









Evaluation of the Association between Genetic Polymorphism of Interleukin-1 Beta (-511C/T and +3953C/T) and Cervical Cancer Susceptibility

Pushpendra D. Pratap¹ Syed Tasleem Raza¹ Ghazala Zaidi² Shipra Kunwar³ Ale Eba¹ Muneshwar Rajput¹

- ¹Department of Biochemistry, Central Research Laboratory, Molecular Diagnostic Unit, Era's Lucknow Medical College, Era University, Lucknow, Uttar Pradesh, India
- ²Department of Allied Health Sciences, Era University, Lucknow, Uttar Pradesh, India
- ³Department of Obstetrics and Gynaecology, Era's Lucknow Medical College, Era University, Lucknow, Uttar Pradesh, India

Address for correspondence Syed Tasleem Raza, PhD Genetics, Department of Biochemistry, Central Research Laboratory, Molecular Diagnostic Unit, Era's Lucknow Medical College, Era University, Lucknow 226003, Uttar Pradesh, India (e-mail: tasleem24@gmail.com).

South Asian | Cancer

Abstract



Pushpendra D. Pratap

Keywords

- ELISA
- gene polymorphism
- **HPV**
- PCR-RFLP
- RT-PCR

Cervical cancer (CC) is the leading cause of cancer-related mortality among women, primarily caused by persistent human papillomavirus (HPV) infection, especially in developing countries. A proinflammatory cytokine, emerging as a major facilitator of carcinogenesis, is termed interleukin-1 beta (IL-1β), which characterizes host-environment interactions. Numerous epidemiological studies have revealed that IL-1β gene polymorphisms have been associated with numerous malignancies, but in the context of CC, results of these studies were inconclusive. Thus, our study aimed to explore the relationship between IL-1β polymorphisms (-511C/T and +3953C/T) and CC susceptibility. Genotyping was conducted on 192 CC patients and 200 healthy controls through polymerase chain reaction-restricted fragment length polymorphism. HPV analysis was done through real-time polymerase chain reaction, and the serum concentration of IL- 1β was measured by enzyme-linked immunosorbent assay. Women with CT and TT genotypes of IL-1 β -511C/T had a threefold increased risk of CC (odds ratio [OR], 3.60; 95% confidence interval [CI], 2.132-6.063; *p* < 0.001 vs. OR, 3.34; 95% CI, 1.952-5.713; p < 0.001) compared to controls. Women with the T allele of IL-1 β -511C/T polymorphism were associated with increased CC susceptibility (OR, 2.00; 95% CI, 1.51-2.66; p = 0.0001) compared to controls. No significant difference was found between patients and controls in the genotype or allele frequencies of IL-1β +3953C/T polymorphism (OR, 0.93; 95% CI, 0.56-1.55; p = 0.86 vs. OR, 0.95; 95% CI, 0.72-1.26; p = 0.74). There was no significant association found between IL-1 β -511C/T promoter (OR, 2.41; 95% CI, 0.46-12.76; p = 0.28 vs. OR, 1.64; 95% CI, 0.13-21.10; p = 0.7) and +3953C/T (OR, 3.76; 95% CI, 0.44-31.82; p = 0.19 vs. OR, 0.21; 95% CI, 0.01-3.92; p = 0.25) polymorphisms in tobacco chewers and smokers compared to

DOI https://doi.org/10.1055/s-0043-1776788 ISSN 2278-330X

How to cite this article: Pratap PD, Raza ST, Zaidi G, et al. Evaluation of the Association between Genetic Polymorphism of Interleukin-1 Beta (-511C/T and +3953C/T) and Cervical Cancer Susceptibility. South Asian | Cancer 2024;00(00):00-00

© 2024. MedIntel Services Pvt Ltd. All rights reserved.

This is an open access article published by Thieme under the terms of the Creative Commons Attribution-NonDerivative-NonCommercial-License, permitting copying and reproduction so long as the original work is given appropriate credit. Contents may not be used for commercial purposes, or adapted, remixed, transformed or built upon. (https://creativecommons.org/licenses/by-nc-nd/

Thieme Medical and Scientific Publishers Pvt. Ltd., A-12, 2nd Floor, Sector 2, Noida-201301 UP, India

controls. The level of serum concentration of IL-1 β was significantly higher in cases compared to controls. Our results conclude that IL-1 β -511C/T polymorphism is associated with CC susceptibility.

Introduction

Cervical cancer (CC) is the second most prevalent gynecological malignancy after breast cancer, and it is one of the major causes of cancer-related mortality in females around the world. As per 2020 GLOBOCAN, it is the fourth most commonly diagnosed cancer and the leading cause of cancer death in women, with an estimated 604,127 new cases and 341,831 fatalities. Epidemiological studies have demonstrated that the human papilloma virus (HPV) is officially designated as the main cause of CC, primarily HPV-16 and HPV-18.² HPV-16 was found to be highly significant in tissues from CC patients, followed by HPV-18.³ According to the molecular mechanism of CC, HPV deoxyribonucleic acid (DNA) was integrated into the host's genome which prompted keratinocytes to change the uterine cervix into cancer.4 The uterine body and vagina are connected via the cervix. The area of the cervix closest to the uterine body is termed the endocervix. The exocervix is the region of the uterus that is located nearest to the vagina. The cells that line the cervix, also known as the uterine cervix, the lowest portion of the uterus, are where CC first develops. ⁵ The glandular cells and the squamous cells, which coat the cervix, are primarily two cell types. The "transformation zone" is the location where these two subtypes of cells collide. As a person gets older and after giving birth, the transition zone's location changes. The transition zone is a common place where CC develops.⁵

CC is caused by a multitude of factors, including multiple sexual partners, age at first sexual intercourse, parity, heavy cigarette smoking, immune suppression, tobacco and alcohol use, and coinfection with other sexually transmitted agents. The immune system is crucial in suppressing or encouraging cancer by activating a variety of immunological components, especially cells and cytokines. The antigen presentation response, which is downregulated in CC and has a negative impact on cellular and humoral responses, appears to alter carcinogenesis. As a result of the dynamic interactions between environmental exposure and host genetic background, it is widely regarded as a complex disease. Polymorphisms in the human genome are now acknowledged as a fundamental cause of the disease.

Interleukin (IL)-1 is a proinflammatory cytokine primarily produced by macrophages, monocytes, and epithelial cells that has a role in inflammation, immunological response, and angiogenesis of tumor or tumor initiation. The interleukin-1 alpha (IL-1 α), interleukin-1 beta (IL-1 β), and IL-1 receptor antagonist genes are located in the 2q13-21 gene family and encode IL-1 α , IL-1 β , as well as the anti-inflammatory cytokine IL-1RA. Breast, lung, colon, and melanomas are solid tumors in which IL-1 β production has a poor prognosis. Polymorphisms in the IL-1 β gene have been linked to susceptibility to a variety of cancers, according to research. To the best of our knowledge, only a few studies

have investigated the association between IL-1 β gene polymorphism and CC risk, but results were inconclusive. As a result, the purpose of this study was to look into the relationship between IL-1 β –511C/T and +3953C/T gene polymorphisms and CC susceptibility.

Materials and Methods

Study Design: Case-Control Study

Patients and Sample Collection

For this case-control study, 192 women with CC and 200 healthy controls were recruited from the outpatient department (OPD) of the Department of Obstetrics and Gynaecology, Era's Lucknow Medical College and Hospital, Lucknow, Uttar Pradesh, India. All patients had recently been diagnosed, histopathologically verified, and staged as per International Federation of Gynaecology and Obstetrics (FIGO) criteria. The institutional ethics committee gave its approval, and each subject gave their informed consent. Patients with history of cancer or who had received radiotherapy or chemotherapy were excluded. Controls were recruited from the same department as those attending OPD, confirmed to be cervical cytology negative in the pathology department, and ascertained to have no history of cancer, infection, or any acute or chronic pathology. Each subject's peripheral blood (5 mL) was collected into tubes containing ethylenediamine tetraacetic acid and plain vials prior to radiation therapy and/or chemotherapy at the time of the initial diagnosis. Each participant's blood and serum samples were kept at -70°C until further use.

Genomic DNA Extraction and Genotyping

The PureLink® Genomic DNA extraction kit was used to extract genomic DNA from each subject according to the manufacturer's instructions. To determine the IL-1 β promoter polymorphism at position –511C/T gene, polymerase chain reaction-restricted fragment length polymorphism (PCR-RFLP) method ¹³ was used with the following specific primer sequence: 5'-TGGCATTGATCTGGTTCATC-3' forward, and 5'-GTTTAGGAATCTTCCCACTT-3' reverse. For each sample, PCR was carried out in a final volume of 20 μ L including 2 μ L of genomic DNA, 25 mM of Deoxynucleotide triphosphates (dNTP), 1 mmol/L of MgCl2, 1.5 units of Taq, that is, DNA polymerase (Fermentas, Thermo Fisher Scientific, BIO-RAD, USA), and 5 μ L of buffer with annealing at 58°C.

The amplified product, that is, 15 µL was digested with 5 U of Aval (New England Biolabs, Beverly, Massachusetts, United States) at 37°C overnight, then the products were separated on 3% agarose gel electrophoresis with EtBr (Invitrogen Life Technologies, Carlsbad, California, United States) and visualized under ultraviolet light with a 100 base pair ladder (►Fig. 1A).

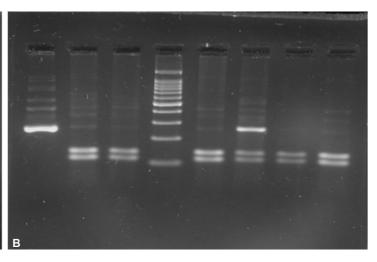


Fig. 1 (A) Agarose gel picture showing interleukin (IL)-1 β (-511C/T) gene polymorphism: Lanes 1 and 3, TT genotype; Lanes 2 and 5–8, CT genotype; Lane 4, molecular marker 100 bp. (B) Agarose gel picture showing IL-1 β +3953 C/T gene polymorphism: Lane 1, TT genotype; Lanes 2, 3, 5, 7, and 8, CC genotype; Lane 6, CT genotype; and Lane 4, molecular marker 100 bp.

For genotyping of the IL-1β polymorphism at position +3953, PCR amplification was followed by RFLP analysis. ¹⁴ Using the primer sequence 5'-GTT GTC ATC AGA CTT TGA CC-3' forward and 5'-TTC AGT TCA TAT GGA CCA GA-3' reverse, the target DNA sequence in exon 5 of the IL-1β gene was amplified with annealing at 59°C. The amplified product was digested with 2.5 U Taq 1 at 37°C overnight and separated on 3% agarose gel electrophoresis and under ultraviolet light (►Fig. 1B).

The concentration of IL-1 β in human serum samples was measured by enzyme-linked immunosorbent assay (ELISA) using the Human IL-1 β Diaclone ELISA Kit as per the manufacturer's protocol (Cat. No.850.006.096).

Statistical Analysis

The SPSS v. 21.0 tool (SPSS Inc., Chicago, Illinois, United States) was used to analyze the data. For continuous variables, the two-tailed *t*-test was employed, and for categorical variables, the Chi-square (v2) test was utilized. Using the v2 test, Hardy-Weinberg equilibrium studies were used to compare observed and anticipated genotype frequencies. The odds ratio (OR) and their 95% confidence interval (CI) from multivariate logistic regression analysis were used to determine the correlations between genotypes and CC risk. A statistically significant *p*-value of 0.05 was used.

Results

The demographic characteristics of cases and controls are summarized in **Table 1**. The mean age of cases was 49.8 years on average, while controls were 44.9 years old. Histopathological distribution of CC was keratinizing squamous cell carcinoma (85.42%) and adenocarcinoma (4.16%), as shown in **Table 1**. Staging of all the cases was scrutinized through FIGO criteria. Among them, majority of the cases (57.30%) were diagnosed with stage II. Between patients and controls, there was a significant difference in serum IL-1β levels.

IL-1 Beta Polymorphism -511C/T Polymorphism

Genotype distribution of IL-1β polymorphism –511C/T in the promoter region among CC patients and controls is summarized in **Table 2**. Overall, three genotypes (CC, CT, and TT) and two alleles (C and T) were seen in this study. Genotype

Table 1 Clinicopathological characteristics of women with cervical cancer

Cases (n = 192)	Controls (n = 200)			
Age (years)	n	%	n	%
o 21–30	10	5.2	6	3
o 31–40	32	16.67	62	31
o 41–50	76	39.59	102	51
o 51–60	44	22.92	16	8
o 61–70	26	13.54	10	5
o >70	4	2.08	4	2
o Mean age	49.8 years		44.9 years	
Histopathological analysis				
o Nonkeratinizing SCC	20	10.42	N/A	N/A
o Keratinizing SCC	164	85.42	N/A	N/A
o Adenocarcinoma	8	4.16	N/A	N/A
FIGO staging				
o Stages I	42	21.87	N/A	N/A
o Stages II	110	57.3	N/A	N/A
o Stages III	30	15.62	N/A	N/A
o Stages IV	10	5.21	N/A	N/A
Tobacco chewing	96	50	57	28.5
Smoking	36	18.75	16	8

Abbreviations: FIGO, International Federation of Gynaecology and Obstetrics; N/A, not available; SCC, squamous cell carcinoma.

Table 2 Distribution of genotype and allele of interleukin-1 β (–511C/T and +3953 C/T) single nucleotide polymorphism in cervical cancer and control samples

IL-1β –511 C/T	Genotype f	requencies		Association analysis			
	Patients		Controls		Patients vs. controls		
Genotype	n = 192	%	n = 200	%	OR (95% CI)	<i>p</i> -Value	
Codominant model							
o CC	32	16.67	82	41	Reference	Reference	
o CT	87	45.31	62	31	3.60 (2.132-6.063)	< 0.001	
o TT	73	38.02	56	28	3.34 (1.952–5.713)	< 0.001	
Dominant model							
o CC	32	16.66	82	41	Reference	Reference	
o CT + TT	160	83.34	118	59	3.47 (2.17–5.57)	< 0.001	
Recessive model							
o CC + CT	119	61.98	144	72	Reference	Reference	
o TT	73	38.02	56	28	1.58 (1.03–2.41)	0.034	
Allele							
οС	151	39.33	226	56.5	Reference	Reference	
οΤ	233	60.67	174	43.5	2.00 (1.51–2.66)	0.0001	
IL-1β +3953 C/T							
Codominant model							
o CC	69	35.94	67	33.5	Reference	Reference	
o CT	71	36.98	79	39.5	0.87 (0.54–1.38)	0.565	
o TT	52	27.08	54	27	0.93 (0.56–1.55)	0.865	
Dominant model							
o CC	69	35.94	67	33.5	Reference	Reference	
o CT + TT	123	64.06	133	61.5	0.90 (0.59-1.36)	0.612	
Recessive model							
o CC + CT	140	72.92	146	73	Reference	Reference	
o TT	52	27.08	54	27	1.00 (0.64–1.57)	0.985	
Allele							
ο С	209	54.42	213	53.3	Reference	Reference	
οΤ	175	45.58	187	46.8	0.95 (0.72–1.26)	0.741	

Abbreviations: CI, confidence interval; IL, interleukin; OR, odds ratio.

frequencies in cases were (CC, CT, and TT) 16.67%, 45.31%, and 38.02% versus 41, 31, and 28% as compared to controls. The allele frequencies were 0.61 for the C allele (ancestral allele) and 0.60 for T allele (variant allele) in the cases while in the controls 0.56 and 0.43 for C and T alleles, respectively. Statistical analyses for genotype and allele frequencies included codominant, dominant, and recessive models, as summarized in **Table 2**. C/T and T/T genotype frequencies were higher in cases as compared to controls (p < 0.001). As compared to controls, CC patients had a higher frequency of C/T+T/T genotype (p < 0.001). Frequency of T allele was higher in cases as compared to controls (p = 0.0001). Women with the C/T genotype (OR, 3.60; 95% CI, 2.132–6.063), T/T genotype (OR, 3.34; 95% CI, 1.952–5.713), C/T+T/T genotypes (OR, 3.47; 95% CI, 2.17–5.57), and T allele (OR, 2.00; 95%

CI, 1.51–2.66) were more likely to develop CC. The polymorphism of IL-1 β –511C/T has been linked significantly to CC.

IL-1β +3953C/T Polymorphism

IL-1 β +3953C/T polymorphism frequency distribution between CC cases and controls is summarized in **Table 2**. In this study, three genotypes and two allele were observed. Frequencies of genotypes were C/C 35.94%, C/T 36.98%, and T/T 27.08% in CC patients group and 33.50%, 39.50%, and 27% in the control group, respectively. In the controls, the allele frequencies for alleles C (ancestral allele) and T (variant allele) were 53.3% and 46.80%, respectively, whereas in the cases, the allele frequencies for alleles C and T were 54.42% and 45.58%, respectively. When compared to the controls, there was no significant association found (p > 0.05).

Data confirming whether IL-1 β (-511C/T and +3953C/T) polymorphism is associated with tobacco chewing and smoking are summarized in **Table 3**. On the comparison to controls there was no association found among genotype and allele frequencies between tobacco chewers and smok-

ers (p > 0.05). In context of -511C/T polymorphism, in the individuals carrying C/T and T/T genotypes (OR, 1.79; 95% CI, 0.91–3.54; p = 0.09 and OR, 2.41; 95% CI, 0.46–12.76; p = 0.28), T allele (OR, 1.58; 95% CI, 0.93–2.68; p = 0.09) was not associated with CC susceptibility in tobacco chewers

Table 3 Distribution of genotype and allele frequency of interleukin-1 β (-511C/T and +3953 C/T) single nucleotide polymorphism with tobacco chewing and smoking versus controls

IL-1β –511 C/T	Genotype	frequencies		Association analysis	Association analysis		
	Patients		Controls		Patients vs. controls	ols	
Tobacco chewing	n = 96	%	n = 57	%	OR (95% CI)	<i>p</i> -Value	
Genotype							
o CC	41	42.71	33	57.89	Reference	Reference	
o CT	49	51.04	22	38.6	1.79 (0.91–3.54)	0.09	
o TT	6	6.25	2	3.51	2.41 (0.46–12.76)	0.28	
o CT + TT	55	57.31	24	42.1	1.84 (0.95-3.58)	0.06	
Allele							
о С	131	68.23	88	77.19	Reference	Reference	
οΤ	61	31.77	26	22.81	1.58 (0.93-2.68)	0.09	
Smoking	n=36	%	n = 16	%	OR (95% CI)	<i>p</i> -Value	
Genotype							
o CC	11	30.56	9	56.25	Reference	Reference	
o CT	23	63.89	6	37.5	3.14 (0.89–11.04)	0.06	
o TT	2	5.55	1	6.25	1.64 (0.13-21.10)	0.7	
o CT + TT	25	69.45	7	43.75	2.92 (0.87-9.86)	0.07	
Allele							
ο С	45	62.5	24	75	Reference	Reference	
οΤ	27	37.5	8	25	1.80 (0.71-4.57)	0.21	
IL-1β +3953 C/T							
Tobacco chewing	n = 96	%	n = 57	%	OR (95% CI)	<i>p</i> -Value	
Genotype							
o CC	49	51.04	23	40.35	Reference	Reference	
o CT	39	40.62	33	57.89	0.55 (0.28-1.09)	0.08	
o TT	8	8.34	1	1.76	3.76 (0.44–31.82)	0.19	
o CT + TT	47	48.96	34	59.65	0.65 (0.33-1.26)	0.2	
Allele							
ο С	137	71.35	79	69.3	Reference	Reference	
οΤ	55	28.65	35	30.7	0.91 (0.55-1.50)	0.7	
Smoking	n=36	%	n = 16	%	OR (95% CI)	<i>p</i> -Value	
Genotype							
o CC	24	66.67	5	31.25	Reference	Reference	
o CT	11	30.56	10	62.5	0.23 (0.06-0.83)	0.2	
o TT	1	2.77	1	6.25	0.21 (0.01-3.92)	0.25	
o CT + TT	12	33.34	11	68.75	0.23 (0.06-0.80)	0.01	
Allele							
οС	59	81.95	20	62.5	Reference	Reference	
οΤ	13	18.05	12	37.5	0.37 (0.14-0.93)	0.03	

Abbreviations: CI, confidence interval; IL, interleukin; OR, odds ratio.

Table 4 Genotype and allele frequencies of interleukin-1 β –511C/T and +3953 C/T gene and their association with stages of cervical cancer and controls

IL-1β –511 C/T	Controls		Cervical cancer with stages I and II (%)		Odds ratio (95% CI)	<i>p</i> -Value	stage	er with	Odds ratio (95% CI)	<i>p</i> -Value
Genotype frequency	n = 200	%	n	%			n	%		
o CC	82	41	21	17.36	Reference	Reference	11	15.51	Reference	Reference
o CT	62	31	54	44.63	3.40 (1.86-6.21)	0.0005	33	46.47	3.97 (1.86–8.47)	0.0002
o TT	56	28	46	38.01	3.21 (1.73-5.95)	0.0001	27	38.02	3.59 (1.65–7.83)	0.0008
o CT+TT	118	59	100	82.64	3.31 (1.91–5.73)	0.0001	60	84.5	3.79 (1.88–7.65)	0.0001
Allele										
о С	226	56.5	96	39.67	Reference	Reference	55	38.73	Reference	Reference
οΤ	174	43.5	146	60.33	1.98 (1.43-2.73)	0.0003	87	61.27	2.05 (1.39–3.04)	0.0002
IL-1β +3953 C/T										
o CC	67	33.5	35	35.35	Reference	Reference	34	36.56	Reference	Reference
o CT	79	39.5	36	36.36	0.87 (0.49–1.54)	0.637	35	37.64	0.87 (0.49–1.55)	0.642
o TT	54	27	28	28.29	0.96 (0.52–1.77)	0.889	24	25.8	0.88 (0.46-1.65)	0.681
o CT + TT	133	61.5	64	64.65	0.92 (0.56–1.53)	0.75	59	63.44	0.87 (0.52–1.46)	0.608
Allele										
οС	213	53.25	106	53.54	Reference	Reference	103	55.37	Reference	Reference
οΤ	187	46.75	92	46.46	0.99 (0.70-1.39)	0.947	83	44.63	0.92 (0.65-1.30)	0.63

Abbreviations: CI, confidence interval; IL, interleukin.

and smokers as compared to controls, as presented in **Table 3**.

As shown in ►Table 4, a strong correlation between the genotypes of the IL-1 β –511C/T polymorphism and the clinical stage of CC patients who met FIGO criteria was confirmed. Individuals carrying C/T (OR, 3.40; 95% CI, 1.86-6.21; p = 0.0005), T/T (OR, 3.21; 95% CI, 1.73-5.95; p = 0.0001), and C/T + T/T genotypes (OR, 3.31; 95% CI, 1.91–5.73; p = 0.0001) along with T allele (OR, 1.98; 95% CI, 1.43-2.73; p = 0.0003) of IL-1 β –511C/T were significantly associated with stages I and II of CC. Similarly, C/T, T/T, and C/T + T/T genotypes and T allele of IL-1β –511C/T polymorphism were also significantly associated with stages III and IV: (OR, 3.97; 95% CI, 1.86-8.47; p = 0.0002), (OR 3.59; 95% CI, 1.65–7.83; p = 0.0008), and (OR 3.79; 95% CI, 1.88–7.65; p = 0.0001); and (OR 2.05; 95% CI, 1.39–3.04; p = 0.0002). However, there is no correlation between CC patients' clinical stage and the IL-1β +3953 C/T genotypes, as shown in **►Table 4**.

Discussion

CC is still a major public health issue in both developed and developing nations. CC is mostly caused by high-risk HPV, according to several prior epidemiological studies. ¹⁵ CC is widely recognized to be a multifaceted disease wherein genetic and environmental factors also play key roles in development. Environmental variables include lifestyle, exposure to tobacco-derived carcinogens, and kitchen smoke,

whereas genetic variables include inheritance of genetic traits or gene variations related to carcinogenesis. ¹⁶

Production of proinflammatory cytokines such as IL-18 and IL-1β triggers by multiprotein oligomer inflammasome. The elevated level and or expression of IL-1β may induce an inflammatory cascade that results in cancerrelated inflammation including chemotaxis, angiogenesis, and adhesiveness. Polymorphisms in cytokine genes can influence the immune response to HPV infection, altering the risk of CC development. Phe p53 gene degradation by the oncogenic protein HPV E6 is the most well-known carcinogenic pathway in CC. Inflammatory cytokines have also been found to cause DNA damage and prevent DNA repair.

Our study revealed that in IL-1 β –511C/T polymorphism, T allele was significantly linked with CC risk with up to twofold increase (p < 0.0001). The women with C/T and/or T/T genotype were strongly related with an increased CC risk of up to 3.60–3.34 folds (p < 0.001). Kang et al establish that carriers of the IL-1 β –511 C/T or T/T genotypes had a greater risk of CC with an OR of 2.42 in Korean women, which is consistent with our findings. Singh et al also have reported that women with T/T genotype of IL-1 β –511 C/T polymorphism were strongly linked with an increased CC risk in Indian population. Numerous studies have revealed the link between IL-1 β –511C/T gene polymorphism and cancers including gastric, liver, breast, and lung. However, the results remain inconclusive in different populations and various disease models.

Our study revealed that IL-1B -511C/T and T/T genotypes were associated with increased risk of CC as compared to controls (p < 0.001). As a result, genotypes of the IL-1β polymorphism may be linked to a higher reactivity to carcinogens (like HPV infection). Therefore, inherited genetic variations in the IL-1β gene may play a significant role in CC susceptibility, also indicating that IL-1B could potentially be a cause for CC. El-Omar et al reported that polymorphism in IL-1β -511C/T was associated with susceptibility to gastric cancer and also stated the T allele has been reported to be linked with increase in transcriptional activities.²⁸ So, due to its association with another polymorphism of the IL-1 locus, which can directly alter the IL-1 β gene expression, the -511C/T polymorphism may still be influencing cancer susceptibility. Wu et al revealed that T/T versus C/C, C/T versus C/C, and C/T + T/T versus C/C of IL-1β -511C/T polymorphism have considerably enhanced the risk of CC in the Asian group.²⁹ We contribute to these data, showing that susceptibility to CC risk linked to the C/T, T/T, and C/T + T/T genotypes becomes more significant in cases as compared to controls and also in tumor stage, which resembles with Wu et al. However, the findings of subgroup analysis revealed contrary effects of IL-1β -511C/T polymorphism with CC. According to Zidi et al, C/C genotype was associated with enhanced risk of development of CC among Tunisian women, while T/T genotype and T allele of IL-1β -511C/T polymorphism may be protective factor against CC development.³⁰ In contrast, findings of Al-Tahhan et al demonstrated that individuals carrying T/T genotype and T allele of IL-1β -511C/T polymorphism may be acknowledged as candidate biomarkers for CC development among Egyptian women.31

The results of our investigation showed a strong correlation between the polymorphism of IL-1 β –511C/T and vulnerability to CC. The allele analysis supported the connection between carrying the T allele and an elevated risk of CC in women with the C/T, T/T, and C/T+T/T genotypes. Our findings also showed the necessity to expand prevention efforts regarding the disease's relevance and promote gynecological follow-ups due to the high prevalence of and death due to CC. A potential strategy for determining CC risk is to create a susceptibility framework that includes both individual traits that raise the likelihood of the disease's occurrence and genetic biomarkers like the IL-1 β –511C/T polymorphism.

Conclusion

In summary, this is preliminary experimental research to investigate the relationship between putatively functional genetic variations of the IL-1 β gene and CC risk. Our findings add to the growing body of evidence linking IL-1 β gene variation to CC susceptibility. Our findings could shed fresh light on proinflammatory cytokines, inflammation, and the etiology of CC. Elevated serum levels are linked to a higher chance of developing CC as compared to controls.

Limitations of the Study

However, these findings should be interpreted with caution, and additional research with a bigger sample size and various demographics should be done to confirm our findings.

Authors' Contributions

Pushpendra D. Pratap conceived the study design, participated in data collection, and carried out the laboratory work. Syed Tasleem Raza, Ghazala Zaidi, and Shipra Kunwar revised the manuscript. Ale Eba and Muneshwar Rajput assisted for technical support. All authors read and approved the final manuscript.

Funding

This study was financially supported by the Department of Biotechnology, Ministry of Science and Technology, Government of India, New Delhi (Sanction Number - 6242-P110/RGCB/PMD/DBT/SHKR/2015).

Conflict of Interest

None declared.

References

- 1 Sung H, Ferlay J, Siegel RL, et al. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin 2021;71(03): 209–249
- 2 Muñoz N, Bosch FX, de Sanjosé S, et al; International Agency for Research on Cancer Multicenter Cervical Cancer Study Group. Epidemiologic classification of human papillomavirus types associated with cervical cancer. N Engl J Med 2003;348(06):518–527
- 3 Pratap P, Raza ST, Zaidi G, et al. Detection and quantitation of high risk human papillomavirus, HRHPV 16 and 18 in tissue of Indian women with cervical cancer: a case control study. EJPMR 2021;8 (03):551–557
- 4 Pratap P, Raza ST, Zaidi G, et al. Molecular biology of human papillomavirus, cervical carcinoma and its management. Can J Clin Nutr 2021;9(01):71–88
- 5 American Cancer Society. Cancer facts and figures. Atlanta, GA: American Cancer Society; 2015. http://old.cancer.org/acs/ groups/content/@editorial/documents/document/acspc-044552. pdf Accessed October 30, 2023
- 6 Castellsagué X, Bosch FX, Muñoz N. Environmental co-factors in HPV carcinogenesis. Virus Res 2002;89(02):191–199
- 7 Mehta AM, Mooij M, Branković I, Ouburg S, Morré SA, Jordanova ES. Cervical carcinogenesis and immune response gene polymorphisms: a review. J Immunol Res 2017;2017:8913860
- 8 Sims JE, Smith DE. The IL-1 family: regulators of immunity. Nat Rev Immunol 2010;10(02):89–102
- 9 Arend WP, Malyak M, Guthridge CJ, Gabay C. Interleukin-1 receptor antagonist: role in biology. Annu Rev Immunol 1998; 16:27–55
- 10 Lewis AM, Varghese S, Xu H, Alexander HR. Interleukin-1 and cancer progression: the emerging role of interleukin-1 receptor antagonist as a novel therapeutic agent in cancer treatment. J Transl Med 2006;4:48
- 11 Ban JY, Kim MK, Park SW, Kwon KH. Interleukin-1 beta polymorphisms are associated with lymph node metastasis in Korean patients with papillary thyroid carcinoma. Immunol Invest 2012; 41(08):888–905

- 12 Xu H, Ding Q, Jiang HW. Genetic polymorphism of interleukin-1A (IL-1A), IL-1B, and IL-1 receptor antagonist (IL-1RN) and prostate cancer risk. Asian Pac J Cancer Prev 2014;15(20):8741–8747
- 13 Achyut BR, Srivastava A, Bhattacharya S, Mittal B. Genetic association of interleukin-1beta (-511C/T) and interleukin-1 receptor antagonist (86 bp repeat) polymorphisms with Type 2 diabetes mellitus in North Indians. Clin Chim Acta 2007;377(1-2):163–169
- 14 Bioque G, Crusius JB, Koutroubakis I, et al. Allelic polymorphism in IL-1 beta and IL-1 receptor antagonist (IL-1Ra) genes in inflammatory bowel disease. Clin Exp Immunol 1995;102(02):379–383
- 15 Crossley B, Crossley J. A review of the use of human papilloma virus (HPV) in cervical screening. Br J Biomed Sci 2017;74(03):105–109
- 16 Velema JP, Ferrera A, Figueroa M, et al. Burning wood in the kitchen increases the risk of cervical neoplasia in HPV-infected women in Honduras. Int J Cancer 2002;97(04):536–541
- 17 Kantono M, Guo B. Inflammasomes and cancer: the dynamic role of the inflammasome in tumor development. Front Immunol 2017;8:1132
- 18 Idris A, Ghazali NB, Koh D. Interleukin 1β-A potential salivary biomarker for cancer progression? Biomark Cancer 2015;7:25–29
- 19 Deshpande A, Nolan JP, White PS, et al. TNF-alpha promoter polymorphisms and susceptibility to human papillomavirus 16associated cervical cancer. J Infect Dis 2005;191(06):969–976
- 20 Fu L, Van Doorslaer K, Chen Z, et al. Degradation of p53 by human Alphapapillomavirus E6 proteins shows a stronger correlation with phylogeny than oncogenicity. PLoS One 2010;5(09):e12816
- 21 Jaiswal M, LaRusso NF, Burgart LJ, Gores GJ. Inflammatory cytokines induce DNA damage and inhibit DNA repair in cholangiocarcinoma cells by a nitric oxide-dependent mechanism. Cancer Res 2000;60(01):184–190
- 22 Kang S, Kim JW, Park NH, et al. Interleukin-1 beta-511 polymorphism and risk of cervical cancer. J Korean Med Sci 2007;22(01):110–113

- 23 Singh H, Sachan R, Goel H, Mittal B. Genetic variants of interleukin-1RN and interleukin-1beta genes and risk of cervical cancer. BJOG 2008;115(05):633-638
- 24 Lee KA, Ki CS, Kim HJ, et al. Novel interleukin 1beta polymorphism increased the risk of gastric cancer in a Korean population. J Gastroenterol 2004;39(05):429–433
- 25 Hirankarn N, Kimkong I, Kummee P, Tangkijvanich P, Poovorawan Y. Interleukin-1beta gene polymorphism associated with hepatocellular carcinoma in hepatitis B virus infection. World J Gastroenterol 2006;12(05):776–779
- 26 Liu J, Zhai X, Jin G, et al. Functional variants in the promoter of interleukin-1beta are associated with an increased risk of breast cancer: a case-control analysis in a Chinese population. Int J Cancer 2006;118(10):2554–2558
- 27 Zienolddiny S, Ryberg D, Maggini V, Skaug V, Canzian F, Haugen A. Polymorphisms of the interleukin-1 beta gene are associated with increased risk of non-small cell lung cancer. Int J Cancer 2004;109 (03):353–356
- 28 El-Omar EM, Carrington M, Chow WH, et al. Interleukin-1 polymorphisms associated with increased risk of gastric cancer. Nature 2000;404(6776):398–402
- 29 Wu S, Hu G, Chen J, Xie G. Interleukin 1β and interleukin 1 receptor antagonist gene polymorphisms and cervical cancer: a meta-analysis. Int J Gynecol Cancer 2014;24(06):984–990
- 30 Zidi S, Sghaier I, Zouidi F, et al. Interleukin-1 gene cluster polymorphisms and its haplotypes may predict the risk to develop cervical cancer in Tunisia. Pathol Oncol Res 2015;21(04): 1101–1107
- 31 Al-Tahhan MA, Etewa RL, El Behery MM. Association between circulating interleukin-1 beta (IL-1β) levels and IL-1β C-511T polymorphism with cervical cancer risk in Egyptian women. Mol Cell Biochem 2011;353(1-2):159–165