Role of Molecular Targeted Therapeutic Drugs in Treatment of Glioblastoma: A Review Article

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Abstract

Glioblastoma is remarkably periodic primary brain tumor, characterizing an eminently heterogeneous pattern of neoplasms that are utmost destructive and threatening cancers.

An enhanced and upgraded knowledge of the various molecular pathways that cause malignant changes in glioblastoma has resulted in advancement of numerous biomarkers and the interpretation of various agents that pointedly target tumor cells and microenvironment. In this review, literature or information on various targeted therapy for glioblastoma is discussed. English language articles were scrutinized in plentiful directory or databases like PubMed, ScienceDirect, Web of Sciences, Google Scholar, and Scopus. The important keywords used for searching databases are "Glioblastoma," "Targeted therapy in glioblastoma," "Therapeutic drugs in glioblastoma," and "Molecular targets in glioblastoma."

Keywords
► glioblastoma
► targeted therapy
► molecular targets
► therapeutic drugs

Introduction

Glioblastoma is the utmost common and destructive primary malignant brain tumor seen in adults including average overall survival (OS) of 10 to 20 months.[1–4] Glioblastoma comprises an eminently heterogeneous collection of protruding malignant tumors of the brain.[5]

In a nutshell, the abovementioned research demonstrated that nearly all tumors suppress periodic molecular modifications eradicating core pathways engaged in the control of growth and deoxyribonucleic acid repair. It is acknowledged that glioblastomas are described by considerable intratumor and intertumor genomic heterogeneity.[6–10] Depending upon the findings of the Cancer Genome Atlas, there are four distinctive subdivisions of glioblastomas. These are the neural, proneural, mesenchymal, and classical subtypes. The neural subdivision illustrates 16% of glioblastoma. The neural subdivision is represented by the expression of various neuron markers like GABRA1, SLC1A5, and NEFL. The proneural subdivision demonstrates mutation in platelet-derived growth factor receptor A (PDGFRA). The classical subdivision demonstrated CDKN2A deletion and epidermal growth factor receptor (EGFR) amplification. The mesenchymal subdivision demonstrates mutations in phosphatase and tensin homolog (PTEN) and NF1 (neurofibromatosis type 1).[11]

In this review, literature or information on various targeted therapy for glioblastoma is discussed. English language articles were scrutinized in plentiful directory or databases like PubMed, ScienceDirect, Web of Sciences, Google Scholar, and Scopus. The important keywords used for searching databases are "Glioblastoma," "Targeted therapy in glioblastoma," "Therapeutic drugs in glioblastoma," and "Molecular targets in glioblastoma."

Various Molecular Targeted Therapeutics for Glioblastoma

Receptor Tyrosine Kinases

They are types of transmembrane proteins. It contains a single transmembrane helix, extracellular ligand-binding domain, and intracellular catalytic domain. The receptor tyrosine kinase group consists of platelet-derived growth factors, fibroblast growth factor receptors (FGFRs), EGFRs, and hepatocyte growth factor receptors. Beneath typical physiological status, receptor tyrosine kinases are implicated in persisting cellular homeostasis by controlling cell–cell communication, cell proliferation, survival, differentiation, and migration. Therefore, dysregulation of the receptor tyrosine kinases pathway performs a crucial aspect in the initiation and progression of glioblastoma.[12–14]

Epidermal Growth Factor Receptor

Genomic interpretation identified that 57% of glioblastoma cells harbor EGFR genetic mutations. Overexpression and amplification of EGFRs were recognized in 60 and 40% of cases of primary glioblastoma accordingly. Overexpression and amplification result in fundamental receptor activation and intensify the survival, proliferation, and resistance to therapeutics of glioblastoma cells.[15–18] Different forms of genetic mutation were also recognized which include point mutations and rearrangement of EGFRs.[19]

The utmost prevalent approach for targeting EGFRs is by way of the adoption of monoclonal antibodies. Various anti-EGFR antibodies have been established since cetuximab (the first chimeric antibody). Cetuximab and panitumumab do not demonstrate encouraging outcomes. Depatuxizumab and nimotuzumab demonstrate survival advantages when mixed with radiotherapy and chemotherapeutic temozolomide (TMZ) accordingly.[20–22]

EGFRs are also aimed by prohibitions of the activity of tyrosine kinase. Different inhibitors have been graded in clinical research with the least possible or no advantage like gefitinib, erlotinib, and dacomitinib. Nonetheless, utilizing afatinib leads to an upsurge in progression-free survival (PFS) in those individuals that demonstrate overexpression of EGFRs.[23]

PDGFR

PDGFR is one of the targeted therapeutics in the glioblastoma–proneural subdivision. Gene amplification in PDGFR is observed in 15% of cases of glioblastoma. In different grades of gliomas, overexpression of PDGFR is observed and is linked with poor prognosis. Until now, various multikinase inhibitors like imatinib, sunitinib, and dasatinib have not demonstrated encouraging clinical advantages.[24,25]

MET

The hepatocyte growth factor receptor is encoded by the MET gene, which is expected to perform an influential function in the invasion, recurrence, migration, and drug resistance of glioma cells.[26,27] Approximately 30% of glioblastoma patients are represented by overexpression of MET.

The usefulness of the rilotumumab antibody only had no action on restricting the advancement of glioblastoma. Clinical research of integrated antibody onartuzumab and anti-vascular drugs proved that there was no meaningful advantage for recurrent glioblastoma patients. Cabozantinib, an MET inhibitor, was moderately active in individuals with recurrent glioblastoma.[27–31]

PI3K/AKT/mTOR Pathway

It is the utmost prevalent alteration pathway in individuals with glioblastoma. PI3K activation in glioblastoma is chiefly because of the alteration of PTEN.[32,33]

Buparlisib, a PI3K pan inhibitor, was also demonstrated to be incompetent in contrast to recurrent glioblastoma in research, either as an individual dose or linked with lomustine or carboplatin.[34,35]

Fibroblast Growth Factor Receptor

It is comprehensively expressed in glioblastoma, but its therapeutic worth may be confined to the limited count of individuals with FGFR-TACC fusion. In the current research, utilization of dovitinib was incompetent in increasing the survival of individuals whether linked with antivascular therapy or not.[36–38]

BRAF Mutation

BRAF takes part in Mek/Erk pathway activation and encourages the proliferation of the cell. BRAF alteration is noticed in different varieties of cancer and is demonstrated to be a trustworthy target.[39–42]

Neurotrophic Tyrosine Receptor Kinases

Three distinctive genes encode the neurotrophic tyrosine receptor kinases (NTRKs). These genes are NTRK3, NTRK2, and NTRK1. The NTRK gene genomic rearrangement results in the union of the gene, which may provoke the TRK pathway activation. This gene fusion occurrence is rarely seen in glioblastoma. Entrectinib was competent in the therapeutics of infantile glioblastoma. Larotrectinib was administered in a lady with infantile glioblastoma and the therapeutic result was noteworthy.[43–46]

The Retinoblastoma Pathway

The cell cycle regulation of the retinoblastoma protein (pRB) pathway is reciprocated because of CDK4/6 amplification, CDKN2A/B homozygous deletion, and modification of the RB1 gene. In phase II research, palbociclib has shown an unsatisfying outcome. Ribociclib was also incompetent.[47–49]

Proteasome

Proteasome encourage apoptosis by controlling p53, which alarmingly controls the cell cycle and alters drug resistance. Presently, various clinically recognized proteasome inhibitors include ixazomib, bortezomib, and marizomib. Bortezomib when combined with vorinostat shows inadequate results in recurrent glioblastoma. But when bortezomib is mixed with definitive radiotherapy, it shows hopeful survival rates and is well accepted. Disulfiram has advantageous

blood–brain barrier penetration competence and improved drug resistance to utilize its antitumor outcome in recently diagnosed glioblastoma and recurrent glioblastoma models.[50–54]

Vascular Endothelial Growth Factor

Glioblastoma is described by irregularity in vascular proliferation. The vascular endothelial growth factor (VEGF) is eminently expressed in glioblastoma and advocates the anomalous proliferation of tumors. VEGFR1 and VEGFR2 pathways are recommended as a significant determinant of tumor survival in glioblastoma.[55] Bevacizumab is attached to endothelial cells and suppresses angiogenesis. In phase II research, bevacizumab demonstrates meaningful anti-glioma and biological activity, increased OS, and radiation response rate in recently diagnosed cases of glioblastoma and recurrent glioblastoma. It is also observed that bevacizumab in phase III clinical research substantially enhances PFS.[56–60]

Bevacizumab along with TMZ demonstrates great competence and resistance. Etoposide shows an identical outcome to bevacizumab monotherapy, but etoposide displays higher toxicity.[61–63] Additional VEGF such as cediranib has demonstrated meaningful potency in phase II clinical research of recurrent glioblastoma. It is observed that cediranib advocates blood perfusion and extended the OS in recently diagnosed cases of glioblastoma.[64,65]

Axitinib could be a promising consolidation ally with immunotherapy. Additional inhibitors such as aflibercept also downregulate the VEGF activity.[66,67]

Integrin

Integrins perform in signal transduction participating in various cellular processes. Integrins also arbitrate cellular transmission inside the extracellular matrix throughout motility, invasion, migration, angiogenesis, and adhesion. In endothelial cells, integrins αvβ5 and αvβ3 are eminently expressed and recognized as therapeutic targets in glioblastoma.[68,69]

Programmed Cell Death Protein 1

One approach to cancer immunotherapy is to forbid the communication among programmed cell death protein 1 (PD-1) on T cells and PD-1 ligand on host or tumor cells. Pembrolizumab has inadequate effectiveness in earlier therapeutics of glioblastoma, exclusive of those cases with definitive mismatch repair defects.[70–73] Nivolumab, when mixed with bevacizumab and chemoradiotherapy in recently diagnosed glioblastoma individuals along with MGMT promoter unmethylation, was unsuccessful.[74]

Lymphocyte-Activation Gene 3

Lymphocyte-activation gene 3 (LAG-3) results in an immune outbreak of tumor cells. LAG-3 is mainly seen in activated immune cells. LAG-3 is consistently expressed in T cells. Therefore, LAG-3 prohibitor evolves to a pleasant immune modulating agent only or in association with additional immune checkpoint inhibitors. In glioblastoma, LAG-3 is expressed along with CD8A, suggesting that LAG-3 targeted therapy in glioblastoma with sufficient CD8+ T cell infiltration may be hopeful.[75–79]

CD73

The nasal application of cationic nanoemulsion when blended with CD73-siRNA conferred hopeful anti-CD33 outcomes in glioblastoma model.[80]

V-Domain Immunoglobulin Suppressor of T Cell Activation

It has been originally acknowledged for its meaningful appearance in immunosuppression. V-domain immunoglobulin suppressor of T cell activation (VISTA) complexly and reciprocally perform as ligand and receptor in the positive and negative control of cancer immunity.[81–83] IgSF11 (immunoglobulin superfamily 11 gene), a VISTA ligand, demonstrates raised expression notably in high-grade glioma and corresponds with poor prognosis, implying the promising prognostic significance of IgSF11 and VISTA.[84]

CD70

CD70 is eminently overexpressed in cells of recurrent glioma in comparison to ordinary tissue and is linked with inadequate survival. Therefore, CD70 is suggested to bring about T cell apoptosis or debilitation and initiate regulatory T cells to intercede immunosuppression.[85,86]

Tumor-Associated Macrophage Therapy

Minocycline could restrain the expression of microglial matrix metalloproteinases and weaken the glioma intrusion. In addition, cyclosporine A demonstrated effectiveness in debilitating the angiogenesis and survival of glioma by restraining the microglia infiltration. Propentofylline was also demonstrated to lower the growth of tumors in glioblastoma by precisely targeting microglia.[87–90]

Conclusion

The prediction of glioblastoma stays worse and poor regardless of radiotherapy, aggressive surgery, and chemotherapies. Furthermore, numerous innovative introductions in elementary and translational researches were made in recent times. Various targeted therapies are being extensively investigated in various clinical researches. Promising advancement in glioblastoma therapeutics will apparently depend on collection of correct association of various targeted agents collectively with different multimodal therapy.

Conflict of Interest
None declared.

References

1 Ostrom QT, Gittleman H, Liao P, et al. CBTRUS statistical report: primary brain and central nervous system tumors diagnosed in the United States in 2007-2011. Neuro-oncol 2014;16 Suppl 4 (Suppl 4):iv1–iv63

2 Stupp R, Mason WP, van den Bent MJ, et al; European Organisation for Research and Treatment of Cancer Brain Tumor and Radiotherapy Groups National Cancer Institute of Canada Clinical Trials Group. Radiotherapy plus concomitant and adjuvant temozolomide for glioblastoma. N Engl J Med 2005;352(10):987–996

3 Rønning PA, Helseth E, Meling TR, Johannesen TB. A population-based study on the effect of temozolomide in the treatment of glioblastoma multiforme. Neuro-oncol 2012;14(09):1178–1184

4 Stupp R, Taillibert S, Kanner AA, et al. Maintenance therapy with tumor-treating fields plus temozolomide vs temozolomide alone for glioblastoma: a randomized clinical trial. JAMA 2015;314(23): 2535–2543

5 Louis DN, Perry A, Reifenberger G, et al. The 2016 World Health Organization classification of tumors of the central nervous system: a summary. Acta Neuropathol 2016;131(06):803–820

6 Snuderl M, Fazlollahi L, Le LP, et al. Mosaic amplification of multiple receptor tyrosine kinase genes in glioblastoma. Cancer Cell 2011;20(06):810–817

7 Sottoriva A, Spiteri I, Piccirillo SG, et al. Intratumor heterogeneity in human glioblastoma reflects cancer evolutionary dynamics. Proc Natl Acad Sci U S A 2013;110(10):4009–4014

8 Johnson BE, Mazor T, Hong C, et al. Mutational analysis reveals the origin and therapy-driven evolution of recurrent glioma. Science 2014;343(6167):189–193

9 Kim H, Zheng S, Amini SS, et al. Whole-genome and multisector exome sequencing of primary and post-treatment glioblastoma reveals patterns of tumor evolution. Genome Res 2015;25(03): 316–327

10 Kim J, Lee IH, Cho HJ, et al. Spatiotemporal evolution of the primary glioblastoma genome. Cancer Cell 2015;28(03):318–328

11 Verhaak RGW, Hoadley KA, Purdom E, et al; Cancer Genome Atlas Research Network. Integrated genomic analysis identifies clinically relevant subtypes of glioblastoma characterized by abnormalities in PDGFRA, IDH1, EGFR, and NF1. Cancer Cell 2010;17(01):98–110

12 Hubbard SR. Structural analysis of receptor tyrosine kinases. Prog Biophys Mol Biol 1999;71(3-4):343–358

13 Montor WR, Salas AROSE, Melo FHM. Receptor tyrosine kinases and downstream pathways as druggable targets for cancer treatment: the current arsenal of inhibitors. Mol Cancer 2018;17(01): 55

14 Blume-Jensen P, Hunter T. Oncogenic kinase signalling. Nature 2001;411(6835):355–365

15 Brennan CW, Verhaak RGW, McKenna A, et al; TCGA Research Network. The somatic genomic landscape of glioblastoma. Cell 2013;155(02):462–477

16 Chakravarti A, Chakladar A, Delaney MA, Latham DE, Loeffler JS. The epidermal growth factor receptor pathway mediates resistance to sequential administration of radiation and chemotherapy in primary human glioblastoma cells in a RAS-dependent manner. Cancer Res 2002;62(15):4307–4315

17 Mazzoleni S, Politi LS, Pala M, et al. Epidermal growth factor receptor expression identifies functionally and molecularly distinct tumor-initiating cells in human glioblastoma multiforme and is required for gliomagenesis. Cancer Res 2010;70(19): 7500–7513

18 Li L, Dutra A, Pak E, et al. EGFRvIII expression and PTEN loss synergistically induce chromosomal instability and glial tumors. Neuro-oncol 2009;11(01):9–21

19 Wen PY, Kesari S. Malignant gliomas in adults. N Engl J Med 2008; 359(05):492–507

20 Cruz Da Silva E, Mercier MC, Etienne-Selloum N, Dontenwill M, Choulier L. A systematic review of glioblastoma- targeted therapies in phases II, III, IV clinical trials. Cancers (Basel) 2021;13(08): 1795

21 Van Den Bent M, Eoli M, Sepulveda JM, et al. INTELLANCE 2/EORTC 1410 randomized phase II study of Depatux-M alone and with temozolomide vs temozolomide or lomustine in recurrent EGFR amplified glioblastoma. Neuro-oncol 2020;22(05):684–693

22 Solomon MT, Miranda N, Jorrín E, et al. Nimotuzumab in combination with radiotherapy in high grade glioma patients: a single institution experience. Cancer Biol Ther 2014;15(05):504–509

23 Reardon DA, Nabors LB, Mason WP, et al; BI 1200 36 Trial Group and the Canadian Brain Tumour Consortium. Phase I/randomized phase II study of afatinib, an irreversible ErbB family blocker, with or without protracted temozolomide in adults with recurrent glioblastoma. Neuro-oncol 2015;17(03):430–439

24 Plate KH, Breier G, Farrell CL, Risau W. Platelet-derived growth factor receptor-beta is induced during tumor development and upregulated during tumor progression in endothelial cells in human gliomas. Lab Invest 1992;67(04):529–534

25 Camorani S, Esposito CL, Rienzo A, et al. Inhibition of receptor signaling and of glioblastoma-derived tumor growth by a novel PDGFRβ aptamer. Mol Ther 2014;22(04):828–841

26 Cheng F, Guo D. MET in glioma: signaling pathways and targeted therapies. J Exp Clin Cancer Res 2019;38(01):270

27 Xie Q, Bradley R, Kang L, et al. Hepatocyte growth factor (HGF) autocrine activation predicts sensitivity to MET inhibition in glioblastoma. Proc Natl Acad Sci U S A 2012;109(02):570–575

28 Wen PY, Schiff D, Cloughesy TF, et al. A phase II study evaluating the efficacy and safety of AMG 102 (rilotumumab) in patients with recurrent glioblastoma. Neuro-oncol 2011;13(04):437–446

29 Cloughesy T, Finocchiaro G, Belda-Iniesta C, et al. Randomized, double-blind, placebo-controlled, multicenter phase II study of onartuzumab plus bevacizumab versus placebo plus bevacizumab in patients with recurrent glioblastoma: efficacy, safety, and hepatocyte growth factor and O^6-methylguanine-DNA methyltransferase biomarker analyses. J Clin Oncol 2017;35(03): 343–351

30 Wen PY, Drappatz J, de Groot J, et al. Phase II study of cabozantinib in patients with progressive glioblastoma: subset analysis of patients naive to antiangiogenic therapy. Neuro-oncol 2018;20 (02):249–258

31 Cloughesy TF, Drappatz J, de Groot J, et al. Phase II study of cabozantinib in patients with progressive glioblastoma: subset analysis of patients with prior antiangiogenic therapy. Neuro-oncol 2018;20(02):259–267

32 Hoxhaj G, Manning BD. The PI3K-AKT network at the interface of oncogenic signalling and cancer metabolism. Nat Rev Cancer 2020;20(02):74–88

33 Zhao H-F, Wang J, Shao W, et al. Recent advances in the use of PI3K inhibitors for glioblastoma multiforme: current preclinical and clinical development. Mol Cancer 2017;16(01):100

34 Wen PY, Touat M, Alexander BM, et al. Buparlisib in patients with recurrent glioblastoma harboring phosphatidylinositol 3-kinase pathway activation: an open-label, multicenter, multi-arm, phase II trial. J Clin Oncol 2019;37(09):741–750

35 Rosenthal M, Clement PM, Campone M, et al. Buparlisib plus carboplatin or lomustine in patients with recurrent glioblastoma: a phase Ib/II, open-label, multicentre, randomised study. ESMO Open 2020;5(04):e000672

36 Singh D, Chan JM, Zoppoli P, et al. Transforming fusions of FGFR and TACC genes in human glioblastoma. Science 2012;337 (6099):1231–1235

37 Di Stefano AL, Fucci A, Frattini V, et al. Detection, characterization, and inhibition of FGFR-TACC Fusions in IDH wild-type glioma. Clin Cancer Res 2015;21(14):3307–3317

38 Sharma M, Schilero C, Peereboom DM, et al. Phase II study of dovitinib in recurrent glioblastoma. J Neurooncol 2019;144(02): 359–368

39 Planchard D, Besse B, Groen HJM, et al. Dabrafenib plus trametinib in patients with previously treated BRAF(V600E)-mutant metastatic non-small cell lung cancer: an open-label, multicentre phase 2 trial. Lancet Oncol 2016;17(07):984–993

40 Brose MS, Cabanillas ME, Cohen EEW, et al. Vemurafenib in patients with BRAF(V600E)-positive metastatic or unresectable papillary thyroid cancer refractory to radioactive iodine: a non-randomised, multicentre, open-label, phase 2 trial. Lancet Oncol 2016;17(09):1272–1282

41 Subbiah V, Kreitman RJ, Wainberg ZA, et al. Dabrafenib and trametinib treatment in patients with locally advanced or metastatic BRAF V600-mutant anaplastic thyroid cancer. J Clin Oncol 2018;36(01):7–13

42 Robert C, Grob JJ, Stroyakovskiy D, et al. Five-year outcomes with dabrafenib plus trametinib in metastatic melanoma. N Engl J Med 2019;381(07):626–636

43 Woo HY, Na K, Yoo J, et al. Glioblastomas harboring gene fusions detected by next-generation sequencing. Brain Tumor Pathol 2020;37(04):136–144

44 Ferguson SD, Zhou S, Huse JT, et al. Targetable gene fusions associate with the IDH wild-type astrocytic lineage in adult gliomas. J Neuropathol Exp Neurol 2018;77(06):437–442

45 Alharbi M, Mobark NA, Balbaid AAO, et al. Regression of ETV6-NTRK3 infantile glioblastoma after first-line treatment with larotrectinib. JCO Precis Oncol 2020;4:PO.20.00017

46 Ku DT-L, Shing MM-K, Chan GC-F, et al. HGG-48. ROS1 inhibitor entrectinib use in relapse/ refractory infantile glioblastoma with positive ROS1 fusion - a case report with promising response. Neuro-oncol 2020;22(Supplement_3):iii352–iii352

47 Taylor JW, Parikh M, Phillips JJ, et al. Phase-2 trial of palbociclib in adult patients with recurrent RB1-positive glioblastoma. J Neuro-oncol 2018;140(02):477–483

48 Miller TW, Traphagen NA, Li J, et al. Tumor pharmacokinetics and pharmacodynamics of the CDK4/6 inhibitor ribociclib in patients with recurrent glioblastoma. J Neurooncol 2019;144(03):563–572

49 Tien AC, Li J, Bao X, et al. A phase 0 trial of ribociclib in recurrent glioblastoma patients incorporating a tumor pharmacodynamic- and pharmacokinetic-guided expansion cohort. Clin Cancer Res 2019;25(19):5777–5786

50 Goldberg AL. Protein degradation and protection against misfolded or damaged proteins. Nature 2003;426(6968):895–899

51 Narayanan S, Cai C-Y, Assaraf YG, et al. Targeting the ubiquitin-proteasome pathway to overcome anti-cancer drug resistance. Drug Resist Updat 2020;48:100663

52 Friday BB, Anderson SK, Buckner J, et al. Phase II trial of vorinostat in combination with bortezomib in recurrent glioblastoma: a north central cancer treatment group study. Neuro-oncol 2012;14(02):215–221

53 Kong XT, Nguyen NT, Choi YJ, et al. Phase 2 study of bortezomib combined with temozolomide and regional radiation therapy for upfront treatment of patients with newly diagnosed glioblastoma multiforme: safety and efficacy assessment. Int J Radiat Oncol Biol Phys 2018;100(05):1195–1203

54 Huang J, Campian JL, Gujar AD, et al. Final results of a phase I dose-escalation, dose-expansion study of adding disulfiram with or without copper to adjuvant temozolomide for newly diagnosed glioblastoma. J Neurooncol 2018;138(01):105–111

55 Szabo E, Schneider H, Seystahl K, et al. Autocrine VEGFR1 and VEGFR2 signaling promotes survival in human glioblastoma models in vitro and in vivo. Neuro-oncol 2016;18(09):1242–1252

56 Ferrara N, Hillan KJ, Novotny W. Bevacizumab (Avastin), a humanized anti-VEGF monoclonal antibody for cancer therapy. Biochem Biophys Res Commun 2005;333(02):328–335

57 Friedman HS, Prados MD, Wen PY, et al. Bevacizumab alone and in combination with irinotecan in recurrent glioblastoma. J Clin Oncol 2009;27(28):4733–4740

58 Kreisl TN, Kim L, Moore K, et al. Phase II trial of single-agent bevacizumab followed by bevacizumab plus irinotecan at tumor progression in recurrent glioblastoma. J Clin Oncol 2009;27(05):740–745

59 Raizer JJ, Grimm S, Chamberlain MC, et al. A phase 2 trial of single-agent bevacizumab given in an every-3-week schedule for patients with recurrent high-grade gliomas. Cancer 2010;116(22):5297–5305

60 Gilbert MR, Dignam JJ, Armstrong TS, et al. A randomized trial of bevacizumab for newly diagnosed glioblastoma. N Engl J Med 2014;370(08):699–708

61 Gilbert MR, Pugh SL, Aldape K, et al. NRG oncology RTOG 0625: a randomized phase II trial of bevacizumab with either irinotecan or dose-dense temozolomide in recurrent glioblastoma. J Neuro-oncol 2017;131(01):193–199

62 Sathornsumetee S, Desjardins A, Vredenburgh JJ, et al. Phase II trial of bevacizumab and erlotinib in patients with recurrent malignant glioma. Neuro-oncol 2010;12(12):1300–1310

63 Reardon DA, Desjardins A, Vredenburgh JJ, et al. Metronomic chemotherapy with daily, oral etoposide plus bevacizumab for recurrent malignant glioma: a phase II study. Br J Cancer 2009;101(12):1986–1994

64 Batchelor TT, Duda DG, di Tomaso E, et al. Phase II study of cediranib, an oral pan-vascular endothelial growth factor receptor tyrosine kinase inhibitor, in patients with recurrent glioblastoma. J Clin Oncol 2010;28(17):2817–2823

65 Batchelor TT, Gerstner ER, Emblem KE, et al. Improved tumor oxygenation and survival in glioblastoma patients who show increased blood perfusion after cediranib and chemoradiation. Proc Natl Acad Sci U S A 2013;110(47):19059–19064

66 Du Four S, Maenhout SK, Benteyn D, et al. Disease progression in recurrent glioblastoma patients treated with the VEGFR inhibitor axitinib is associated with increased regulatory T cell numbers and T cell exhaustion. Cancer Immunol Immunother 2016;65(06):727–740

67 de Groot JF, Piao Y, Tran H, et al. Myeloid biomarkers associated with glioblastoma response to anti-VEGF therapy with aflibercept. Clin Cancer Res 2011;17(14):4872–4881

68 Schnell O, Krebs B, Carlsen J, et al. Imaging of integrin alpha(v)beta(3) expression in patients with malignant glioma by [18F] Gal-acto-RGD positron emission tomography. Neuro-oncol 2009;11(06):861–870

69 Mikkelsen T, Brodie C, Finniss S, et al. Radiation sensitization of glioblastoma by cilengitide has unanticipated schedule-dependency. Int J Cancer 2009;124(11):2719–2727

70 Blumenthal DT, Yalon M, Vainer GW, et al. Pembrolizumab: first experience with recurrent primary central nervous system (CNS) tumors. J Neurooncol 2016;129(03):453–460

71 Bouffet E, Larouche V, Campbell BB, et al. Immune checkpoint inhibition for hypermutant glioblastoma multiforme resulting from germline biallelic mismatch repair deficiency. J Clin Oncol 2016;34(19):2206–2211

72 Johanns TM, Miller CA, Dorward IG, et al. Immunogenomics of hypermutated glioblastoma: a patient with germline POLE deficiency treated with checkpoint blockade immunotherapy. Cancer Discov 2016;6(11):1230–1236

73 Lukas RV, Rodon J, Becker K, et al. Clinical activity and safety of atezolizumab in patients with recurrent glioblastoma. J Neuro-oncol 2018;140(02):317–328

74 Reardon DA, Brandes AA, Omuro A, et al. Effect of nivolumab vs bevacizumab in patients with recurrent glioblastoma: the Check-Mate 143 phase 3 randomized clinical trial. JAMA Oncol 2020;6(07):1003–1010

75 Workman CJ, Rice DS, Dugger KJ, Kurschner C, Vignali DAA. Phenotypic analysis of the murine CD4-related glycoprotein, CD223 (LAG-3). Eur J Immunol 2002;32(08):2255–2263

76 Triebel F, Jitsukawa S, Baixeras E, et al. LAG-3, a novel lymphocyte activation gene closely related to CD4. J Exp Med 1990;171(05):1393–1405

77 Maruhashi T, Sugiura D, Okazaki I-M, Okazaki T. LAG-3: from molecular functions to clinical applications. J Immunother Cancer 2020;8(02):e001014

78 Harris-Bookman S, Mathios D, Martin AM, et al. Expression of LAG-3 and efficacy of combination treatment with anti-LAG-3 and anti-PD-1 monoclonal antibodies in glioblastoma. Int J Cancer 2018;143(12):3201–3208

79 Mair MJ, Kiesel B, Feldmann K, et al. LAG-3 expression in the inflammatory microenvironment of glioma. J Neurooncol 2021; 152(03):533–539

80 Azambuja JH, Schuh RS, Michels LR, et al. Nasal administration of cationic nanoemulsions as CD73-siRNA delivery system for glioblastoma treatment: a new therapeutical approach. Mol Neurobiol 2020;57(02):635–649

81 Wang L, Rubinstein R, Lines JL, et al. VISTA, a novel mouse Ig superfamily ligand that negatively regulates T cell responses. J Exp Med 2011;208(03):577–592

82 Huang X, Zhang X, Li E, et al. VISTA: an immune regulatory protein checking tumor and immune cells in cancer immunotherapy. J Hematol Oncol 2020;13(01):83

83 Flies DB, Han X, Higuchi T, et al. Coinhibitory receptor PD-1H preferentially suppresses CD4+ T cell-mediated immunity. J Clin Invest 2014;124(05):1966–1975

84 Ghouzlani A, Rafii S, Karkouri M, Lakhdar A, Badou A. The Promising IgSF11 immune checkpoint is highly expressed in advanced human gliomas and associates to poor prognosis. Front Oncol 2021;10:608609

85 Wischhusen J, Jung G, Radovanovic I, et al. Identification of CD70-mediated apoptosis of immune effector cells as a novel immune escape pathway of human glioblastoma. Cancer Res 2002;62(09):2592–2599

86 Jin L, Ge H, Long Y, et al. CD70, a novel target of CAR T-cell therapy for gliomas. Neuro-oncol 2018;20(01):55–65

87 Hu F, Ku M-C, Markovic D, et al. Glioma-associated microglial MMP9 expression is upregulated by TLR2 signaling and sensitive to minocycline. Int J Cancer 2014;135(11):2569–2578

88 Cohen AL, Anker CJ, Salzman K, Jensen RL, Shrleve DC, Colman H. A phase 1 study of repeat radiation, minocycline, and bevacizumab in patients with recurrent glioma (RAMBO). J Clin Oncol 2014;32 (15):2066–2066

89 Gabrusiewicz K, Ellert-Miklaszewska A, Lipko M, Sielska M, Frankowska M, Kaminska B. Characteristics of the alternative phenotype of microglia/macrophages and its modulation in experimental gliomas. PLoS One 2011;6(08):e23902

90 Jacobs VL, Landry RP, Liu Y, Romero-Sandoval EA, De Leo JA. Propentofylline decreases tumor growth in a rodent model of glioblastoma multiforme by a direct mechanism on microglia. Neuro-oncol 2012;14(02):119–131