

Antimicrobial Constituents from Leaves of *Dolichandrone spathacea* and Their Relevance to Traditional Use



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ABSTRACT

Five new compounds, three iridoid glycosides (**1–3**) and two triterpenoid saponins (**4, 5**), along with thirty-two known compounds were isolated from the methanolic extract of the leaves of *Dolichandrone spathacea*. This traditional medicinal plant is widely used in Asia and India as antiseptic, for bronchitis and thrush treatment, and the methanolic extract has been shown to possess antibacterial activity against methicillin-resistant *Staphylococcus aureus*. The new iridoids were esterified derivatives of 6-ajugol and 6-catalpol, and the new saponins were glucosides of two polyhydroxy triterpenes with ursane skeleton. Their structures were elucidated by spectroscopic methods, including 1D and 2D NMR experiments and HR-ESI-MS analysis, and from comparison with the literature. This study aimed at investigating extracts and isolated compounds for their antimicrobial activities against bacterial and yeast strains, in order to validate the uses of the plant in folk medicine. The 6-O-esterified iridoids had weaker antibacterial activity; verbascoside and p-methoxycinnamic acid, the major compounds of the methanol extract, possessed strong antibacterial activity, which could account for the traditional antiseptic and anti-infectious uses of the leaves of *D. spathacea*.

Introduction

Dolichandrone spathacea (L.f.) Seem. (Bignoniaceae) also known as mangrove trumpet tree, is a common tree growing wild in river banks and mangroves of the Asia-Pacific area [1]. The leaves are used as an antitumor, antiseptic, and to treat oral thrush (as mouthwash), nervous diseases and flatulence in many countries of Southeast Asia [1, 2]. The juice of the leaves is used orally against bronchitis in India [3]. In Vietnam, this plant is part of a traditional medication (Tiêu Phong Nhuận Gan), used against hepatic disorders, skin diseases, allergies, and as detoxifier, anti-inflammatory and laxative. Several

biological activities of leaves extracts were measured showing for example, that the polar extracts possessed a high antiradical activity [2] and that the aqueous methanol extract exhibited an inhibitory effect against rat intestinal maltase [4]. Literature also reports that a methanol extract from the leaves exhibited a strong growth inhibition of six strains of methicillin-resistant *Staphylococcus aureus* (MRSA) [5]. The same article demonstrated, in a preliminary approach, the detection of flavonoids, saponins, triterpenes, and tannins in the leaves but without any structural characterization [5]. In the present paper, we report the isolation and characterization of

phytochemical constituents from leaves of *D. spathacea*, and the evaluation of antimicrobial activities of extracts and isolated compounds against different microorganisms.

Results and Discussion

The leaves of *D. spathacea* were extracted by successive percolations with petroleum ether, chloroform, ethyl acetate, methanol, and 80% aqueous methanol. The methanolic extract was purified by using a combination of liquid chromatographic techniques to yield 37 compounds. Among these, five were new compounds: three iridoid glucosides (**1-3**) and two triterpenoid saponins (**4** and **5**) (► **Fig. 1**). The 32 known isolated compounds were: 6-O-p-coumaroyl-ajugol [6], 6-O-E-caffeoyl-ajugol and 6-O-E-isoferuloyl-ajugol [7], nemoroside [8], 6-O-(E)-cinnamoyl-catalpol [9], specioside [10], 6-(E)-p-methoxycinnamoyl-catalpol [11], a mixture of (E)- and (Z)-6-p-methoxycinnamoyl-catalpol [12], verminoside and minecoside [13], nemoroside and 6''(Z)-nemoroside [14], ixoside [15], arjunglucoside I [16], decaffeoylacteoside [17], verbascoside (or acteoside) [18], isoverbascoside (or isoacteoside) [19], luteolin, luteolin-7-O-β-D-glucopyranoside, luteolin-7-O-β-D-glucuronide and luteolin-7-O-rutinoside [20–23], (2E,6E)-8-hydroxy-2,6-dimethyl-2,6-octadienoic acid [24], (2E,6Z)-8-hydroxy-2,6-dimethyl-2,6-octadienoic acid [25], 6R- and 6S-(2E)-8-hydroxy-2,6-dimethyl-2-octenoic acid in mixture [26, 27], p-hydroxybenzoic acid [28], vanillic acid [29], p-hydroxycinnamic and p-methoxycinnamic acids [30], isoferulic acid [31], and 6S,9S-roseoside [32, 33].

The new compounds (**1-5**) possessed the same sugar residue in their structures (► **Table 1** and **2**), which was determined as D-glucose after acid hydrolysis and chiral HPLC analysis. NMR showed that it was in the β-D-pyranosyl configuration.

The ¹H-NMR data (► **Table 1**) of **1** and **2** revealed the characteristic signals of an ajugol part [6] with an acetal proton at δ_H 5.53, two cis olefinic protons at δ_H 6.24 and 5.00, one methyl singlet at δ_H 1.41, and a deshielded hydroxymethine proton at δ_H 4.96. The ¹³C-NMR (DEPT and HSQC) data confirmed the presence of the ajugol moiety (► **Table 1**) [6], and the relative stereochemistry was confirmed by observation of ROE effects between H-5β/H-9β, H-1α/H-10α, H-10α/H-7α, H-7α/H-6α, and the absence of an ROE effect between H-5β/H-1α and H-6α/H-9β.

The molecular formula of **1** was determined to be C₂₅H₃₂O₁₁ by HR-ESI-MS analysis [molecular ion at m/z 531.1848 [M + Na]⁺ (calcd. for C₂₅H₃₂O₁₁Na, 531.1842)]. By subtracting the ajugol part, there remained a C₁₀H₂₄O₉ (161 uma) residue. It was assigned to a p-O-methoxycinnamoyl group by NMR: AA'BB' system of four aromatic protons at δ_H 7.56 (2H) and 6.97 (2H), one E-double bond at δ_H 7.68 and 6.43 (J = 16.0 Hz), and one methoxy group at δ_H 3.85. The ester carbonyl carbon at δ_C 167.4 (C-9'') showed an HMBC correlation with H-6 of ajugol moiety and therefore, compound **1** was determined as 6-O-(p-methoxy-E-cinnamoyl)-ajugol.

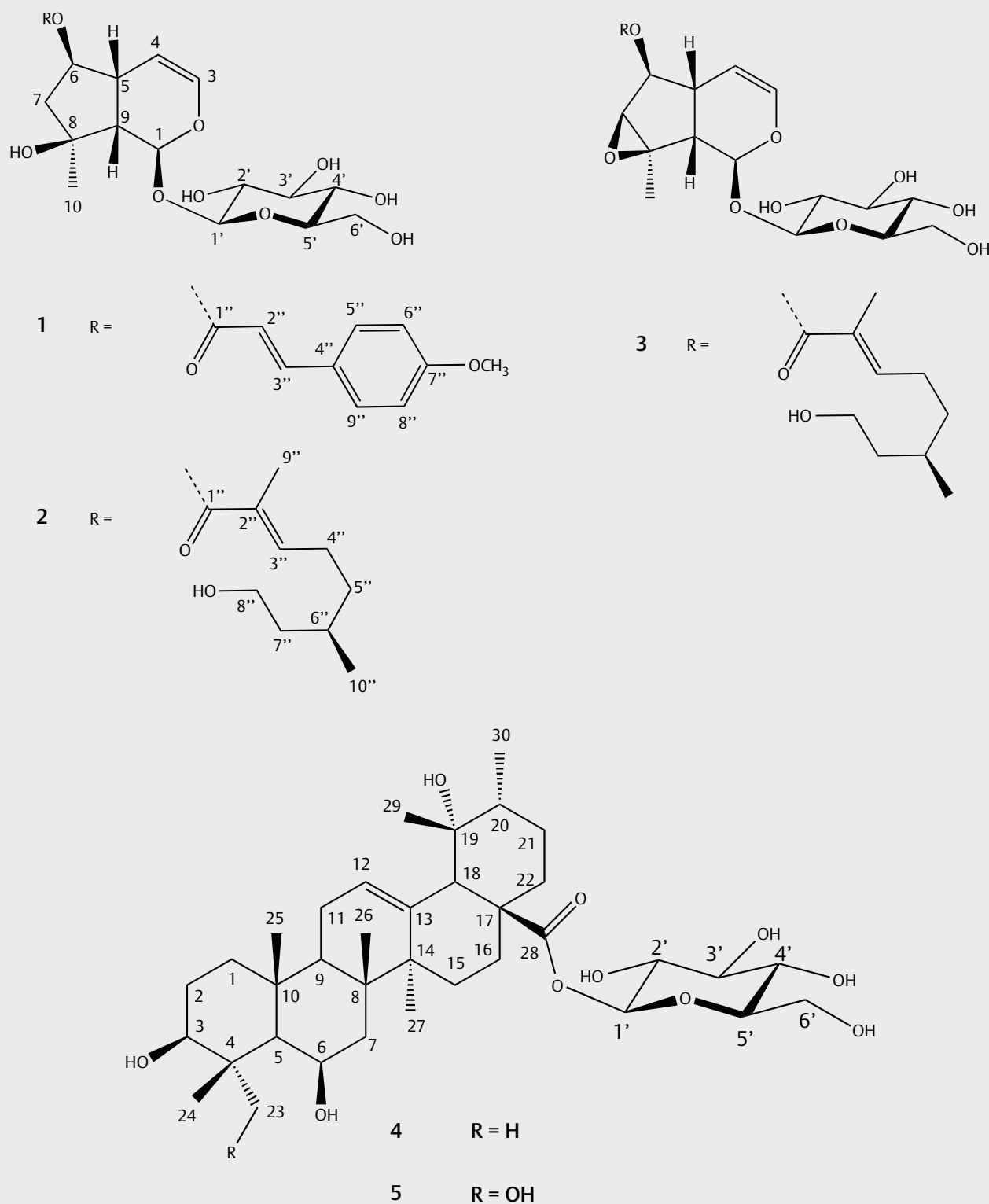
Iridoid glucoside (**2**) showed a molecular ion peak [M + Na]⁺ at m/z 539.2462 (calcd. for C₂₅H₄₀O₁₁Na, 539.2468) in agreement with a C₂₅H₄₀O₁₁ formula. Its ¹H- and ¹³C-NMR spectra (► **Table 1**) were similar to those of **1** with characteristic signals for a 6-esterified derivative of ajugol. The ¹³C-NMR and DEPT spectra of **2** exhibited ten carbons for a monoterpene acid elucidated as 8-hy-

droxy-2,6-dimethyl-2-octenoic acid which was isolated as pure known compound (see below) [26, 27] (► **Table 1**). An E-configuration could be assigned to the trisubstituted double bond according to the shielding of CH₃-9'' at δ_C 12.4 in comparison with δ_C 22–25 for Z-configuration [34]. To determine the absolute configuration of the CH-6'', we carried out the alkaline hydrolysis of **2** and then the monoterpene acid was purified by semi-preparative HPLC [18]. The positive sign of the optical rotation of the isolated acid indicated that it was 6R-(2E)-8-hydroxy-2,6-dimethyl-2-octenoic acid. The absence of any supplementary peak in the ¹³C as well as ¹H NMR spectra demonstrated that **2** was diastereoisomerically pure despite the fact that the optical rotation value of the acid ([α]_D = +3.7) was lower than those found in the literature [26, 27]. This discrepancy is most probably due to difficulties in measuring rotations on tiny amounts of material. We concluded that **2** was 6''R-O-(2E)-8-hydroxy-2,6-dimethyl-2-octenoyl-ajugol.

The HR-ESI-MS analysis of the third new iridoid glucoside (**3**) revealed a molecular formula of C₂₅H₃₈O₁₂ with the molecular ion at m/z 553.2256 [M + Na]⁺ (calcd. for C₂₅H₃₈O₁₂Na, 553.2261). The comparison of ¹H- and ¹³C-NMR spectral data (► **Table 1**) of compounds **2** and **3** showed great similarities for the ester monoterpene parts. The iridoid skeleton of **3** was identified as catalpol [14] with only one deshielded proton H-7 at δ_H 3.70 (brs) and two oxygenated carbons for CH₂OH-10 at δ_C 61.3 and -CHO-7 at δ_C 60.2 [35]. The relative stereochemistry of the six asymmetric centers was confirmed by analysis of ROE effects. The stereochemistry of the carbon C-6'' of the monoterpene acid was determined by the same method as that employed for hydrolysis of **2**. The same optical rotation ([α]_D = +3.7) was measured and compound **3** was identified as 6''R-O-(2E)-8-hydroxy-2,6-dimethyl-2-octenoyl-catalpol.

The positive HR-ESI-MS of saponin (**4**) gave a molecular ion peak [M + Na]⁺ at m/z 673.3934 (calcd. for C₃₆H₅₈O₁₀Na 673.3928) corresponding to a molecular formula C₃₆H₅₈O₁₀. The presence of 36 carbons was observed in the ¹³C-NMR spectrum including 6 carbons for a β-D-glucopyranose and 30 carbons for a Δ¹² ursene skeleton with δ_C 130.0 (CH-12) and 138.9 (C-13; ► **Table 2**). The glucose unit was linked to the triterpene moiety at position C-28 since an HMBC correlation was observed between the anomeric proton H-1' and the carbonyl C-28 at δ_C 178.5 [36]. The aglycone was determined to be triterpene uncaric acid (or 3β,6β,19α-trihydroxyurs-12-en-28-oic acid) [37]. The 3β-OH configuration was confirmed by the large coupling constant ³J_{H-3ax/H-2ax} = 11.7 Hz suitable with an (α) axial H-3, the hydroxyl group being in equatorial position (β). The ROE effects observed between H-6 and H-5α, and between H-6 and H-23α indicated a 6β-OH configuration. The third hydroxyl group was localized in axial position (19α-OH) due to the observation of ROE between the axial proton H-18β and the equatorial methyl CH₃-29β. Thus, compound **4** was identified as 28-O-β-D-glucopyranosyl-3β, 6β, 19α-trihydroxyurs-12-en-28-oic acid, named 28-O-β-D-glucopyranosyl uncaric acid.

The positive HR-ESI-MS of saponin (**5**) displayed a molecular ion peak [M + Na]⁺ at m/z 689.3869 (calcd. for C₃₆H₅₈O₁₁Na, 689.3877) consistent with the molecular formula C₃₆H₅₈O₁₁, therefore a hydroxylated derivative of saponin **4**. Comparison of the NMR spectral data of **4** and **5** revealed significant similarities (► **Table 2**); the difference was the existence of only six methyl groups and an additional hydroxyl group located at position C-23 or C-24 according



► **Fig. 1** Chemical structures of iridoids (**1**, **2**, **3**) and saponins (**4**, **5**).

to HMBC correlations between the supplementary protons CH₂-OH and carbons C-3, C-4, and C-5. The ROE effects observed between H-6 α /H-5 α , and between H-6 α and CH₂-OH confirmed the α -equatorial position of 23-CH₂-OH. The expected effects due to

24-hydroxylation were well observed on C-23 ($\Delta\alpha = +38.4$), C-4 ($\Delta\beta = +3.5$), C-24 ($\Delta\gamma = -3.5$), C-5 ($\Delta\gamma = -6.7$), and C-3 ($\Delta\gamma = -6.3$; ► **Table 2**). Thus, the aglycone of **5** was 3 β , 6 β , 19 α , 23-tetrahydroxyurs-12-en-28-oic acid [38], and saponin **5** was identified as

► **Table 1** ¹H- and ¹³C-NMR spectroscopic data (CD₃OD) of iridoids (1), (2) and (3).

| | (1) | | (2) | | (3) | |
|--------------------------------|---|------------|---|------------|--|------------|
| | δ_H (J in Hz) | δ_C | δ_H (J in Hz) | δ_C | δ_H (J in Hz) | δ_C |
| 1 | 5.53, d (2.5) | 92.0 | 5.56, d (2.4) | 93.4 | 5.18, d (9.6) | 95.1 |
| 3 | 6.24, dd (6.3, 2.2) | 139.7 | 6.23, dd (6.3, 2.2) | 141.1 | 6.39, d (6.0) | 142.4 |
| 4 | 5.00, dd (6.3, 2.6) | 103.2 | 4.97, dd (6.3, 2.7) | 104.6 | 4.98, dd (5.6, 4.1) | 102.9 |
| 5 | 2.95, dd (9.1, 2.3) | 38.0 | 2.90, dq (9.2, 2.2) | 39.3 | 2.61-2.56, m | 36.7 |
| 6 | 4.96, ddd (6.5, 4.1, 2.7) | 78.9 | 4.89, m | 80.5 | 4.98, d (7.7) | 81.6 |
| 7 | 2.02, dd (14.2, 4.0) β 2.27, dd (14.2, 6.4) α | 46.5 | 2.23, dd (14.3, 6.5) α 1.98, dd (14.3, 4.0) β | 47.8 | 3.70, brs | 60.2 |
| 8 | | 77.7 | | 79.0 | | 66.8 |
| 9 | 2.60, dd (9.3, 2.3) | 50.2 | 2.59, dd (9.2, 2.0) | 51.6 | 2.63, dd (9.3, 7.9) | 43.2 |
| 10 | 1.41, s | 24.7 | 1.39, s | 26.1 | 4.18, d (13.2) 3.85, d (13.1) | 61.3 |
| β -D-glucopyranose (C-1) | | | | | | |
| 1' | 4.69, d (8.0) | 98.0 | 4.68, d (7.9) | 99.4 | 4.81, d (7.9) | 99.7 |
| 2' | 3.22, dd (9.2, 7.9) | 73.4 | 3.22, dd (9.2, 8.1) | 74.8 | 3.30, dd (8.8, 8.3) | 74.9 |
| 3' | 3.39, dd (9.2, 8.7) | 76.6 | 3.59, t (8.9) | 78.0 | 3.43, t (9.0) | 77.7 |
| 4' | 3.31, dd (9.2, 8.6) | 71.8 | 3.29, dd (9.7, 8.5) | 71.7 | 3.28, t (9.3) | 71.8 |
| 5' | 3.32-3.35, m | 76.8 | 3.32-3.35, m | 78.2 | 3.35, ddd (9.4, 6.3, 1.5) | 78.7 |
| 6' | 3.68, dd (12.0, 6.0) 3.92, dd (12.0, 2.2) | 61.5 | 3.68, dd (12.0, 5.9) 3.92, dd (12.0, 3.1) | 62.9 | 3.67, dd (12.2, 6.1) 3.95, brd (11.9) | 62.9 |
| 6-Acyl ester | | | | | | |
| 1'' | | 167.4 | | 169.5 | | 169.5 |
| 2'' | 6.43, d (16.0) | 115.0 | | 128.8 | | 128.3 |
| 3'' | 7.68, d (16.0) | 144.8 | 6.83, td (7.5, 1.3) | 144.3 | 6.88, t (7.3) | 145.0 |
| 4'' | | 127.0 | 2.20-2.32, m | 27.2 | 2.22-2.35, m | 27.3 |
| 5'' | 7.56, d (8.6) | 129.6 | 1.47-1.54, m 1.29-1.34, m | 36.9 | 1.49-1.56, m 1.30-1.37, m | 36.9 |
| 6'' | 6.97, d (8.7) | 114.0 | 1.64-1.66, m | 30.5 | 1.60-1.67, m | 30.5 |
| 7'' | | 161.9 | 1.59-1.64, m 1.35-1.40, m | 40.6 | 1.60-1.65, m 1.41.37-1.43, m | 40.6 |
| 8'' | 7.56, d (8.6) | 114.0 | 3.62-3.67, m 3.58-3.3.62, m | 60.9 | 3.58-3.66, m | 60.9 |
| 9'' | 6.97, d (8.7) | 129.6 | 1.85, s | 12.4 | 1.89, s | 12.5 |
| 10'' | | | 0.97, d (6.5) | 19.8 | 0.97, d (6.4) | 19.8 |
| OCH ₃ | 3.85 (s) | 54.5 | | | | |

28-O- β -D-glucopyranosyl-3 β , 6 β , 19 α 23-tetrahydroxyurs-12-en-28-oic acid, or 28-O- β -D-glucopyranosyl-23-hydroxy-uncaric acid.

MIC were determined for the five extracts prepared from the leaves of *D. spathacea* against 8 Gram-positive bacterial strains, 9 Gram-negative bacterial strains, and 5 yeast strains (► **Table 3**). The results demonstrated stronger antibacterial activity of all extracts against Gram-positive bacteria, and against two pathogenic bacteria, *Streptococcus pyogenes* and *Shigella sonnei* (MIC \leq 0.3 mg/mL). The AcOEt extract was the most active against all tested microorganisms. The antimicrobial activity of the isolated compounds was first evaluated with an immersion bioautography method [39] against *Staphylococcus aureus* CIP 53.154 (► **Table 4**). Five compounds, among them decaffeoylacteoside and verbascoside that showed growth inhibition areas close to the ones observed with the antibiotic controls, as well as luteolin, p-methoxycinnamic acid and 6-O-E-caffeoyl-ajugol, seemed promising antimicrobial candi-

dates. Our results were in agreement with the literature on antibacterial activity of verbascoside [40], luteolin [41], and p-methoxycinnamic acid [42].

A serial dilution technique in 96-well plates was used to determine the MIC of the active compounds against five bacteria (► **Table 4**); the best inhibitory activity was found for the two phenylethanoid diglycosidic compounds, decaffeoylacteoside and verbascoside (MIC = 31 μ g/mL), and for p-methoxycinnamic acid (MIC = 62 μ g/mL).

In summary, it may be concluded from this study that iridoid glucosides could be considered as chemical markers of Bignoniaceae, represented here by sixteen compounds amongst which three were never described before. The 6-O-esterified iridoids like minecoside and 6-O-caffeoyl-ajugol had weaker antibacterial activity than reported for the non-esterified aucubin [43]. Six compounds of the methanol extract of *D. spathacea* showed a strong inhibitory activity against resistant bacterial strains, particularly against

► **Table 2** ¹H- and ¹³C-NMR spectroscopic data (CD₃OD) of saponins (4) and (5).

| | (4) | | (5) | |
|---------------------------------|--|------------|--|------------|
| | δ_H (J in Hz) | δ_C | δ_H (J in Hz) | δ_C |
| 1 | 0.98-1.03 m 1.58-1.66, m | 42.1 | 0.94-1.04, m 1.58-1.65, m | 41.7 |
| 2 | 1.58-1.66, m 1.70-1.78, m | 28.0 | 1.58-1.65, m 1.71-1.79, m | 27.6 |
| 3 | 3.10, dd (11.7, 4.1) | 80.2 | 3.58, dd (11.7, 3.8) | 73.9 |
| 4 | | 40.7 | | 44.2 |
| 5 | 0.75, brs | 57.2 | 1.19, brs | 49.5 |
| 6 | 4.50, brs ($w_{1/2}$ = 7.5) | 69.0 | 4.41, brs ($w_{1/2}$ = 7.8) | 68.9 |
| 7 | 1.55, dd (14.5, 2.0) eq 1.70-1.78, m ax | 41.8 | 1.51, brd (14.2) 1.78-1.85, m | 41.4 |
| 8 | | 40.3 | | 40.3 |
| 9 | 1.70-1.76, m | 49.0 | 1.71-1.79, m | 48.9 |
| 10 | | 37.6 | | 37.3 |
| 11 | 2.00-2.12, m | 24.7 | 2.01-2.13, m | 24.7 |
| 12 | 5.36, t (3.4) | 130.0 | 5.36, brt (3.4) | 130.0 |
| 13 | | 138.9 | | 138.9 |
| 14 | | 43.1 | | 43.1 |
| 15 | 1.91, td (14.0, 4.4) ax 1.03, m eq | 29.7 | 1.91, td (13.9, 3.9) ax 1.01-1.04, m eq | 29.7 |
| 16 | 2.62, td (13.3, 4.3) ax 1.64-1.68, m eq | 26.6 | 2.62, td (13.3, 4.3) ax 1.66, brd (13.0) eq | 26.6 |
| 17 | | 49.3 | | 48.6 |
| 18 | 2.55, brs | 55.0 | 2.25, s | 55.0 |
| 19 | | 73.7 | | 73.7 |
| 20 | 1.34-1.40, m | 43.0 | 1.35-1.40, m | 43.0 |
| 21 | 1.29-1.24, m 1.73-1.77, m | 27.2 | 1.24-1.29, m 1.71-1.79, m | 27.2 |
| 22 | 1.64-1.68, m 1.78-1.83, m | 38.3 | 1.66, brd (12.6) 1.78-1.85, m | 38.3 |
| 23 | 1.07, s | 28.4 | 3.50, d (10.9) 3.62, d (11.0) | 66.8 |
| 24 | 1.18, s | 17.6 | 1.09, s | 14.1 |
| 25 | 1.33, s | 17.4 | 1.35, s | 17.7 |
| 26 | 1.07, s | 18.7 | 1.07, s | 18.7 |
| 27 | 1.32, s | 24.7 | 1.33, s | 24.7 |
| 28 | | 178.5 | | 178.5 |
| 29 | 1.23, s | 27.1 | 1.23, s | 27.1 |
| 30 | 0.95, d (6.7) | 16.6 | 0.95, d (6.7) | 16.6 |
| β -D-glucopyranose (C-28) | | | | |
| 1' | 5.33, d (8.1) | 95.9 | 5.33, d (8.2) | 95.9 |
| 2' | 3.34, brt (8.2) | 73.9 | 3.35, t (8.2) | 73.9 |
| 3' | 3.43, t (8.9) | 78.3 | 3.43, t (8.8) | 78.3 |
| 4' | 3.37, t (9.0) | 71.2 | 3.38, dd (9.4, 8.5) | 71.2 |
| 5' | 3.32-3.36, m | 78.6 | 3.32-3.36, m | 78.6 |
| 6' | 3.82, dd (11.8, 2.0) 3.70, dd (11.9, 4.6) | 62.4 | 3.83, dd (12.0, 1.9) 3.71, dd (12.0, 4.6) | 62.4 |

P. aeruginosa and *E. faecalis*. Dicaffeoylacteoside, verbascoside, and p-methoxycinnamic acid, the major compounds of the methanol extract, possessed strong antibacterial activity against most of the strains tested (MIC < 100 μ g/mL). This result may explain the use of this species in folk medicine to treat oral thrush, bronchitis, and eye

infection as well as cutaneous antiseptic. The large number and wide diversity of isolated compounds from leaves of *D. spathacea* supported the other traditional uses of this plant in gastrointestinal diseases as depurative, carminative, and against liver disease.

► **Table 3** Minimal inhibitory concentration of extracts from *Dolichandrone spathacea*.

| | Microbial strain | MIC of extract (mg/mL) | | | | | Vancomycine | Gentamicine | Amphotericin B |
|------------------------|---|------------------------|-------------------|-------|-------|--------------|-------------|-------------|----------------|
| | | Petroleum ether | CHCl ₃ | AcOEt | MeOH | Aqueous MeOH | | | |
| Gram-positive bacteria | <i>Bacillus subtilis</i> ATCC 6633 | 5 | 1.2 | 5 | 10 | 10 | S | S | NT |
| | <i>Enterococcus faecalis</i> ATCC 1034 | 5 | 2.5 | 10 | 10 | 10 | R | R | NT |
| | <i>Staphylococcus aureus</i> CIP 8325-4 | 5 | 2.5 | 5 | NA | 10 | S | S | NT |
| | <i>Staphylococcus aureus</i> CIP 53.154 | 5 | 2.5 | 5 | 2.5 | 10 | S | S | NT |
| | <i>Micrococcus luteus</i> (lab. collection) | 5 | 1.2 | 5 | 5 | 10 | S | S | NT |
| | <i>Listeria innocua</i> (lab. collection) | 5 | 2.5 | 5 | NA | 10 | S | S | NT |
| | <i>Staphylococcus epidermidis</i> (lab. collection) | ≤ 0.3 | 1.2 | 2.5 | 10 | 10 | S | S | NT |
| | <i>Streptococcus pyogenes</i> (lab. collection) | ≤ 0.3 | ≤ 0.3 | ≤ 0.3 | ≤ 0.3 | ≤ 0.3 | S | S | NT |
| Gram-negative bacteria | <i>Providencia stuartii</i> (lab. collection) | 5 | 1.2 | 5 | NA | 10 | R | S | NT |
| | <i>Pseudomonas aeruginosa</i> ATCC 9027 | 10 | NA | 10 | 10 | 10 | R | S | NT |
| | <i>Shigella sonnei</i> (lab. collection) | ≤ 0.3 | ≤ 0.3 | ≤ 0.3 | ≤ 0.3 | ≤ 0.3 | I | S | NT |
| | <i>Proteus vulgaris</i> (lab. collection) | NA | NA | 10 | NA | 10 | R | S | NT |
| | <i>Klebsiella pneumoniae</i> (lab. collection) | NA | NA | 10 | NA | NA | R | R | NT |
| | <i>Serratia marcescens</i> (lab. collection) | NA | NA | 10 | NA | 10 | R | S | NT |
| | <i>Escherichia coli</i> CIP 54.127 | NA | NA | 10 | NA | NA | R | S | NT |
| | <i>Enterobacter cloacae</i> (lab. collection) | NA | NA | 10 | NA | NA | R | S | NT |
| | <i>Salmonella enterica</i> (lab. collection) | NA | NA | 10 | NA | 10 | R | S | NT |
| Yeast | <i>Candida glabrata</i> (lab. collection) | 2.5 | 1.2 | 5 | NA | 10 | R | R | S |
| | <i>Candida tropicalis</i> (lab. collection) | 5 | 10 | 5 | NA | 10 | R | R | S |
| | <i>Candida kefyr</i> (lab. collection) | 5 | 10 | 10 | NA | NA | R | R | S |
| | <i>Cryptococcus neoformans</i> (lab. collection) | 10 | NA | 10 | NA | NA | R | R | S |
| | <i>Candida albicans</i> ATCC 2091 | 5 | 10 | 5 | NA | 10 | R | R | S |

NA = not active; S = sensitive; I = intermediate sensitivity; R = resistant.

MIC (µg/mL) of positive controls: Gentamicin, S: ≤ 4, R: > 8; Vancomycin, S: ≤ 4, R: > 16; Amphotericin B, S: ≤ 1, R: > 4.

Material and Methods

General experimental procedures

Optical rotations were determined with a Perkin-Elmer 341 polarimeter. UV-Vis spectra were measured with a Shimadzu UV/Vis U-2450 spectrophotometer. IR spectra were recorded on a Nicolet Impact 410 FTIR spectrometer. HR-ESI-MS experiments were performed using a Micromass Q-TOF micro instrument. NMR spectra

were acquired in CD₃OD on a Bruker Avance DRX-600 instrument using standard pulse sequences and parameters. TLC was performed on pre-coated silica-gel 60 A: Alugram UV₂₅₄ Macherey-Nagel in normal phase, and Silicagel 60 RP-18 F₂₅₄₅ Merck in inverted phase. CC and vacuum liquid chromatography (VLC) were realized on Kieselgel 60 (63-200 mesh), Merck. Flash chromatography (FC) was carried out on a Grace Reveleris apparatus equipped with a detector ELSD, a detector UV/vis and Reveleris Flash System software. HPLC was performed on a Dionex apparatus equipped

► **Table 4** Bioautography and Minimal Inhibitory Concentration of antimicrobial compounds of *Dolichandrone spathacea*.

| Compound | Bioautography | MIC (µg/mL) | | | | |
|--|--------------------------------|--------------------------------|---------------------------------|---|-----------------------------------|---------------------------------------|
| | <i>S. aureus</i> CIP 53.154 | <i>S. aureus</i> CIP 53.154 | <i>E. faecalis</i> ATCC 1034 | <i>S. pyogenes</i> (lab. collection) | <i>P. aeruginosa</i> ATCC 9027 | <i>S. sonnei</i> (lab. collection) |
| decaffeoylacteoside | +++ | 62.5 | 31.2 | 62.5 | 250 | 31.2 |
| verbascoside | +++ | 125 | 31.2 | 125 | 250 | 31.2 |
| luteolin | ++ | 250 | 250 | 125 | 500 | 125 |
| <i>p</i> -methoxycinnamic acid | ++ | 62.5 | 125 | 62.5 | 125 | 62.5 |
| minicoside | NA | NA | 500 | 250 | 500 | 250 |
| 6- <i>O</i> - <i>E</i> -caffeoylajugol | ++ | 125 | 250 | 125 | 500 | 125 |
| Vancomycin | +++ | 3.9 | 62.5 | ≤ 1.9 | 62.5 | 7.8 |
| Gentamicin | +++ | ≤ 1.9 | 15.6 | 1.9 | 7.8 | ≤ 1.9 |

(+): Anti-staphylococcal effect, rated from + (0.5 cm inhibition zone) to ++++ (>1.5 cm inhibition zone); NA: no activity.

with a LPG 3400AB pump, an ASI-100 auto-sampler, a diode array detector UVD 340 S, an oven STH 585, and a Chromeleon software. Phenomenex Luna C18 columns (5 µm, 100 Å; 250 × 10 mm: column 1; 250 × 15 mm: column 2) and Interchrom Uptisphere Strategy C18 column (2–5 µm, 250 × 10 mm: column 3) were used for semi-preparative HPLC with gradient eluent [solvent A, H₂O or H₂O (pH 2.4, 0.025 % TFA); solvent B, MeCN or MeOH], and the chromatogram was monitored at 205 and 254 nm. Preparative HPLC chain comprises a Merck column of 200 × 50 mm packed with C-18 silica gel, an Armen AP 250/500 pump, an ACC 250/500 injector, and an UV Merck K-2501 Knauer detector; fractions were collected using a Büchi C-660 collector. Sugars were purified on a Waters chromatographic chain with a Rezex ROA column (250 × 21.2 mm: column 4), Empower software, a 600 E pump, a 717 plus auto-sampler, and a refractive index (RI) detector (flow rate = 3.5 mL/min, pressure = 1400 psi); Chiralpak IC column (5 µm, 250 × 4.6 mm: column 5) was used for the chiral identification of sugars (flow rate = 0.5 mL/min, pressure = 300 psi).

Plant material

The leaves of *D. spathacea* were collected in Vinh Long Province, Vietnam in December 2011. The plant was identified by Professor Nghia Thin Nguyen (Faculty of Biology, University of Sciences Hanoi, Vietnam) and a voucher specimen (CTU-CD001) was deposited at the Department of Biology, Faculty of Pedagogy, Can Tho University, Vietnam.

Extraction and isolation

Dried and powdered leaves (150 g) were macerated for 7 h with petroleum ether (1.5 L) to furnish 0.98 g of extract (yield 0.65 %), and treated in the same way successively with 1.5 L CHCl₃ for 20 h (5 g; yield 3.4 %), 1.5 L EtOAc (2 g; yield 1.4 %), 1.5 L MeOH (23 g; yield 15.3 %), and 1.5 L of 80 % aqueous MeOH (17 g; yield 12 %).

The MeOH extract (2 × 10 g) was subjected to silica gel VLC (7 × 9.5 cm) using a gradient of solvent CHCl₃ – MeOH – H₂O (100:0:0 – 0:100:5) to give 20 fractions. The *p*-methoxycinnamic acid (480 mg) was obtained in VLC-fraction 4 eluted with CHCl₃ – MeOH (90:10).

VLC-fraction 5 [620 mg, eluted with CHCl₃ – MeOH (85:15)] was separated by C-18 FC using a gradient of MeOH – H₂O (10:90 – 100:0); sub-fraction 5.9 [53 mg, eluted with MeOH – H₂O (45:55)]

was purified by semi-preparative HPLC (column 2; 6 mL/min) using an isocratic solvent of 25 % MeCN (pH = 2.4) during 30 min to yield (2*E*,6*E*)-8-hydroxy-2,6-dimethyl-2,6-octadienoic acid (6.2 mg; R_t = 10.66 min) and (2*E*,6*Z*)-8-hydroxy-2,6-dimethyl-2,6-octadienoic acid (3.5 mg; R_t = 11.31 min); sub-fraction 5.11 (40 mg) was purified by the same way using an isocratic solvent of 28 % MeCN to furnish a mixture of 6*R*- and 6*S*-(2*E*)-8-hydroxy-2,6-dimethyl-2-octenoic acids (5.8 mg; R_t = 14.62 min).

VLC-fraction 6 [2.3 g, eluted with CHCl₃ – MeOH (80:20)] was chromatographed by silica gel FC using a gradient of CHCl₃ – MeOH (100:0 – 0:100) to obtain *p*-hydroxycinnamic acid (22 mg) and 6-(*E*)-*p*-methoxycinnamoyl-catalpol (240 mg); sub-fraction 6.3 [31 mg, eluted with CHCl₃ – MeOH (93:7)] was purified by semi-preparative HPLC (column 1; 3 mL/min) using an isocratic mode of 23 % MeCN (pH = 2.4) to give vanillic acid (1 mg; R_t = 7.13 min) and isoferulic acid (2.3 mg; R_t = 12.39 min); the purification of sub-fraction 6.5 [28 mg, eluted with CHCl₃ – MeOH (91:9)] by semi-preparative HPLC (column 2; 6 mL/min) using a gradient solvent of 20–45 % MeCN (pH = 2.4) gave *p*-hydroxybenzoic acid (3 mg; R_t = 8.69 min) and luteolin (4.5 mg; R_t = 16.18 min); sub-fraction 6.7 (470 mg) eluted with CHCl₃ – MeOH (90:10) was separated by C-18 FC using a gradient solvent of MeOH – H₂O (10:90 – 100:0), then fractions eluted with MeOH – H₂O (60:40) were subjected to silica gel CC using a gradient system of CHCl₃ – MeOH (100:0 – 0:100) to obtain 6-*O*-(*p*-methoxy-*E*-cinnamoyl)-ajugol (**1**) (50 mg) eluted with 5 % MeOH and a mixture of (*E*)- and (*Z*)-6-*p*-methoxycinnamoyl-catalpol (2 mg; R_t = 25.46 min), finally purified by semi-preparative HPLC (column 1, 4 mL/min) using an isocratic mode of 28 % MeCN.

VLC-fraction 7 [2.6 g, eluted with CHCl₃ – MeOH (75:25)] was chromatographed by preparative HPLC using a gradient of MeOH – H₂O (25:75 – 100:0); sub-fraction 7.5 [32 mg, eluted with MeOH – H₂O (35:65)] was separated by semi-preparative HPLC (column 3, 4 mL/min) using an isocratic of 28 % MeCN (pH = 2.4) to obtain a mixture (R_t = 10.43 min) which was purified by a semi-preparative HPLC using an isocratic of 17 % MeOH during 55 min to yield 6*S*,9*S*-roseoside (6 mg; R_t = 46.98 min); the purification of sub-fraction 7.9 [90 mg, eluted with MeOH – H₂O (35:65)] by semi-preparative HPLC (column 3; 4 mL/min) using an isocratic mode of 23 % MeCN gave minicoside (5 mg; R_t = 14.49 min) and 6-*O*-*E*-isoferuloyl-ajugol (4 mg; R_t = 16.15 min); in the same way, sub-fraction 7.10 [87 mg, eluted with MeOH – H₂O (35:65)] yielded nemorososide (4 mg;

R_t = 14.70 min) and 6''(Z)-nemoside (7.5 mg; R_t = 19.02 min); the 6-O-p-E-coumaroyl-ajugol (23 mg; R_t = 25.19 min) was obtained from sub-fraction 7.12 [50 mg, eluted with MeOH – H₂O (40:60)] by a semi-preparative HPLC (column 1; 4 mL/min) using an isocratic of 20% MeCN; semi-preparative HPLC (column 3; 3.5 mL/min) of sub-fraction 7.15 [62 mg, eluted with MeOH – H₂O (60:40)] using an isocratic of 25% MeCN furnished mixture of 6''R-(2E)-8-hydroxy-2,6-dimethyl-2-octenoyl-ajugol (**2**) (7.5 mg; R_t = 22.54 min) and mixture of 6''R-(2E)-8-hydroxy-2,6-dimethyl-2-octenoyl-catalpol (**3**; 10 mg; R_t = 25.91 min); in the same way, sub-fraction 7.17 (52 mg, eluted with MeOH – H₂O (65:35)) gave 6-O-(E)-cinnamoyl-catalpol (14 mg; R_t = 20.72 min); the 28-O-β-D-glucopyranosyl uncaric acid (**4**; 7.3 mg; R_f = 0.28) was obtained from sub-fraction 7.21 (27 mg, eluted with MeOH – H₂O (100:0)) by using C-18 preparative TLC eluted with MeOH – H₂O (70:30).

VLC-fraction 8 [2.2 g, eluted with CHCl₃ – MeOH (70:30)] was separated by preparative HPLC using a gradient mode of MeOH – H₂O (25:75 – 100:0) to yield verbascoside [397 mg, eluted with MeOH – H₂O (25:75)]; the purification of sub-fraction 8.4 [397 mg, eluted with MeOH – H₂O (35:65)] by semi-preparative HPLC (column 3; 4 mL/min) using an isocratic of 10% MeCN gave decaffeoyl-acteoside (8 mg; R_t = 8.61 min); semi-preparative HPLC (column 1; 4 mL/min) applied to sub-fraction 8.12 [33 mg, eluted with MeOH – H₂O (40:60)] was carried out by using an isocratic mode of 20% MeCN to give verminoside (7 mg; R_t = 11.33 min); semi-preparative HPLC (column 3, 3.5 mL/min) of sub-fraction 8.14 [40 mg, eluted with MeOH – H₂O (45:55)] yield isoverbascoside (14 mg; R_t = 18.41 min) by using an isocratic mode of 19% MeCN; sub-fraction 8.15 [12 mg, eluted with MeOH – H₂O (45:55)] was purified by semi-preparative HPLC (column 3; 4 mL/min) using an isocratic mode of 18% MeCN to furnish luteolin-7-O-β-D-glucopyranoside (7.5 mg; R_t = 15.81 min); 6-O-E-caffeoyl-ajugol (7 mg; R_t = 24 min) was obtained from sub-fraction 8.16 [40 mg, eluted with MeOH – H₂O (50:50)] by semi-preparative HPLC (column 1; 4 mL/min) using an isocratic mode of 18% MeCN; the same protocol applied to sub-fraction 8.18 [33 mg, eluted with MeOH – H₂O (55:45)] gave specioside (9 mg; R_t = 12.87 min) using an isocratic mode of 23% MeCN; the purification of sub-fraction 8.21 [90 mg, eluted with MeOH – H₂O (65:35)] yielded arjunglucoside I (6 mg; R_t = 13.13 min) and nemoroside (9.5 mg; R_t = 17.16 min) by semi-preparative HPLC (column 3; 4 mL/min) using an isocratic mode of 25% MeCN; sub-fraction 8.23 [20 mg, eluted with MeOH – H₂O (65:35)] was purified by silica gel preparative TLC eluted with CHCl₃ – MeOH – H₂O (80:20:2) to yield 28-O-β-D-glucopyranosyl-23-hydroxy-uncaric acid (**5**; 7.5 mg; R_f = 0.33).

VLC-fraction 13 [617 mg, eluted with CHCl₃ – MeOH – H₂O (70:30:5)] was separated by C-18 FC using a gradient of MeOH – H₂O (10:90 – 100:0) to give 17 sub-fractions; luteolin-7-O-rutinoside (6 mg; R_t = 14.85 min) was obtained by semi-preparative HPLC (column 1; 3.5 mL/min) of sub-fraction 13.9 [18 mg, eluted with MeOH – H₂O (45:55)] using an isocratic solvent of 18% MeCN (pH = 2.4).

Purification of VLC-fraction 17 [90 mg, eluted with CHCl₃ – MeOH – H₂O (30:70:5)] by semi-preparative HPLC (column 1; 4 mL/min) using a gradient mode of 10 – 35% MeCN (pH = 2.4) furnished ixoside (6 mg; R_t = 5.79 min) and luteolin-7-O-glucuronide (4.5 mg; R_t = 15.89 min).

The purities of all isolated compounds were ≥ 95%, as determined by HPLC and ¹H NMR.

Compound 1: amorphous powder; $[\alpha]_D$ -143 (c 0.39, MeOH); UV (MeOH) λ_{max} nm (log ϵ) 202 (2.88), 226 (2.82), 308 (3.09); IR (KBr) ν_{max} cm⁻¹: 3428, 2931, 1706, 1662, 1632, 1575, 1514, 1170, 1002, 825; ¹H- and ¹³C-NMR: see ► **Table 1**; HR-ESI-MS m/z 531.1848 [M + Na]⁺ (calcd. for C₂₅H₃₂O₁₁Na 531.1842), 483.2442 [M + Na-MeO-OH]⁺, 185.2039 [Ajugol-Glc]⁺.

Compound 2: amorphous powder; $[\alpha]_D$ -94 (c 0.25, MeOH); UV (MeOH) λ_{max} nm (log ϵ) 224 (3.34), 308 (2.33); ¹H- and ¹³C-NMR: see ► **Table 1**; HR-ESI-MS m/z 539.2462 [M + Na]⁺ (calcd. for C₂₅H₄₀O₁₁Na 539.2468).

Compound 3: amorphous powder; $[\alpha]_D$ -107 (c 0.22, MeOH); UV (MeOH) λ_{max} nm (log ϵ) 224 (3.33), 308 (2.33); ¹H- and ¹³C-NMR: see ► **Table 1**; HR-ESI-MS m/z 553.2256 [M + Na]⁺ (calcd. for C₂₅H₃₈O₁₂Na 553.2261), 385.1032 [M + Na-Ester]⁺, 351.2457 [M-Glc]⁺.

Compound 4: white powder; $[\alpha]_D$ -18 (c 0.12, MeOH); UV (MeOH) λ_{max} nm (log ϵ) 210 (3.37), 308 (3.05); IR (KBr) ν_{max} cm⁻¹: 3423, 2929, 1734, 1653, 1458, 1383, 1074; ¹H- and ¹³C-NMR: see ► **Table 2**; HR-ESI-MS m/z 673.3934 [M + Na]⁺ (calcd. for C₃₆H₅₈O₁₀Na 673.3928), 457.2697 [M-Glc-OH]⁺.

Compound 5: white powder; $[\alpha]_D$ -7 (c 0.19, MeOH); UV (MeOH) λ_{max} nm (log ϵ) 214 (3.43), 298 (2.83); IR (KBr) ν_{max} cm⁻¹: 3422, 2927, 1732, 1598, 1455, 1384, 1072; ¹H- and ¹³C-NMR: see ► **Table 2**; HR-ESI-MS m/z 689.3869 [M + Na]⁺ (calcd. for C₃₆H₅₈O₁₁Na 689.3877), 503.3242 [M-Glc]⁺.

Acid hydrolysis

The MeOH extract (1 g) was refluxed in 25 mL of 2 N HCl in 4 h. After extraction with EtOAc (3 × 25 mL), the aqueous layer was neutralized with 0.5 M NaOH and freeze-dried. The glucose was identified by comparison with an authentic sample by TLC using MeCOEt – i-PrOH – MeCOME – H₂O (20/10/7/6). It was purified by semi-preparative HPLC (column 4) using 2.5 μM H₂SO₄ as solvent, and identified by chiral analytic HPLC (column 5) using an isocratic mixture of n-hexane/EtOH/TFA (80/20/1) in comparison with the authentic samples of D-glucose (R_t = 19.44 min) and L-glucose (R_t = 18.73 min).

Alkaline hydrolysis

4 mg of (**2** or **3**) were dissolved in 1 mL of 2 N NaOH and stirred overnight. The solution was treated with Et₂O (3 × 1 mL) to remove neutral material. Acidification with dilute HCl was followed by extraction with Et₂O (3 × 1 mL). The Et₂O extract were evaporated in vacuo and residue was purified by semi-preparative HPLC to give 1.2 mg (or 1.5 mg) of monoterpenic acid.

Antimicrobial assays

After an overnight culture at 37 °C in Mueller-Hinton (MH) medium, the bacteria (ATCC or CIP collections or clinical strains) and yeasts (clinical strains) were resuspended in fresh MH medium up to the working solution of 105 microorganisms/mL. The clinical strains were part of the internal laboratory of microbiology or parasitology collections. All the antibiotics were used in serial dilutions with the higher working concentration of 64 mg/L.

To determine the minimum inhibitory concentration (MIC) of the extracts, agar plates containing six different concentrations (10, 5, 2.5, 1.2, 0.6, and 0.3 mg/mL) of each extract were loaded with the different microorganism using a multiple inoculator (Steers). After 24 h of incubation at 37 °C, the activity was estimated looking at the presence or absence of colonies. Solvents used to prepare extracts and MH agar medium were checked for absence of antibacterial activity (negative controls). Positive active controls were gentamicin ($\geq 99\%$ of purity) and vancomycin ($\geq 99\%$ of purity) for antibacterial assays, and amphotericin B ($\geq 99\%$ of purity) for anti-yeast assays.

The immersion bioautography method was used to identify the active compounds against *S. aureus* CIP 53.154. 10 μ L of each compound (2 mg/mL of methanol) or gentamicin (50 μ g/mL, positive control) were spotted onto TLC plates (10 \times 10 cm). The plates were then dried, sterilized and covered by the MH agar medium containing *S. aureus* (10^5 bacteria/mL) in square Petri dishes. After incubation for 24 h at 37 °C, bacterial growth was revealed by a 2 mg/mL solution of thiazolybluetetrazolium bromide (MTT) and growth inhibition zones were measured [39].

The broth microdilution method in 96-well plates was used to determine MIC of active compounds against five bacteria sensitive to the extracts. Nine concentrations of each compound, from 500 μ g/mL to 1.9 μ g/mL in a serially twofold dilution, were tested against 0.5×10^5 bacteria/mL, in a total volume of 200 μ L, and incubated overnight at 37 °C. Positive control wells with bacteria in MH medium, gentamicin or vancomycin, as well as negative controls such as medium alone or medium with methanol were systematically added to the test. Bacterial growth was followed visually and by spraying MTT incubating at 37 °C for at least 10 min. The MIC value was determined as the lowest concentration of a compound leading to a clear well. This test was performed in triplicate.

Supporting information

Tables with ^1H - and ^{13}C -NMR data of the 30 known compounds, and ^1H - and ^{13}C -NMR Spectra of compounds (1-5) are available as Supporting Information.

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Conflict of Interest

The authors declare no conflict of interest.

References

- [1] Wiart C. Medicinal plants of the Asia-pacific: drugs for future? Singapore: World Scientific Publishing; 2006: 565–566
- [2] Kaewpiboon C, Lirdprapamongkol K, Srisomsap C, Winayanuwattikun P, Yongvanich T, Puwaprisirisan P, Svasti J, Assavalapsakul W. Studies of the in vitro cytotoxic, antioxidant, lipase inhibitory and antimicrobial activities of selected Thai medicinal plants. *BMC Complement Altern Med* 2012; 12: 217–224
- [3] Prasad PRC, C. Reddy S, Raza SH, Dutt CBS. Folklore medicinal plants of North Andaman Island, India. *Fitoterapia* 2008; 79: 458–464
- [4] Jong-Anurakkun N, Bhandari MR, Kawabata J. α -Glucosidase inhibitors from Devil tree (*Alstonia scholaris*). *Food Chem* 2007; 103: 1319–1323
- [5] Saiful AJ, Mastura M, Mazurah MI, Nuziah H. Inhibitory potential against methicillin-resistant *Staphylococcus aureus* (MRSA) of *Dolichandrone spathacea*, a mangrove tree species of Malaysia. *Lat Am J Pharm* 2011; 30: 359–362
- [6] Nishimura H, Sasaki H, Morota T, Chin M, Mitsushashi H. Six iridoid glycosides from *Rehmannia glutinosa*. *Phytochemistry* 1989; 28: 2705–2709
- [7] Harinantenaina L, Kasai R, Rakotovo M, Yamasaki K. New iridoid and phenethyl glycosides from Malagasy medicinal plant, *Phyllarthron madagascariense*. *J Nat Med* 2001; 55: 187–192
- [8] Junior P. Nemoroside and Nemoroside, zwei neue Iridoidglucoside aus *Penstemon nemorosus*. *Planta Med* 1983; 47: 67–70
- [9] Sarg T, Salama O, El-Domiaty M, Bishr M, Mansour ES, Weight E. Iridoid glucosides from *Gentiana kurroo* Royle. *Alex J Pharm Sci* 1991; 5: 82–86
- [10] El-Naggar SF, Dorskotch RW. Specioside: a new iridoid glycoside from *Catalpa speciosa*. *J Nat Prod* 1980; 43: 524–526
- [11] Houghton PJ, Hikino H. Anti-hepatotoxic activity of extracts and constituents of *Buddleja* species. *Planta Med* 1989; 55: 123–126
- [12] Dellar JE, Conn BJ, Cole MD, Waterman PG. Cinnamate esters of catalpol from *Westringia fruticosa* and *Westringia viminalis*. *Biochem Syst Ecol* 1996; 24: 65–69
- [13] Sticher O, Afifi-Yazar FU. Minecosid und Verminosid, zwei neue Iridoidglucoside aus *Veronica officinalis* L. (Scrophulariaceae). *Helv Chim Acta* 1979; 62: 535–539
- [14] Arslanian RL, Anderson T, Stermitz FR. Iridoid glucosides of *Penstemon ambiguus*. *J Nat Prod* 1990; 53: 1485–1489
- [15] Takeda Y, Nishimura H, Inouye H. Two new iridoid glucosides from *Ixora chinensis*. *Phytochemistry* 1975; 14: 2647–2650
- [16] Abe F, Yamauchi T. Glycosides of 19 α -hydroxyoleanane-type triterpenoids from *Trachelospermum asiaticum* (*Trachelospermum*. IV). *Chem Pharm Bull* 1987; 35: 1833–1838
- [17] Karasawa H, Kobayashi H, Takizawa N, Miyase T, Fukushima S. Studies on the constituents of *Cistanchis* Herba. VII: Isolation and structures of Cistanosides H and I. *Yakugaku Zasshi* 1986; 106: 562–566
- [18] Andary C, Wylde R, Laffite C, Privat G, Winternitz F. Structures of verbascoside and orobanchoside, caffeic acid sugar esters from *Orobancha rapum-genistae*. *Phytochemistry* 1982; 21: 1123–1127
- [19] Suo M, Ohta T, Takano F, Jin S. Bioactive phenylpropanoid glycosides from *Tabebuia avellaneda*. *Molecules* 2013; 18: 7336–7345
- [20] Liu QM, Zhao HY, Zhong XK, Jiang JG. *Eclipta prostrata* L. phytochemicals: Isolation, structure elucidation, and their antitumor activity. *Food Chem Toxicol* 2012; 50: 4016–4022
- [21] Orhan F, Barış O, Yanmış D, Bal T, Güvenalp Z, Güllüce M. Isolation of some luteolin derivatives from *Mentha longifolia* (L.) Hudson subsp. *longifolia* and determination of their genotoxic potencies. *Food Chem* 2012; 135: 764–769

- [22] Özgen U, Mavi A, Terzi Z, Kazaz C, Asçi A, Kaya Y, Seçen H. Relationship between chemical structure and antioxidant activity of luteolin and its glycosides isolated from *Thymus sipyleus* subsp. *sipyleus* var. *sipyleus*. *Rec Nat Prod* 2011; 5: 12–21
- [23] Kokotkiewicz A, Luczkiewicz M, Sowinski P, Glod D, Gorynski K, Bucinski A. Isolation and structure elucidation of phenolic compounds from *Cyclopia subternata* Vogel (honeybush) intact plant and in vitro cultures. *Food Chem* 2012; 133: 1373–1382
- [24] Iwagawa T, Asai H, Hase T, Sako S, Su R, Hagiwara N, Kim M. Monoterpenoids from *Radermachia sinica*. *Phytochemistry* 1990; 29: 1913–1916
- [25] Ono T, Koutari S, Marumoto S, Miyazawa M. Novel compound, (2Z,6E)-1-hydroxy-3,7-dimethyl-2,6-octadien-8-oic acid produced from biotransformation of nerol by *Spodoptera litura* Larvae. *J Oleo Science* 2013; 62: 313–318
- [26] Yamaguchi K, Shinohara C, Kojima S, Sodeoka M, Tsuji T. (2E,6R)-8-Hydroxy-2,6-dimethyl-2-octenoic acid, a novel anti-osteoporotic monoterpene, isolated from *Cistanche salsa*. *Biosci Biotechnol Biochem* 1999; 63: 731–735
- [27] Machida K, Ando M, Yaoita Y, Kakuda R, Kikuchi M. Studies on the constituents of *Catalpa* species. VI. Monoterpene glycosides from the fallen leaves of *Catalpa ovata* G. Don. *Chem Pharm Bull* 2001; 49: 732–736
- [28] Peungvicha P, Tamsiririrkkul R, Prasain JK, Tezuka Y, Kadota S, Thirawarapan SS, Watanabe H. 4-Hydroxybenzoic acid: a hypoglycemic constituent of aqueous extract of *Pandanus odoratus* root. *J Ethnopharmacol* 1998; 62: 79–84
- [29] Sakushima A, Coşkun M, Maoka T. Hydroxybenzoic acids from *Boreava orientalis*. *Phytochemistry* 1995; 40: 257–261
- [30] Silva AMS, Alkorta I, Elguero J, Silva VLM. A ¹³C NMR study of the structure of four cinnamic acids and their methyl esters. *J Mol Struct* 2001; 595: 1–6
- [31] Kelley CJ, Harruff RC, Carmack M. Polyphenolic acids of *Lithospermum ruderale*. II. Carbon-13 nuclear magnetic resonance of lithospermic and rosmarinic acids. *J Org Chem* 1976; 41: 449–455
- [32] Çalış I, Kuruüzüm-Uz A, Lorenzetto PA, Rüedi P. (6S)-Hydroxy-3-oxo- α -ionol glucosides from *Capparis spinosa* fruits. *Phytochemistry* 2002; 59: 451–457
- [33] Yamano Y, Ito M. Synthesis of optically active vomifoliol and roseoside stereoisomers. *Chem Pharm Bull* 2005; 53: 541–546
- [34] Couperus PA, Clague ADH, van Dongen JPCM. ¹³C chemical shifts of some model olefins. *Org Magn Reson* 1976; 8: 426–431
- [35] Dinda B, Debnath S, Harigaya Y. Naturally occurring iridoids. A review, Part 1. *Chem Pharm Bull* 2007; 55: 159–222
- [36] Acebey-Castellon IL, Voutquenne-Nazabadioko L, Doan TMH, Roseau N, Bouthagane N, Muhammad D, Le Magrex Debar E, Gangloff SC, Litaudon M, Sévenet T, Nguyen VH, Lavaud C. Triterpenoid saponins from *Symplocos lancifolia*. *J Nat Prod* 2011; 74: 163–168
- [37] Aquino R, De Simone F, Vincieri FF, Pizza C, Gacs-Baitz E. New polyhydroxylated triterpenes from *Uncaria tomentosa*. *J Nat Prod* 1990; 53: 559–564
- [38] Ma X, Yang C, Zhang Y. Complete assignments of ¹H and ¹³C NMR spectral data for three polyhydroxylated 12-ursen-type triterpenoids from *Dischidia esquirolii*. *Magn Reson Chem* 2008; 46: 571–575
- [39] Abedini A, Roumy V, Mahieux S, Biabiany M, Standaert-Vitse A, Rivière C, Sahpaz S, Bailleul F, Neut C, Hennebelle T. Rosmarinic acid and its methyl ester as antimicrobial components of the hydroxymethanolic extract of *Hyptis atrorubens* Poit. (Lamiaceae). *Evid Based Complement Alternat Med* 2013; 2013: 604536
- [40] Nazemiyeh H, Rahman MM, Gibbons S, Nahar L, Gharamani MA, Talebpour AH, Sarker SD. Assessment of the antibacterial activity of phenylethanoid glycosides from *Phlomis lanceolata* against multiple-drug-resistant strains of *Staphylococcus aureus*. *J Nat Med* 2008; 62: 91–95
- [41] Joung DK, Lee YS, Han SH, Lee SW, Cha SW, Mun SH, Kong R, Kang OH, Song HJ, Shin DW, Kwon DY. Potentiating activity of luteolin on membrane permeabilizing agent and ATPase inhibitor against methicillin-resistant *Staphylococcus aureus*. *Asian Pac J Trop Med* 2016; 9: 19–22
- [42] Rigano D, Formisano C, Basile A, Lavitola A, Senatore F, Rosselli S, Bruno M. Antibacterial activity of flavonoids and phenylpropanoids from *Marrubium globosum* ssp. *libanoticum*. *Phytother Res* 2007; 21: 395–397
- [43] Yuan CS, Zhang Q, Xie W, Yang XP, Jia ZJ. Iridoids from *Pedicularis kansuensis* forma *albiflora*. *Pharmazie* 2003; 58: 428–430