18 hr-HPV 87.9%). The specificity of HPV genotyping was also clearly higher than non-specific hr-HPV detection in a study by Heymans et al. [25].

We found a significantly lower sensitivity for CIN2+ at follow-up (50.0%) with exclusive testing for HPV 16 (hc2 100%, RealTime pooled 75.0%). Reasons for this are again the above-mentioned two HPV negative recurrences and two cases with recurrence due to non-16/18 hr-HPV genotypes and negative HPV 16 results.

4/5 patients with HPV 16 persistence had a recurrence (PPV 80.0%). This result is similar to those of a Norwegian study by Vintermyr et al. [26] in which 94.8% of patients with recurrence were shown to have persistent hr-HPV infection, of which 70.9% were genotypes 16/18. This study also described no change in HPV 16 prevalence before and after conization, a finding not confirmed by our study. Only five of the initial 22 positive patients (22.7%) still tested positive for HPV 16 infection postoperatively (Table 5).

These results confirm that partial HPV genotyping is a promising option. The PPV of 80% for HPV 16 persistence allows relatively reliable identification of women in imminent danger of recurrence. In contrast to this, the PPV of exclusive hc2 testing at follow-up is only around 16%.

Exclusive use of non-16/18 hr-HPV genotypes for testing at first follow-up and diagnosis of persistence after conization are, with sensitivities of 25.0 and 40.0% respectively, not meaningful parameters for post-conization monitoring. It is possible that identification of type-specific persistence with certain non-16/18 hr-HPV genotypes could also have relatively high PPV. To answer this question, however, complete HPV genotyping would be required.

The postoperative status and course of HPV 18 infections could not be evaluated since only three patients were identified with this genotype preoperatively and in all three cases the virus was no longer detectable after conization.

Co-testing

Positive co-testing (positive HPV result and/or ≥ LSIL) with hc2 and cytology did not improve sensitivity and specificity compared to hc2 alone (n = 103; 100 and 78.8%). The correlation between positive co-testing and the incidence of recurrence remained significant (p = 0.003).

The combination of RealTime and cytology increased sensitivity to 100% for positive co-testing compared to HPV testing alone (75.0%), specificity remaining almost unchanged (n = 41; 78.8 vs. 84.8%).

Other parameters

Cone height was not significant for recurrence prediction. Recession margins were clear 92.6% of the time at a cone height of 11 mm, 94.4% at 13 mm and 100% when cone height was 20 mm. These results correspond to the findings of Kliemann et al. [21] (11 mm = 90%, 13 mm = 95%, 20 mm = 100%).

Only a limited analysis of resection margins was possible in this study since involvement (R1) was pooled with no differentiation between endo- and ectocervical involvement. 6/112 patients (5.4%) with R0 resection on histology had recurrences while 5/13 patients (38.5%) with R1 resections had recurrences (p = 0.002).

However Johnson et al. [27], in agreement with previous studies [28, 29], showed that only involvement of the endocervical resection margin can be regarded as predictive of recurrence.

Endocervical re-resection was not significant for the incidence of recurrence (p = 1.000). Of the 94 patients (65.7%) who underwent re-resection 7 (7.4%) had recurrence. Among the 49 patients (34.3%) with no re-resection at conization 4 (8.2%) had recurrence, a similar proportion to those who underwent re-resection. Our results support the supposition that endocervical re-resection and the associated additional shortening of the cervix can potentially be avoided.

Conclusion

Postoperative HPV status determined using the two test methods Hybrid Capture 2 and RealTime was shown to be highly specific for non-recurrence. Whereas the sensitivity of pooled HPV results for recurrence detection is very high, the PPV (16%) of hc2 is extremely low. Here partial HPV genotyping has clear advantages. With a PPV of 80% recurrence should be expected especially in the context of (recurrent) detection of HPV 16 after conization.

Thus the combination of pooled HPV testing (high sensitivity and NPV) and partial genotyping of HPV 16 (high PPV) can simplify follow-up management after conization.

Further prospective studies with systematic biopsy after conization are required for the further evaluation of the promising genotype specific monitoring of HPV 16 infection course and to compare this to pooled HPV testing.

Conflict of Interest

P. Hillemanns received research grants from GSK and Abbott as well as presentation remuneration from SMMD, Roche and Hologic. M. Jentschke received presentation remuneration, travel and congress support from Abbott Molecular, Wiesbaden.

References

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