

# ESGE–ESGENA guideline for quality assurance in reprocessing: Microbiological surveillance testing in endoscopy



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## Institutions

Institutions are listed at the end of the guideline.

## Bibliography

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## 1. Introduction



Microbiological surveillance is an important means for evaluating the outcome quality of reprocessing procedures and is an instrument of regular quality control in gastrointestinal endoscopy, whether endoscopic procedures are performed in hospitals, in private clinics or doctors' offices. It is an instrument for detecting and redressing weaknesses and mistakes in the reprocessing procedure and for preventing the transmission of infectious agents through endoscopy. This guideline, from the European Society of Gastrointestinal Endoscopy (ESGE) and the European Society of Gastroenterology and Endoscopy Nurses and Associates (ESGENA), addresses the necessity for microbiological surveillance in endoscopy and provides practical information about testing the quality of the microbiological out-

comes of manual and automated reprocessing procedures used in endoscopy.

It is a consensus guideline, prepared in co-operation with endoscopists, microbiologists, hygienists, endoscopy nurses, and representatives from industry.

## Aims of this ESGE–ESGENA guideline

These are:

- a) To support individual endoscopy departments in developing local standards and protocols for regular microbiological surveillance
- b) To support national societies and official bodies in developing national recommendations and quality assurance programs for hygiene and infection control in gastrointestinal endoscopy

<b>a. Inadequate reprocessing of endoscopes and accessories</b> <ul style="list-style-type: none"><li>– Inadequate cleaning (e. g. inadequate manual cleaning and brushing of endoscope channels)</li><li>– Contaminated cleaning accessories (e. g. cleaning brushes)</li><li>– Use of unsuitable or incompatible detergents and disinfectants</li><li>– Inadequate concentrations and contact time of agents</li><li>– Contaminated or time-expired solutions</li><li>– Contaminated rinsing water</li><li>– Fixed organic material (biofilm) in endoscopes, water pipes, containers, or washer-disinfectors</li><li>– Use of nonsterile accessories in invasive diagnosis and treatment (e. g. nonsterile biopsy forceps, polypectomy snares)</li><li>– Inadequate reprocessing of water bottles (e. g. no sterilization)</li><li>– Use of tap water in water bottles</li></ul>
<b>b. Inadequate transport and storage of endoscopes</b> <ul style="list-style-type: none"><li>– Insufficient drying before storage (e. g. <i>Pseudomonas</i> spp.)</li><li>– Inappropriate storage conditions</li></ul>
<b>c. Contaminated or defective washer-disinfector</b> <ul style="list-style-type: none"><li>– Contaminated pipes, containers, etc.</li><li>– Contaminated final rinsing water</li><li>– Mechanical/electronic defects of washer-disinfector</li><li>– Incorrect use of washer-disinfector (e. g. wrong connections)</li><li>– Lack of regular maintenance of washer-disinfector according to manufacturer's recommendations</li></ul>
<b>d. Design limitations and damaged endoscopes</b> <ul style="list-style-type: none"><li>– Small lumina, branched channels, not accessible to cleaning brushes</li><li>– Damage to the surfaces (internal and external) of the endoscope, providing potential for contamination</li></ul>
<b>e. Contaminated water in the endoscopy unit</b> <ul style="list-style-type: none"><li>– Contaminated main water pipes/supply</li><li>– Contaminated or inadequate water supply systems (filtration etc.)</li></ul>

**Table 1** Weaknesses and deficiencies in endoscope reprocessing (modified from reference [12])

2. Risk of infections and potential problem areas during reprocessing

Endoscopic procedures have become an essential tool in the diagnosis and treatment of gastrointestinal diseases, and every patient has the right to be examined and treated without risk of transmission of infectious agents or complications that may result from inadequate reprocessing of endoscopes and endoscopic accessories [1].

Since the late 1970s there have been sporadic reports of nosocomial infections linked to endoscopic procedures. Bacterial infections have been acquired during endoscopy, caused for example by *Salmonella* spp., *Helicobacter pylori* and *Pseudomonas* spp. [2–6]. Viral diseases such as hepatitis B and C have also been transmitted during endoscopy [7–8]. The majority of documented cases were caused by non-compliance with national and international reprocessing guidelines [2–8].

Manual cleaning including brushing is the first and most important step in reprocessing flexible endoscopes [1,9,10], regardless of whether an automated system is used. Non-cleaned or insufficiently cleaned endoscope channels promote the formation of microbial plaques and biofilms. Organic material which is not removed by manual brushing can additionally be fixed by aldehydes and promote the growth of organisms. Manual cleaning must include all accessible endoscope channels, all valve ports, the outer surface, and parts that are difficult to access such as the bridge elevator. Therefore, thorough cleaning is a prerequisite for adequate disinfection, regardless of whether the endoscope is reprocessed manually or in an automatic washer-disinfector.

In addition to the endoscopes themselves, water bottles can be a source of endoscope contamination. This can be caused by inadequate cleaning of water bottles, lack of sterilization or use of tap water instead of sterile water [11]. Therefore, testing of water bottles should be part of regular quality control.

As the design of endoscopes varies depending on the manufacturer, it is essential that staff is familiar with the design and construction of all the equipment in order to ensure safe and adequate cleaning and disinfection.

Table 1 presents a summary of areas of weakness and deficiencies with regard to endoscope reprocessing.

3. Responsibilities

A quality assurance program with regular microbiological surveillance should be established for endoscopy rooms, whether endoscopy is carried out in hospitals, private clinics, or doctors' offices.

The clinical service providers have a duty of care to provide hygiene supervision and to carry out microbiological surveillance. Furthermore, every professional also has a duty of care to comply with the relevant national recommendations and regulations.

The collection, culturing, and interpretation of test results should be performed in close co-operation with the endoscopists, endoscopy nurses, hygienists and appropriate microbiology personnel/microbiologists. All test details (including test media, method of collecting samples, culture, and interpretation) should be discussed with the relevant microbiological authorities prior to testing.

4. Frequency

Depending on the reprocessing procedure (manual or automated), routine quality assurance of the whole endoscope reprocessing system must be established. Routine testing can cover periodic microbiological surveillance of endoscopes, washer-disinfectors, accessories, and the water supply used in endoscopy.

Test area/material	Test method	Standard
Each endoscope channel	Flushing or rinsing of: – Suction/biopsy channel – Water channel – Air channel – Additional rinsing channels – Elevator channel on duodenoscopes	– Fill a sterile syringe with 20 ml sterile saline – Connect the syringe to the entry port of each channel – Ensure that the connection permits complete flushing of the whole channel – Adequate connectors should be used in order to guarantee correct rinsing – Collect the fluid in a sterile container
Outer surfaces	Swabs from e.g.: – Distal end – Valve ports – Bridge elevator	– Use sterile swabs, moistened with sterile saline with or without appropriate neutralizer – Take swabs from each separate part of the endoscope – Put each swab separately in a suitable medium and container (eg. Tryptic Soy Broth, containing neutralizer)
Water bottle	Liquid sample	– Water bottle ready for use should be tested – Sample volume: 2 × 100 ml – Adequate connectors should be used in order to take liquid samples from the water bottle via the connection tube of the water bottle – Use the appropriate and adequate connector and a sterile syringe to collect liquid samples from the water bottle
Final rinse water	Water sample	– Use a sterile syringe – Collect the water in a sterile container – Sample volume: 2 × 100 ml

**Table 2** Sampling methods for testing the quality of reprocessing of endoscopes

National guidelines and laws on hygiene and infection control in gastrointestinal endoscopy vary from country to country [9,10]. Consequently the frequency of microbiological surveillance and recommended test procedures differs across Europe (see Appendix). This guideline will therefore need to be modified locally in compliance with the appropriate national regulations. As a point of reference, the ESGE–ESGENA guideline committee recommends routine testing at intervals no longer than 3 months.

**Endoscopes.** Depending on the number of endoscopes in use, it may be impractical to test each endoscope at each occasion. Therefore a sample of each type/series should be tested at each sampling session in rotation, making sure that at the end of the year each endoscope has been sampled at least once.

**Water supply.** Microbiological surveillance of the final rinse water used in endoscopy should be carried out on the same day as the microbiological surveillance of the endoscopes, in order to assure the testing of the complete reprocessing cycle.

**Washer-disinfectors.** In the case of automated endoscope reprocessing, validation of the whole procedure is necessary to ensure that it complies with the requirements of the European Standard, prEN ISO 15883 parts 1, 4 and 5 [13]. The ESGE–ESGENA guideline for process validation and for routine testing for endoscope reprocessing in washer-disinfectors should be considered in this regard [14].

The microbiologists should record the data and make a simple trend analysis to see if any contamination detected increased or decreased between tests in order to identify any changing circumstances before a potential outbreak.

## 5. Sampling for routine tests

### 5.1. Endoscopes

Microbiological testing of endoscopes should cover:

- ▶ all channels
- ▶ the outer surfaces, and
- ▶ the connected water bottle.

A sampling plan for each endoscope type has to be established, that takes into account the critical parts of each type.

Sterile saline 0.9% is the most popular test solution. Depending on requirements, a dose of neutralizer can be added to the saline solution (or into the sterile sampling container) in order to neutralize any traces of chemicals which may limit detection of microorganisms. Reports on *in vitro* efficacy tests of the disinfectant used may serve as indicators for the choice of an appropriate neutralizer.

To avoid contamination from the environment, the collection of samples must be done under aseptic conditions.

**Method of sampling.** Standard testing of the endoscope includes collection of:

- a) liquid samples from endoscope channels,
- b) swabs from outer surfaces, and
- c) liquid samples from water bottles.

**a. Liquid samples from endoscope channels.** Flush the appropriate channels with 20 ml sterile saline and collect the liquid in a sterile container (see Table 2).

**Note:**

- ▶ Because of the complex construction of endoscope channels, each channel should be tested separately.
- ▶ Adequate connectors should be used to ensure the complete and separate flushing of each channel.
- ▶ The endoscope manufacturer should give clear instructions on how to connect and test each channel.
- ▶ Because of its small lumen, the elevator channel of duodenoscopes should be tested by flushing with 5 ml sterile saline with or without an appropriate neutralizer.

**Table 3** Indicator organisms for quality control

Organisms identified in microbiological tests	Indication of origin	Troubleshooting
<i>Escherichia coli</i> , enterococci and Enterobacteriaceae	A: Insufficient cleaning and/or disinfection procedures, e. g.: – No brushing – Inadequate concentrations or exposure times of process chemicals	A: Review whole reprocessing cycle with special emphasis on manual cleaning
	B: Mechanical or electronic defects of washer-disinfector, e. g.: – Incorrect amounts and/or concentration of processing chemicals – Design flaws of washer-disinfector, with dead volumes	B: Initiate full maintenance of washer-disinfector
<i>Pseudomonas aeruginosa</i> and other gram-negative nonfermenters	A: – Insufficient final rinsing – Contamination of final rinsing water – Contamination of washer-disinfector due to mechanical or electronic defects – Contamination of filter systems – Design flaws of washer-disinfector with dead volumes	A: Review water supply systems and procedures: – Water quality – Manual and/or washer-disinfector rinsing – Initiate full maintenance of washer-disinfector and filtration systems – Initiate autodisinfection cycle according to manufacturer's instructions (thermal disinfection is preferred)
	B: Insufficient drying of endoscopes before storage	B: Review drying procedures before storage, and ventilation of storage facilities
<i>Staphylococcus aureus</i> , <i>Staphylococcus epidermidis</i>	Recontamination of endoscopes due to: – Inadequate storage and transport – Inadequate hand hygiene	Review of hygiene arrangement for storage, transport and of manual handling
	Contamination from sampling	Repeat sampling
Atypical mycobacteria <i>Legionella</i> organisms	Contamination of washer-disinfector and water system	Review water supply systems and procedures: – Manual and/or washer-disinfector rinsing – Initiate autodisinfection cycle according to manufacturer's instruction (thermal disinfection is preferred) – Initiate full maintenance of washer-disinfector and filtration systems

**b. Swabs from the outer surfaces of the endoscope.** These swabs are taken to test the adequacy of cleaning and disinfection. The method is described in [Table 2](#).

**c. Liquid samples from water bottles.** Water samples should be taken from water bottles at the end of the defined application period (see [Table 2](#)).

## 5.2. Final rinse water of washer-disinfectors

Depending on the design of the washer-disinfector, the options for collecting samples may vary. Irrespective of the type of washer-disinfector, the complete reprocessing cycle should be tested. Manufacturers should provide advice regarding appropriate means of microbiological sampling. A sample of 2 × 100 ml should be taken from the final rinse water.

## 5.3. Water supply

Take water samples according to national recommendations for testing of tap water.

# 6. Cultures/culturing

The samples should be processed shortly after collection. If any delay is likely, the samples should be refrigerated (e. g. for transportation).

## 6.1. Liquid samples from endoscope channels

**Total microbiological count.** Take 1 ml of the sample and place on an appropriate number of plates (depending on plate size) of a

complete medium (e. g. Tryptic Soy Agar [TSA]). Incubate for 48 h at 30 °C.

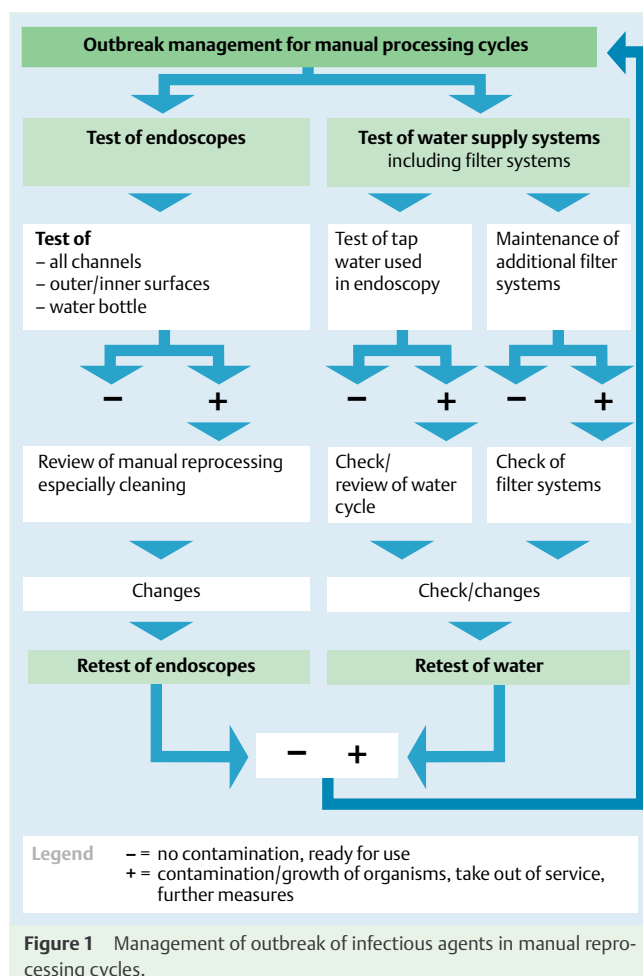
Depending on the required detection limit, another 10 ml of the same sample can be filtered (pore diameter not greater than 0.45 µm). The filter is incubated on an agar plate containing a complete medium (e. g. TSA) at 30 °C for 48 h.

### Detection of special microorganisms.

Enterobacteriaceae, *Pseudomonas aeruginosa* and staphylococci should be tested as indicator organisms. We would recommend that not only the final rinse water but also the endoscopes should be tested for atypical mycobacteria. This would ensure that the whole system is tested according to prEN ISO 15883–4. Add the same volume of double-concentrated Tryptic Soy Broth (TSB) to the rest of the sample and incubate it at 37 °C for 48 h. Double concentrated MADC broth and incubation at 37 °C for 21 days should be used if a test for mycobacteria is deemed appropriate. Streak out on selective agar plates and incubate for an appropriate time and temperature according to the manufacturer's instructions, for example, using:

- ▶ Violet Red Bile Dextrose (VRBD) agar as a selective medium for detection of Enterobacteriaceae
- ▶ Ceftrimid agar for detection of *Pseudomonas aeruginosa*
- ▶ Baird–Parker agar for detection of staphylococci
- ▶ Middlebrook agar for detection of mycobacteria

Further identification tests (e. g. commercially available biochemical test systems for bacteria or acid-fast stain for mycobacteria) may be necessary to confirm the presence of certain groups or species of bacteria.



## 6.2. Swabs

Extract the swab in 10 ml of TSB plus neutralizer, using a vortex. *Detection of special microorganisms.* Incubate the 10 ml volume at 37 °C in an incubator for 48 h. Streak out on selective agar plates and incubate for an appropriate time and temperature, according to the manufacturer's instructions (see section 6.1.)

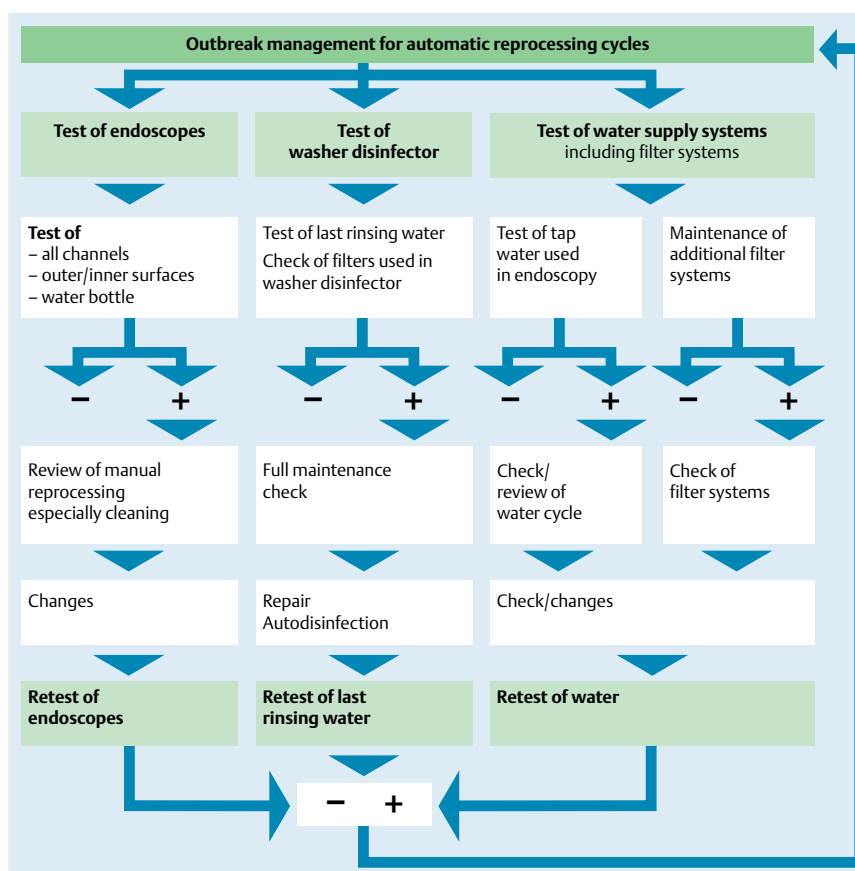
## 6.3. Water samples (from water bottle + final rinse water)

According to the European Standard, prEN ISO 15883–4 the final rinse water should be free of *Pseudomonas aeruginosa*, atypical mycobacteria and *Legionella* spp.

The aerobic total microbial count is determined by filtration (0.45 µm pore size) of 10 ml and 100 ml water samples. The sample is incubated at 30 ± 2 °C on R<sub>2</sub>A medium or another appropriate low nutrient medium for 5 days. The colonies are counted, and the type of microbe is determined by subculture on appropriate selective media and/or standard identification techniques (e.g. commercially available biochemical test systems) are used. For the detection of mycobacteria, Middlebrook 7H10 agar should be used and incubated at 37 °C for up to 21 days.

The methodology of the International Standard ISO 11731 can be used for the detection of *Legionella* spp.

For higher sensitivity in the detection of indicator organisms, an enrichment technique can be used, as follows. A 100 ml water sample is added to 100 ml double strength TSB and incubated at 37 °C for 48 h. The culture is then streaked onto selective media (see section 6.1) for identification of any grown organisms. For the detection of atypical mycobacteria, double strength MADC broth (at 37 °C for 21 days) and subculture on Middlebrook 7H10 agar (at 37 °C for 21 days) should be used.



## 6.4. Additional tests

If clinical or epidemiological data suggest the transmission of infectious agents, in addition to the routine sampling, the test methods should focus on the suspicious organism (see section 8, Management of outbreak of infectious agent).

## 7. Interpretation of results and corrective measures in case of contamination

A variety of bacteria and viruses have been associated with endoscopy-related transmission of infectious agents [2–8]. The detection of viruses is complex, time-consuming and expensive, especially that of intact, infective viruses. Therefore routine microbiological surveillance does not include viruses.

In the case of regular microbiological surveillance, it is not necessary to test for all possible bacteria. A number of organisms can be used as indicators of weaknesses or mistakes in the reprocessing procedure. An overview is given in [Table 3](#).

### 7.1. Total microbiological count

**a. Liquid samples from endoscope channels.** The maximal total count should be <20 cfu/channel. It should be calculated taking into account the amount of saline used to rinse the channel. Indicator organisms should not be found at any time.

**b. Swabs.** Cultures taken from swabs should be focused on the growth of indicator organisms (see section 7.2. and [Table 3](#)). Quantification of microorganisms is not recommended.

**c. Water samples.** The maximum total count should be <10/100 cfu/ml. Indicator organisms should not be found at any time.

### 7.2. Detection of special microorganisms

A criterion for acceptability is the **absence of growth of indicator organisms** (see [Table 3](#)).

## 8. Management of outbreak of infectious agent

In the case of manual reprocessing, the endoscopes and water used in endoscopy rooms must be tested ([Figure 1](#)).

In the case of automated reprocessing, the endoscopes, washer-disinfector, and the water used in endoscopy must be tested at the same time, in order to identify the cause of infection ([Figure 2](#)).

If any contamination is found, it is the responsibility of the clinical service provider to take the suspect piece of equipment out of service (e.g. endoscopes, washer-disinfector, accessories, etc), until corrective actions have been taken and satisfactory results have been achieved [14].

### Institutions

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## 10. Appendix

A survey was performed in July 2006 in the ESGE and ESGENA membership countries, and there were replies from 20 countries. In the majority of countries endoscopes are reprocessed in automated systems (18/20). Among the 20 countries, 13 have national guidelines on reprocessing of endoscopes and accessories, while national guidelines on microbiological surveillance are available in eight of the 20 (see [Table 4](#)). Microbiological testing varies greatly between countries in 12 of the 20. Many countries recommend testing every 3 months (see [Table 5](#)).

Country	Manual reprocessing of endoscopes	Automated reprocessing of endoscopes	National guidelines on endoscopes and accessories	National guideline on microbiological surveillance
Austria		X	X	X
Belgium	X	X	X	
Croatia		X		X
Czech Republic	X			
Denmark		X		
Finland		X	X	
France		X	X	X
Germany		X	X	X
Iceland		X		
Italy	X	X	X	
Jordan	X		X	
Luxembourg		X		
Monaco		X	X	X
Netherlands		X	X	
Norway		X		
Slovenia		X	X	X
Spain		X		
Sweden		X	X	X
Switzerland		X	X	
UK		X	X	X
Total	4	18	13	8

**Table 4** National variations in reprocessing of endoscopic equipment and provision of guidelines

Country	Interval between tests, months (except where indicated)		
	Endoscopes	Automatic washer-disinfectors	Water
Austria	12	12	NA
Croatia	3	3	3
Denmark	1	6	0
France	12	3	1
Germany	3	3	3
Italy	6	6	12
Monaco	1	1	1
Slovenia	Individual tests	Individual tests	6
Spain	3	No	No
Sweden	3	3	No
Switzerland	3	NA	NA
UK	No tests of endoscopes	According to manufacturers' recommendations	Once a week
NA, = Not announced			

**Table 5** National variations in testing intervals for routine microbiological surveillance