New Pharmacological Opportunities for Betulinic Acid

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Introduction
Betulinic acid, or 3β-hydroxy-lup-20(29)-en-28-oic acid (▶ Fig. 1), is a naturally occurring pentacyclic lupane-type triterpenoid usually isolated from birch trees, but present in many other botanical sources. It is found in different plant organs, both as a free aglycon and as glycosyl derivatives. A wide range of pharmacological activities has been described for this triterpenoid, including antiviral and antitumor effects. In addition, several other interesting properties have been identified in the fields of immunity and metabolism, namely antidiabetic, antihyperlipidemic, and anti-inflammatory activities. Taken together, these latter three properties make betulinic acid a highly interesting prospect for treating metabolic syndrome. The present review focuses on the therapeutic potential of this agent, along with several of its semisynthetic derivatives, which could open new frontiers in the use of natural product-based medicines.
Metabolic Syndrome

Metabolic syndrome is characterized by a series of interconnected physiological, biochemical, clinical, and metabolic factors, which directly increase the risk of cardiovascular disease and type 2 diabetes mellitus. Some of these risks, such as insulin resistance, atherogenic dyslipidemia, and endothelial dysfunction, can be prevented or treated [12]. The major risk factors for developing metabolic syndrome are physical inactivity and a diet high in fats and carbohydrates. Both contribute to central obesity and insulin resistance [13], which are associated with chronic inflammation, characterized by increased production of adipocytokines such as TNF-α, IL-1, IL-6, leptin, and adiponectin [12]. For this reason, the reduction of central obesity and insulin resistance, along with the control of the proinflammatory state, are all desirable for avoiding atherogenic dyslipidemia and endothelial dysfunction in vascular inflammation. Natural products, such as betulinic acid, can modulate all these risk factors.

Antidiabetic Properties

Betulinic acid has been described as a potential antidiabetic agent of interest for the treatment of type 2 diabetes mellitus [9]. To this end, various in vitro and in vivo studies have been developed to establish the compound’s activity and mechanism of action. Several studies have described its effects on absorption and uptake of glucose [14–16], insulin resistance and insulin sensitivity [17], and endogenous glucose production and synthesis of glycogen [18–21]. In addition to these effects, a number of other relevant aspects of the compound’s potential mechanisms have been established.

The first mechanism to be described was the inhibition of α-amylase and α-glucosidase, which diminishes the hydrolysis of polysaccharides. This leads to a clear reduction of free glucose and, consequently, a decrease in its absorption. The inhibitory effects on these enzymes and the delay in glucose absorption were demonstrated both in vitro and in vivo in mice. In addition, researchers were able to establish the active concentration and doses, which differed depending on the cell lines and animals used, varying from 14 µM to 100 µM [14–16].

The second mechanism is associated with increases in glycogen synthesis. In this way, betulinic acid stimulates glucose uptake and glycogen synthesis through the AMPK- GSK-3β pathway. In fact, betulinic acid (10 µM) was shown to activate AMPK, thereby decreasing the expression of phosphoenolpyruvate carboxykinase and glucose-6-phosphatase [20]. Moreover, AMPK activation increases both glucose transport across the cell membrane and the expression of GLUT-1 and GLUT-2 [20, 21].

Some relevant negative effects of type 2 diabetes mellitus are due to insulin resistance and β-cell dysfunction. These problems are thus a high priority in the treatment of the disease. In this context, Castro et al. [17] studied the effect of betulinic acid on intracellular signal transduction in glucose homeostasis and observed that rats treated with this triterpene (10 mg/kg, orally) produced a sustained insulin secretion and exhibited reduced levels of glycemia, with values close to those found in an euglycemic state.

Because different enzymes and pathways are implicated in insulin biosynthesis, secretion, and sensitivity, the PTP1B and PI3K-B–Akt pathways constitute a good target for studying the effects and mechanisms of betulinic acid [17]. Indeed, in vitro, this triterpene (0.70 µg/mL) reduced human recombinant PTP1B activity by 95% [22], with an IC50 ~ 3.5 µM [23]. In vivo, treatment of hyperglycemic rats with betulinic acid (10 mg/kg, p.o.) increased the glycogen content and glucose uptake in muscles by acting as an insulin secretagogue and insulin mimetic agent via PI3K, mitogen-activated protein kinase, and mRNA translation [17]. It is well known that protein PTP1B inhibits PI3K/Akt signaling, which leads to the appearance of insulin resistance. In contrast, the inhibition of PTP1B increases the activity of the PI3K/Akt pathway, increasing glucose uptake into skeletal muscle both through the translocation of GLUT-4 to the plasma membrane and by inhibiting gluconeogenesis [9].

The bile acid membrane receptor TGR5 is a G protein-coupled receptor that is normally expressed in brown adipose tissue and muscle. It acts as a cell surface receptor for bile acids, but is also implicated in energy homeostasis and insulin resistance [24]. Betulinic acid has TGR5 agonist activity with an EC50 ~ 1.04 µM and an efficacy of 83% [25]. The activation of the TGR5 receptor induces glucagon-like peptide-1 secretion in different enteroendocrine cells and contributes to the effects of bile acids in glucose homeostasis, but other mechanisms may also be involved, such as the stimulation of oxidative phosphorylation [26, 27]. The principal effects of betulinic acid on glucose metabolism are given in ▶ Fig. 2.

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Effects on Dyslipidemia

Betulinic acid (50 mg/L, administered in drinking water for 15 days) decreased total cholesterol and triglycerides in high-fat diet-induced obese mice [9]. The triterpene also decreased body weight, abdominal fat accumulation, blood glucose, plasma triacylglycerides, and total cholesterol. The results showed significant increases of insulin and leptin (anorexigenic hormone) in plasma, whereas the level of ghrelin (orexigenic hormone) decreased [15]. These effects explain the reduced appetite regulation observed in high-fat diet-induced obese mice [9]. Betulinic acid was shown to cause a greater decrease in plasma amylose activity than in that of lipase [15], but the results of studies conducted by Jang et al. [28] and Kim et al. [10] indicated that betulinic acid inhibits porcine pancreatic lipase (IC₅₀ = 21.1 µM) and has a lipolytic effect mediated by cAMP-dependent phosphodiesterase inhibition, which could lead to the observed reduction of lipid absorption in the small intestine and the increase of fat mobilization [10]. Betulinic acid also reduces cholesterol absorption in the intestine via inhibition of hACAT. Indeed, the triterpene had an inhibitory effect on hACAT-1 (responsible for foam cell formation in macrophages) and hACAT-2 (responsible for the cholesterol absorption process in intestinal mucosal cells), with IC₅₀ values of 16.2 and 28.8 µM, respectively [29].

In addition to its lipolytic effect in adipose tissues [10], betulinic acid reduced lipogenesis and lipid accumulation in various experimental models, both in vitro (HepG2 cells) and in vivo (ICR mice and Wistar rats) [30]. In the HepG2 cell assays, the following effects were demonstrated: suppression of intracellular lipid accumulation via modulation of lipogenic and lipolytic factors, inhibition of hepatic lipid accumulation via activation of the AMPK signaling pathway, increase of CAMKK expression, downregulation of the mTOR, and protein expression of S6 kinase. Ex vivo and in vivo assays showed the following effects: inhibition of SREBP1 activity and expression via modulation of a CAMKK–AMPK–mTOR–S6 kinase pathway in primary rat hepatocytes, suppression of hepatic triglyceride accumulation via modulation of a CAMKK–AMPK–SREBP1 signaling pathway in the livers of ICR mice (high-fat diet), and suppression of triglyceridemia in ICR mice fed with a high-fat diet [30]. Activation of AMPK suppresses mRNA expression and nuclear translocation of SREBP-1 while positively regulating fatty acid oxidation through the activation of the PPAR-α and the PPARγ coactivator (PGC)-1. These effects make sense because human PPARs is expressed in several metabolically active tissues and plays a critical role in the regulation of cellular uptake, activation, and oxidation of fatty acids [31]. For its part, PGC-1 is expressed at high levels in both heart and skeletal muscle. Its expression decreases in situations associated with mitochondrial dysfunction, such as diabetes, suggesting that decreases in PGC-1 activity may contribute to this pathology [32]. In this context, betulinic acid may be able to regulate lipid homeostasis. Betulinic acid also inhibited diacylglycerol acyltransferase in rat liver microsomes (IC₅₀ = 9.6 µM) in a noncompetitive manner, while also inhibiting triacylglycerol synthesis in HepG2 cells [33].

Taken together, these results indicate that betulinic acid may help reduce hepatic lipid accumulation by modulating the AMPK–SREBP signaling pathway, which would explain its hepatic antihyperlipidemic activity [30]. Several relevant aspects of the modification of lipid metabolism are summarized in Fig. 3.

Anti-Inflammatory Effects

Previous studies used various experimental models to demonstrate the anti-inflammatory activity of betulinic acid [11]. Indeed, this triterpene inhibited carrageenan-induced mouse paw edema by 46% (100 mg/kg), 12-O-tetradecanoylphorbol acetate-induced mouse ear edema by 86% (0.5 mg/ear), and ethyl phenylpropionate-induced mouse ear edema by 30% (0.5 mg/ear). In addition, the vascular permeability caused by serotonin (50 mg/kg) in mice was blocked by the anti-glucocorticoid used in the experiment as well as by the mRNA or protein synthesis inhibitors, indicating that the compound may be a potential corticoid-like agent [2]. In complementary research, the same authors [34] studied the effects of betulinic acid on the inflammation induced by various activators of protein kinase C and other agents. The results
showed that betulinic acid (0.5 mg/ear) inhibited the edema induced by mezerein, 12-deoxyphorbol-13-tetradecanoate, and 12-deoxyphorbol-13-phenylacetate by 48, 51, and 61% (ID50 = 0.77 µmol/ear in this case), respectively. Other positive effects were observed in bryostatin 1-induced mouse ear edema (65% at 0.5 mg/ear), bradycin-induced mouse paw edema (54% at 10 mg/kg), and rat skin inflammation induced by glucose oxidase (39% at 0.25 mg/site). No effects were observed in ear edema induced by arachidonic acid, resorufin, or xylene. Because betulinic acid was inactive against arachidonic acid-induced inflammation as well as in neurogenic inflammatory models, it is likely that this type of inflammation may depend on in vivo inhibition of protein kinase C. This possible target was previously reported by Wang and Polya [35] in vitro against rat liver cyclic AMP-dependent protein kinase, wheat embryo Ca2+-dependent protein kinase, and rat brain protein kinase C, with IC50 values of 45, 84, and 145 µM, respectively.

In other studies, betulinic acid also inhibited bovine pancreatic PLA2 by 40% (5 µM) [36], bovine prostaglandin synthase (IC50 = 101 µM), and proinflammatory cytokine-induced neutrophil chemoattractant-1 (34%, 1 µM) in stimulated rat macrophages and IL-1β-stimulated rat fibroblast cells [37]. The triterpene (30 µM) also suppressed NF-κB activation induced by different agents as well as the NF-κB-dependent gene expressions of cyclooxygenase-2 and matrix metalloproteinase-9 [38]. It selectively inhibited cyclooxygenase-2 (IC50 = 11.4 µM) over cyclooxygenase-1 (IC50 = 115 µM) [39], and moderately reduced the production of nitric oxide in stimulated RAW 264.7 cells [40].

Moreover, of the compounds assayed, betulinic acid was the best inhibitor of PLA2. This result is in agreement with previous reports describing the importance of a carboxylate group for the inhibition of PLA2 [36] as well as with the results of previous triterpene studies [41–43].

Betulinic acid can also prevent vascular inflammation and atherosclerosis. Indeed, this triterpene (0.01–10.0 µM) blocked the TNF-α-induced expression levels of intracellular adhesion molecule-1, vascular cell adhesion molecule-1, and endothelial cell selectin, all while reducing the production of reactive oxygen species and NF-κB activation [44]. It also plays a protective role in the development of vascular inflammation and atherosclerosis via ATP-binding cassette transporter A1, the expression of which is promoted through the downregulation of miR-33s, a family of microRNA precursors. This mechanism involves the suppression of iκB phosphorylation, p65 phosphorylation, and nuclear translocation as well as the transcription of the related NF-κB-dependent gene [45].

Endocrinological and Cardiovascular Effects

As mentioned above, PTPIB is an enzyme involved in several processes such as diabetes, inflammation, and cancer. Interactions with PTPIB are thus of great pharmacological interest. The vast majority of molecules that inhibit this enzyme do so by binding to the active cationic site; however, several lupane-derived triterpenes have been described as allosteric inhibitors because they interact with a hydrophobic zone of the protein. Betulinic acid, like betulin, lupeol, and lupenone, acts through hydrophobic interactions with the residues Ala189, Leu192, Phe196, Phe280, Trp291, and Leu294 of PTPIB. In addition, it forms hydrogen bridges through its two polar, alcoholic, and acidic functions with the residues Glu276, Gln288, and Lys292. Lupeol and betulinic acid have...
also been shown to inhibit TNFα-induced PTP1B expression in mouse hypothalamic cells while also exhibiting the highest potency as inhibitors of enzyme activity, with IC₅₀ values of 5.6 and 1.5 µM, respectively [46].

Betulinic acid’s interaction with the ER has also been assessed; in fact, the acid was recently characterized as one of the antiestrogenic principles of Prunella vulgaris L. (Lamiaceae). In addition, it was shown to inhibit the genomic response to estradiol and to reduce the mRNA of growth regulation by estrogen in breast cancer 1 protein (GREB1) in MCF7 cells. Moreover, it reduced the expression of ER-α [47]. Another way that betulinic acid restricts estrogen signaling is through its effect on the Pygopus protein, the expression of which is strongly indicative of breast cancer, both at the cultured cell level and in samples of invasive ER- and ER+ tumors. Betulinic acid affects the coactivation of the estrogen-responsive gene promoter by transcription factor SP-1 (specificity protein-1), a step that entails a synergistic effect with antagonists or selective estrogen modulators [48].

These reports describe mechanisms involving genomic effects, since they correspond to classic pathways of so-called nuclear or hormonal signaling. However, betulinic acid has also been shown to exert a type of non-genomic activation of estrogen function, as described by Hohmann et al. [49]. Such an effect likely occurs upstream of the production of nitric oxide as a result of the increase in nitric oxide synthase activity in endothelial cells, an effect that was blocked by estrogen antagonist IC1182780. The authors postulated that the activation of the enzyme comes from rapid phosphorylation of the Ser1177 residue.

**Antiviral Activity**

Although several reports on the antiviral activity of betulinic acid have been published, the most recent reviews focus almost exclusively on its anti-HIV activity [3, 50]. Our survey therefore covers only the latest studies and research.

Betulinic acid acts against HIV by preventing the cleavage of the capsid-spacer peptide of the Gag protein, thereby impeding viral maturation. This causes the host cell to release virions with no infective capacity. The efficacy of betulinic acid is influenced by various polymorphisms in this protein, especially in the residues 369-371 (QVT in wild type), with one of the best known being V370A. In a study carried out by Swidorski et al. [51], various derivatives were studied, namely C-28 amides carrying a C2−C3 double bond and a benzoic moiety at C-3. The most potent inhibitors of viral expansion in MT-2 cells were the ethylene diamine derivates, including pyridin-methylamines, at C-28 (EC₅₀ = 2−7 nM). The benzoic substitution at C-3 was also present in many of the 2,3-dehydrobetulin and 2,3-dehydrobetulinic acid derivatives assayed by Liu et al. [52], which all bore the bare natural -CH₂OH or -COOH substitution, respectively. The most potent compounds (EC₅₀ < 50 nM) were those with an ortho halogen substitution in the benzoic moiety of the dehydrobetulinic and betulin derivatives, and the simple benzoic or phthalic acids of dehydrobetulin.

One of the betulinic acid derivatives with the most widely recognized antiviral activity is bevirimat (▶Fig. 5), the C-3 ester of 2,2-dimethylsuccinic acid, which was synthesized in 1996 by Kashiwada et al. [53] and which has been the subject of several previous studies. Bevirimat-related compounds lacking the carboxyl C-28, where an ethylamine chain is substituted by linear or cycled alkylamines, have been described by Urano et al. [54].

Evaluated in terms of the level of reverse transcriptase activity, the antiviral potency of all the compounds assayed was higher against wild-type than against the V370A mutant, with the exception of the N-hydroxethylpiperidine derivative. However, the fluorination of bevirimat in different positions did not improve its potency in comparison with the parent group [55].

Studies of the antiviral activity of betulinic acid, along with research on various other triterpenes, have previously demonstrated an effect against herpes viruses, notably against the most common clinical strain, HSV-1. The different effects shown against this strain by the main triterpenes from the bark of birch trees (Betula sp.) were studied by Heidary Navid et al. [56], who evaluated the exact moment of application in *in vitro* treatment. After incubating the active principle with the virus, both sensitive and acyclovir-resistant strains lost their infectivity. However, neither pretreatment of the cell nor administrations at the time of viral propagation were effective.

Among the different possibilities for increasing the efficacy of betulinic acid as an antiviral agent, the most interesting is perhaps that envisaged by Visalli et al. [57], who sought to form polar derivatives on the -COOH in 28, either directly by neutralizing the acid with choline, or indirectly through the formation of intermediacy cinemamines followed by the formation of salts of choline or benzalkonium. This type of structural modification results in increased solubility, and hence a better absorption and greater effect, as demonstrated by improvements in the potency of several compounds (IC₅₀ = 0.6–0.9 µM) in relation to betulinic acid (IC₅₀ = 1.6 mM).

Two lupanes exhibiting significant activity against various viruses were isolated from Schefflera hexaphylla (L.) Frodin (Araliaceae). Their structures were similar to that of betulinic acid, but differed in the orientation of the hydroxyl in C-3. They were thus dubbed as 3-epi-betulinic acid derivatives. One of them was a 3-sulfate conjugate and the other a 4-methyl-4-carboxylic derivative. The dicarboxylic acid had a maximum potency (IC₅₀ = 20 µg/mL) in the range of that of ribavirin, especially against the influenza A virus (H1N1, in MDCK cells) and the Coxackie virus B3 (in HEp-2 cells) [58].

In their study on the active principles of Ziziphus jujuba Mill. (Rhamnaceae), Hong et al. [59] found that betulinic acid inhibited the proliferation of the influenza A/PR/8 virus in A549 cells in a dose-dependent fashion (0.4–50 µM). This effect was subsequently tested in mice to observe its therapeutic efficacy against
the disease. Although betulinic acid was unable to reduce the weight loss caused by the infection, its activity was comparable to that of oseltamivir. From a histological point of view, the test compound reduced both the edema and the inflammation markers in the lungs.

Recent research, based on the idea that the antiviral action of vitamin C could be combined with that of betulinic acid and/or other triterpenes, has focused on the synthesis and testing of various conjugates of ascorbate. The results showed that a triazole amide derived from dibenzylascorbic acid was the most potent compound (Fig. 6), but that it was only able to manifest its antiviral effect in the process of binding the virus to the host cell. Pretreatment of the virus produced a much greater inhibitory effect (87%) than pretreatment of the cell (75%). The main mechanism of action involved in binding the influenza A/WSN/33 virus to the surface of chicken erythrocytes was the inhibition of hemagglutinin-sialic acid binding [60].

Another virus susceptible to betulinic acid is hepatitis B, which has an obvious and important health impact. The inhibition of hepatitis B replication exerted by the triterpene is based on the down-regulation of SOD2 through the dephosphorylation (Ser133) of the cAMP response element-binding transcription factor at its binding site with the SOD2 promoter. In addition, betulinic acid was also shown to facilitate the translocation of the hepatitis B virus X protein into the mitochondria of mouse hepatocytes. The antiviral activity was strictly dependent on SOD2 since overexpression of this enzyme was shown to suppress the effect [61].

Cytotoxic and Potential Anticancer Properties

Betulinic acid’s antitumor activity is one of the most widely studied aspects of the compound’s pharmacology. Although its effect extends to different cell types, it seems clear that the main mechanisms involved are the stimulation of apoptosis and the inhibition of kinases, both accompanied by a prominent antioxidant effect. Due to the sheer number of reviews on the cytotoxic and anticancer properties of betulinic acid and its derivatives, we have decided to limit our review to the most recent papers that were not included in the review published by Ali-Seyed et al. [62].

Betulinic acid was found to reduce many of the toxicity indicators of the antitumor agent doxorubicin, which is known to have strong cardiotoxic activity. These parameters, measured in human blood lymphocytes, include generation of reactive oxygen species, production of inflammatory cytokines such as IL-12 or TNF-α, alteration of mitochondrial membrane potential, and various morphological and histochemical changes to the apoptotic process. It is important to note that the betulinic acid used in this research consisted of a number of plurimolecular aggregates formed by a process of self-assembly, which is typical for this type of substance in hydroalcoholic solution. These self-assembled particles were found to exert proapoptotic and genotoxic activity in KG-1A and K562 leukemia cells, with higher efficacy than betulinic acid itself [63].

In an in vitro study using ovarian cancer cells (OVCAR 432 or RMS 13), a combination of betulinic acid with 5-fluorouracil produced a synergistic increase in the cytotoxic effect of this antitumor agent, with a concurrent increase in the subG1 population and a marked increase of apoptosis, measured by increases of cytochrome c and caspase 3, among other parameters. However, when betulinic acid treatment preceded that of 5-fluorouracil, there was a notable decrease in cytotoxicity, a result that was not explained by the authors [64].

In order to evaluate the effect of betulinic acid on tumor progression in the uterus, Karna et al. [65] studied tumor angiogenesis, assessed as the expression of hypoxia-inducible factor-1α and one of its main target genes, that of vascular endothelial growth factor, in Ishikawa cells. This is a standard model used to study human endometrial adenocarcinoma. In this context, a strong inhibition of the expression of both proteins was observed, as well as a more moderate decrease in the activity of prolinase, which is associated with angiogenesis. This finding led to the proposal that the antitumor effects of the triterpene may, in part, be due to this activity.

A triazolonaphtol derivative of betulinic acid inhibited the expression of PI3K isoforms p110α and p85α, as well as that of p-Akt and NF-κB, in in vitro models of human leukemia (HL-60 cells) and breast cancer. This caused an arrest of the cell cycle in the G0/G1 stage due to an affectation of cyclin-dependent kinase inhibitor proteins and pGSK3β [66]. Another triazole derivative, 3-[1N(2-cyanophenyl)-1H-1,2,3-triazol-4yl]methoxy betulinic acid (Fig. 7), was also reported to boost these same activities [67].

Betulinic acid was also found to facilitate autophagy, even when it had been inhibited by cytosporin A. The induction of autophagy is one way to counteract mitochondrial death by apoptosis. Although the inhibition of caspases generally has an effect on apoptosis, these enzymes were not important in this case as they did not prevent cellular death through betulinic acid. After studying several cancer cell lines, the authors proposed that autophagy probably occurs in a later step, after the mitochondrial damage has taken place [68].

Khan et al. [69] studied various betulinic acid derivatives and found that one strongly inhibited the growth of some types of human cancer cells (HL-60, MIAPaCa2, PC-3, and A549) in a narrow margin of 5–7 µM. This compound (Fig. 8) was selected from a series of relatively simple aryl derivatives of the triazole ring at C-28 carboxyl. In HL-60 cells, an increase in the ratio of Bax/Bcl-2 proteins, a marked sign of apoptosis, was very apparent at 10 and 20 µM, and was accompanied by an accumulation of cells in the G1 stage of the cell cycle. Furthermore, the mitochondrial
membrane potential was decreased even at the lowest concentration assayed (5 µM).

Several derivatives of betulinic and dihydrobetulinic acids, among them several difluorates, were synthesized from betulin. This type of double halogenation usually results in increased cytotoxic potency, which, in this case, was tested on lymphoblastic leukemia CCRF-CEM T cells. The best inhibitory results were obtained for those compounds with a 3-oxo group and a free carboxyl group (C-28), while reduction to dihydrobetulin derivatives did not significantly improve the activity [70].

Bache et al. [71] evaluated the improvements observed in betulinic acid’s efficacy after acetylation in the C-28 primary alcohol analog and glycosylation of both C-3 (compound B-10) and the amino acid ester NVX-207 (Fig. 9). The authors focused on the effects of these compounds on the growth of U251GM and U343GM glioblastoma cells with concomitant radiotherapy. The cells were examined for both their survival under conditions of normoxia and hypoxia with or without irradiation as well as for the expression of certain proteins, such as the apoptotic PARP, the inhibitor of apoptosis survivin, and the marker of tumor and hypoxia CAIX. Under hypoxia, CAIX levels, which were only measurable in this case, were virtually unchanged by betulinic derivatives, whereas survivin was dose-dependently downregulated. PARP protein expression was increased only at the highest concentration assayed (10 µM) of B10.

The p-methoxy-phenylacetic ester of betulinic acid is known as SYK023 (Fig. 10). This fairly simple derivative was assayed as an antitumor agent in a model of xenograft lung cancer in severely immunodeficient BALB/c mice as well as in a model of doxycycline-induced murine lung cancer. SYK023 substantively inhibited tumor progression in both models by means of endoplasmic reticulum stress-mediated apoptosis. The test compound affected the expression of different cyclins, inhibiting metastasis through the downregulation of synaptopodin, an actin-binding protein implicated in cytoskeletal function [72].

Another simple halogenated ester at C-3 is the dichloroacetate derivative, which was studied for its apoptotic, DNA damaging, and cytotoxic activity against several human cancer cell lines, DU 145 and PC-3 (prostate), MCF7, MDA-MB-231, and MDA-MB-468 (breast), and B16-F10 (melanoma). In each case, the potency of the ester surpassed that of betulinic acid. It was also more potent than the effect of betulinic acid + dichloroacetic acid combined as well as the effect observed in non-tumoral cells (WI-38 and 3T3 fibroblasts). In vivo, the results of the test compound on pulmonary metastases were positive, since both the number and, more importantly, the size of the nodules were notably reduced [73].

**Parasiticidal and Anti-Infectious Activity**

A study with several imidazolcarboxylate derivatives of betulin and betulinic acid found that several of them exhibited leishmanicidal activity. The compound with the highest potency (IC_{50} = 25.8 µM) was an ester of 3-keto-betul-1-en-oic acid at C-2 (Fig. 11). When exposed to this imidazole derivative, the promastigote forms of *Leishmania infantum* suffered reductions in the size of both the whole cell and the flagellum. The associated loss of motility suggests that the compound alters the cytoskeleton and affects mitochondrial function. The authors also observed cell cycle arrest in the G0/G1 phase, which would be consistent with the topoisomerase I inhibitory activity described above for other betulinic acid derivatives [74].

From the bark of *Platanus acerifolia* (Aiton) Willd. (Platanaceae) and apple peels, Innocente et al. obtained betulinic and ursolic acids [75]. These compounds, along with several alkyl-piperazinyl-
amide derivatives, were evaluated for their toxicity against Trichomonas vaginalis trophozoites. In addition to the suppression of static and kinetic growth, the rupture of the parasite membrane was also studied. The most potent compound was the amide of 3-acetyl-betulinic acid with N,N-bis-2-aminoethylpiperazine. It should be noted that this compound was also the most potent against different species of fungi belonging to Candida, Tricophyton, Epidermophyton, Microsporum, and Scytalidium [76].

One of the most pressing health problems in the field of parasitology is the control of Schistosoma, both in its infective dimension and in its effects on the environment, by influencing its vectors. Recent research has focused on the activity of lupane derivatives. Although neither betulinic acid nor its alcoholic homologue betulin were found to be active, two ionic phospho-derivatives demonstrated significant inhibitory effects on Schistosoma mansoni, especially in vitro; these were the 30-triphenylphosphonium derivatives of methyl 3-O-acetyl betulate (IC50 = 0.76 µg/mL) and 3,28-diacetylbetulin (IC50 = 0.64 µg/mL). Nevertheless, the efficacy observed in experiments in mice was slight [77].

Betulinic acid (but not its aldehyde homologue) obtained from the anti-infectious plant Callicarpa tornasola (L.) L. (syn.: Callicarpa farinosa Roxb. ex C.B.Clarke, Lamiaceae) showed a potency (MIC = 4–64 µg/mL) similar to that of mexitellicin (MIC = 16–64 µg/mL) against Staphylococcus aureus in several mexitellicin-resistant strains. Still, it was far less effective than the more toxic antibiotic vancomycin. The genes affected by treatment with subinhibitory doses of betulinic acid were components of ABC transporters (two), ribosomal assembly (one), and a two-component regulatory system (two). This last type is associated with both the virulence and antibiotic response of the bacterial cell [78].

Li et al. [79] studied the antimycobacterial constituents of the bark of Alnus incana (L.) Moench (Betulaceae), a species used as an anti-infection or antiseptic agent in different provinces of Canada. The active compounds, evaluated on Mycobacterium tuberculosis H37Ra, were triterpenic in nature. In this case, betulinic acid (IC50 = 83.5 µM; MIC > 400 µg/mL) was not the most potent compound; that distinction went to betulin (IC50 = 2.4 µM; MIC = 12.5 µg/mL).

Of great interest is the work of Lingaraju et al. [80] on the effect of betulinic acid in a model of polymicrobial sepsis in mice with cecal ligation and a puncture to release intestinal contents. Intraperitoneal administration of the triterpenoid in doses ranging from 3 to 30 mg/kg clearly improved the essential parameters of antioxidant defense, catalase and superoxide dismutase, in the kidney, an organ that suffers the consequences of sepsis in a very prominent way. Regarding the inflammation markers, there was a dose-dependent reduction of metalloproteinase-9 and TNF-α, although there was no such effect on IL-6 or IL-10.

Pharmacokinetic Properties

Because of its poor solubility in water, betulinic acid has a low gastrointestinal absorption, which never exceeds 1% [9]. As a result of their work on pharmacokinetic and tissue distribution in CD-1 mice, Udeani et al. [81] proposed a two-compartment, first-order model for pharmacokinetic modeling. Betulinic acid at doses of 250 or 500 mg/kg (intraperitoneal) had distribution volumes of 106 L/kg and 108 L/kg, respectively, with elimination half-lives of 11.5 and 11.8 h, and total clearances of 13.6 and 13.5 L/kg/h, respectively, and serum concentrations peaking at 0.15 and 0.23 h. The binding of betulinic acid to plasma proteins after intraperitoneal or intravenous administration in vivo was 99.99% (15 and 25 µg/mL in mouse, rat, or dog plasma). At 5 µg/mL, the serum protein binding reached ≥99.97% [82].

Betulinic acid distribution in tissues (at 500 mg/kg, intraperitoneal) was as follows: fat tissues (2260 µg/g at 24 h), bladder (3523 µg/g, 24 h), lymph nodes (4218 µg/g, 4 h), mammary glands (1184 µg/g, 24 h), ovaries (3055 µg/g, 4 h), spleen (1287 µg/g, 24 h), uterus (908 µg/g, 24 h), liver (223.9 µg/g, 24 h), and kidney (95.8 µg/g, 24 h), with serum levels of only 1.8 µg/mL after 24 h [37, 81]. In their previous studies on the level of betulinic acid in the blood of athymic mice bearing human melanoma, as well as in tumor and tissue homogenates, Shin et al. [83] observed high concentrations in tumors (545.2 mg/g) and liver tissue (223.9 mg/g), whereas blood levels remained low (1.8 mg/mL).

Betulinic acid’s low water solubility, described above, gives rise to certain problems for developing studies using the standard procedures for in vitro assays [37]. For this reason, researchers have studied different methods by which to dissolve the sample for use in pharmacological experiments, also in vivo. The results have been varied, as they are intrinsically influenced by the specific preparation techniques employed, that is, depending on the formulation or solvent used in the studies. Some relevant preparation techniques include the formation of a polyvinylpyrrolidone–betulinic acid complex [81], spray dried mucoadhesive microparticle formulations [84], phospholipid nanosom formats [85], and vari-
ous structural modifications [52, 67, 86, 87]. Although many of these studies focus mainly on improving the compound’s pharmacological selectivity and potency as an anticancer or antiviral agent [87], several generalizations can be made. For example, in in vivo assays, the compound is principally administered intraperitoneally, with rats and mice being the animals of choice for these studies. To assess the concentrations of the triterpene, the principal systems of analysis used are HPLC-MS [88] and GC-FID [89].

Godugu et al. [84] used a patented, dual-channel, spray gun technology for improving both the oral bioavailability and the efficacy of betulinic acid. These researchers studied the gastrointestinal permeability of the compound in vitro on the Caco-2 cell monolayer. To assess the oral bioavailability and pharmacokinetic profile, they employed Sprague Dawley rats. These studies demonstrated that betulinic acid spray in a dried mucoadhesive microparticle formulation results in a significant 3.90-fold increase in plasma C_max concentration, with the area under the curve level increasing 7.41-fold with this formulation.

The biotransformation of betulinic acid was studied in an in vitro model of metabolism designed to predict the compound’s potential mammalian metabolites. The sites of metabolism mediated by human CYP enzyme systems in the acid were also studied. The results indicated that the free carboxylic acid group (C-28) of this triterpene may act as a substrate for human CYP2C9, with the most likely sites of oxidative metabolism being the carbons C-23 and C-6a. In addition to CYP2C9, a homologous model for human CYP3A4 predicted similar sites of metabolism: positions C-6 and C-23 or C-24 [37].

Because the major problem for the potential therapeutic use of betulinic acid is its poor hydrosolubility, which limits its application to a topical use, various derivatives and forms of administration have been proposed. Modifications at positions C-3, C-20, and C-28 have been shown to increase the solubility without affecting the pharmacological activity. Alternatively, new administration routes have been proposed, such as in the form of liposomes, transdermal applications, nanoemulsions, and slow-release materials. In this context, further studies are necessary to develop novel administration formulations and methods, and this does indeed constitute one of the biggest challenges over the next few years [90].

**Toxicological Aspects**

Various studies have demonstrated a selective effect of betulinic acid on cancerous cells compared to normal cells, along with a low toxicity in vitro against normal dermal fibroblast and peripheral blood lymphocytes with respect to different human neoplastic cell lines [91]. In addition, in vitro studies with Madin Darby kidney cells showed that betulinic acid produces a significant increase in the concentration of intracellular-free calcium with only a slight decrease in cell viability [92]. These results indicate that, at standard doses, betulinic acid possesses a very narrow spectrum of cytotoxicity; indeed, only minor negative effects were observed at relatively high therapeutic doses [37].

In this regard, several in vivo studies have been developed, with varied results. For example, in 1999, Steele et al. [93] found that betulinic acid (250 mg/kg/day for 4 days, intraperitoneal) administered to mice preinfected with *Plasmodium berghei* caused the death of one of the experimental mice. However, in a previous screening, betulinic acid had shown no toxicity in rats (200 and 400 mg/kg, intraperitoneal) or mice (500 mg/kg, 6 doses, each 4th day, or 250 mg/kg, 6 doses, each 3rd day) [37].

**Summary, Future Perspectives, and Conclusions**

The reduction of glycemia through the inhibition of α-amylase and α-glucosidase, together with the stimulation of glucose uptake and glycogen synthesis through the AMPK-GSK3β pathway, constitute relevant mechanisms that justify the potential of betulinic acid as an antidiabetic agent. These effects act together with the increase of insulin biosynthesis, secretion, and sensitivity through the inhibition of PTP-1B and the resulting increase of P3K/Akt activity. Moreover, the activation of AMPK suppresses the mRNA expression and nuclear translocation of SREBP-1 and activates PPAR-α, which positively regulates lipid oxidation because PPAR-α plays a critical role in the regulation of cellular uptake, activation, and oxidation of fatty acids. In addition, betulinic acid has been shown to decrease abdominal fat accumulation, plasma triglycerides, and total cholesterol, while simultaneously increasing levels of the anorexigenic hormone and decreasing those of the orexigenic hormone ghrelin in plasma. Betulinic acid also inhibits pancreatic lipase and reduces lipid absorption in the small intestine while increasing fat mobilization through lipolysis in adipose tissues. It also reduces cholesterol absorption in the intestine via inhibition of hACAT.

The third relevant symptom of metabolic syndrome is inflammation. With this in mind, the anti-inflammatory effects of betulinic acid could be of great interest in the treatment of this increasingly prevalent pathology. To date, in vivo experiments have demonstrated that the activity of betulinic acid depends on the inhibitory effects on both PLA2 and NF-kB activation, as well as on the NF-κB-dependent gene expressions of cyclooxygenase-2 and matrix metalloproteinase-9, endow it with anti-inflammatory properties. Betulinic acid also decreases collagen biosynthesis, most likely through the disruption of the insulin-like growth factor I receptor, which regulates collagen synthesis [94]. However, while these authors observed that this effect was accompanied by an increased expression of NF-κB, a known inhibitor of collagen gene expression, Takada and Aggarwal [38] had previously described its inhibitory effects on NF-κB activation in different tumor cell lines. These contradictory effects are most likely due to the different inducers and cell lines used for NF-κB in various studies. For example, betulinic acid blocked the activation of NF-κB in colon cancer (HCT116 and Caco2) and lung cancer (H1299) cells with different agents (IL-1β, okadaic acid, H2O2, phorbol-esters, and TNF-α), whereas Karna and Palka reported a direct effect of betulinic acid on human endometrial adenocarcinoma cells (non-stimulated) [94].

Reductions of central obesity and insulin resistance, along with control of the proinflammatory state, are all desirable in order to avoid atherogenic dyslipidemia and endothelial dysfunction in vascular inflammation processes. Betulinic acid increases the
production of nitric oxide by activating the endothelial nitric oxide synthase activity that is linked to the activation of the estrogen receptor, all of which are involved in endothelial integrity.

The second group of studies on betulinic acid have focused on its potential as an antiviral and anticancer agent. In both cases, the effect of its derivatives is higher than that of the compound itself. For example, bevirimat or 3'-O-(3',3'-dimethylsuccinyl)betulinic acid and its derivatives were found to inhibit reverse transcriptase activity. A benzoic substitution at C-3 in 2,3-dehydrobetulinic acid with an ortho halogen substitution in the benzoic moiety showed the highest potency of all the inhibitors of HIV-1 maturation. Other antiviral derivatives were obtained from the original triterpene by modifying different characteristics, such as C-28 derivatives, in order to increase its solubility and improve its potency. In this way, its effect on several viruses, such as the influenza A virus (H1N1), the Coxackie virus B3, herpes viruses (HSV-1), and hepatitis B, was also examined. In these studies, the best results were obtained with conjugated structures of the triazole amide derived from dibenzylascorbic acid against the influenza A virus. Similar cases were reported for studies on the cytotoxic and antitumor properties of the compound. In these cases, betulinic acid showed both proapoptotic and genotoxic activity, though its derivatives showed higher efficacy when assayed against various leukemia cells. In some cases, the activity was increased either through modification of the pharmacokinetic properties, through different administration routes (spray dried mucoadhesive microparticle formulations or phospholipid nanosomes), or, as described above, through structural modifications.

Betulinic acid has serious limitations with regard to its direct application as a medicinal agent; however, its derivatives could be highly interesting compounds for treating various pathologies. For this reason, future studies should focus on the elaboration of new derivatives with the activities described above.

In conclusion, betulinic acid should be considered a promising drug for controlling metabolic syndrome in all its different aspects, including as a means to reduce glycemia and inflammation, as well as exerting an effect on triglycerides and cholesterol. In addition, the possible use of some of the acid’s derivatives in antiviral or oncological pharmacological interventions is also relevant, particularly for compounds such as bevirimat, which acts as an inhibitor of reverse transcriptase activity.

**Conflict of Interest**

The authors state no conflict of interest.

**References**


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